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4 Antibiotics, gut microbiota, environment in early life and type 1 diabetes
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40 Key words: gut microbiota, neonatal immune response, type 1 diabetes, immune
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60 Abstract
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62 The gut microbiota interact with innate immune cells and play an important role in
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64 shaping the immune system. Many factors may influence the composition of the
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66 microbiota such as mode of birth, diet, infections and medication including antibiotics.
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68 In diseases with a multifactorial etiology, like type 1 diabetes, manipulation and
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70 alterations of the microbiota in animal models has been shown to influence the
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72 incidence and onset of disease. The microbiota are an important part of the internal
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74 environment and understanding how these bacteria interact with the innate immune
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76 cells to generate immune tolerance may open up opportunities for development of new
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78 therapeutic strategies. In this review, we discuss recent findings in relation to the
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80 microbiota, particularly in the context of type 1 diabetes.
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116 *1. Gut microbiota and type 1 diabetes*
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118 Type 1 diabetes (T1D) is a T cell-mediated autoimmune metabolic disease which is
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120 commonly seen in children and young adults (1) although it can also present in older
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122 adults. The insulin-producing beta cells of the pancreatic islets are damaged and
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124 destroyed by activated autoreactive T cells resulting in disordered blood glucose
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126 regulation (2). This destruction is the result of a complex interaction between genetic
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128 susceptibility genes and environmental factors (3, 4). Genetic screening has shown that
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130 certain major histocompatibility complex (MHC) class II genes, also called human
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132 leukocyte antigen (HLA) genes, *DQA1*0301* (DQ2), *DQB1*0302* (DQ8), *DRB1*DR301*
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134 (DR3) and a number of DR4 alleles are associated with susceptibility to T1D in patients (5,
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136 6). However, only a small portion of individuals carrying those alleles will develop T1D
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138 (7). Yet, a sharp rise of T1D incidence has been seen in recent years (8) in a time frame
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140 that is not sufficient for genetic change, indicating that environmental factors may play
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142 a crucial role in diabetes development (9). Prenatal influence, viral infections, dietary
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144 factors in the young as well as “hygiene” can all affect the disease onset (10). More
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146 recently, several studies have shown commensal microbiota to be connected with the
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148 development of this autoimmune disease (11). Although triggering factors for T1D
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150 have not yet been clearly identified, the gut microbiota are believed to play an
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152 important role in the development of the disease (12, 13).

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161 The gut microbiota are associated with the development of several diseases including
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163 obesity and type 2 diabetes (14), liver disorders (15), intestinal inflammatory syndromes
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172 (16), allergic diseases (17), disorders in the central nervous system (18), and especially,
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174 autoimmune diseases (19-22). We, and others, have recently reported that alteration
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176 of gut microbiota by pharmacological means can protect from or accelerate T1D
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178 development in non-obese diabetic (NOD) mice (23-27), a well established animal model
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180 for T1D research (28).
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184 We were among the first to demonstrate that the gut microbiota shape the NOD mouse
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186 innate immune system (11). MyD88 is a central adaptor in most innate immune Toll-
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188 like receptor signaling pathways and MyD88-deficient NOD mice do not develop
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190 autoimmune diabetes in a clean, but not sterile, housing environment; however, germ-
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192 free MyD88-deficient mice develop full-blown diabetes (11). This indicates that
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194 commensal bacteria, especially gut bacteria play a very important role in triggering the
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196 autoimmune disease. When a defined microbial mixture was introduced orally into
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198 the germ-free MyD88-deficient mice, diabetes development in these mice was
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200 attenuated (11). Similar results were later observed in different mouse models of
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202 human diseases including Celiac Disease (29), obesity/type 2 diabetes (30), and
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204 autoimmune uveitis (31).
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211 There are 10-fold more microorganisms residing in the gut than the total number of
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213 human cells (32), and they protect the host from infection by various pathogens (33).
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215 The main roles of gut bacteria are to aid in nutrition derived from the diet and to
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217 generate energy. A healthy microbiota composition helps to keep the gut epithelia
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219 intact and reduce permeability (34, 35). Furthermore, the interaction between gut
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228 epithelia and the bacteria promotes the development of a normal immune system (36,
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230 37). Several reports have demonstrated that colonization by some specific bacteria in
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232 the gut can protect mice from developing type 1 diabetes; these bacteria include SFB
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234 (Segmented Filamentous Bacteria) (38), *Lactobacillus johnsonii* N6.2 (39), as well as
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236 some Streptococcal species (40), and glycoprotein extracts from *Klebsiella pneumoniae*
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238 (41).
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245 *2. Modification of the gut microbiota*

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247 Although controversial, germ-free mice (11, 42-44) may have accelerated T1D.
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249 Conversely, there has been speculation that gut bacteria may trigger T1D development
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251 in genetically susceptible humans (45) and mouse models of T1D (46). One possible
252
253 means by which this could occur could be transfer of metabolites or cell components of
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255 the bacteria through a “leaky” gut wall and uptake by antigen presenting cells,
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257 processing and presentation of the antigen to activate T cells (47). Tight junctions
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259 represent the major barrier within the paracellular pathway between intestinal
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261 epithelial cells. Alterations in intestinal permeability allow access of bacterial toxin
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263 (45), infectious agents and dietary antigens from the lumen to mucosal immune
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265 elements (48, 49). Another possible mechanism is some bacterial product(s) share the
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267 molecular homology with islet autoantigen(s) and the islet beta cells are attacked by the
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269 immune cells that are reactive to the bacterial antigens (46).
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276 Autoantibodies have been observed in T1D patients as young as several months old (50,
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284 51). Animal model studies have also shown that alteration of gut microbiota early in
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286 life, and gut permeability are important in shaping the host immune system (25, 52, 53),
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288 especially at the prenatal or neonatal stages.
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291 Efforts have been made to investigate which bacteria in the gut may be beneficial or
292
293 harmful in the development of T1D (45, 46, 54-57). Researchers have studied altered
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295 gut microbiota in experimental mice after treating with a combination of 4 antibiotics,
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297 including Ampicillin, Metronidazole, Neomycin and Vancomycin (58). Although there
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299 are studies using germ-free (GF) mice to test whether one or more species of bacteria
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301 introduced into the mice has an impact on diabetes development (42, 59), which species
302
303 are probiotic and which are detrimental have not been conclusively determined, as most
304
305 of the bacteria in the gut are non-culturable.
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309 Other studies have been conducted using vancomycin, a specific gram-positive bacterial
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311 inhibitor, to modify the gut bacteria. Antibiotic intervention during the prenatal period
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313 revealed an acceleration of diabetes onset (27, 60), whereas NOD mice receiving
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315 vancomycin from birth onwards gave the opposite result (52). Recently, Brown and
316
317 colleagues showed that using Neomycin and Vancomycin to treat NOD mouse pups from
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319 the neonatal period for their lifetime (61) accelerated diabetes development. These
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321 studies indicated that the time at which antibiotic treatment is commenced is crucial
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323 and that treating the mothers may be a way of having an effect while avoiding direct
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325 administration of the antibiotics to the pups. Many of these studies used an approach
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327 giving long-term antibiotic treatment, although long-term antibiotic treatment rarely
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340 occurs in humans. Thus, the advantage of studying short-term treatment makes the
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342 studies in animals closer to humans (25, 27). In addition, human studies have shown
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344 that approximately 30% of pregnant women in the USA have had a short-course of
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346 antibiotic medication during their pregnancy (62) and the number could be higher in
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348 other countries. It should be noted that long-term antibiotic treatment could cause
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350 resistant bacteria to propagate in the gut (63).
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357 *3. Protective bacteria that arise from pharmacological alteration of gut microbiota in*
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359 *early life.*
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362 The colonization of gut microbiota is strongly influenced by microbial exposure at birth
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364 (64). When antibiotic treatment is used to study the effect of gut microbiota on
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366 disease, it is clear that the timing of administration, duration of treatment, as well as the
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368 type of antibiotic used must be taken into account. We have published a study
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370 showing that Neomycin/Polymyxin B/Streptomycin- treated NOD mice were protected
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372 from T1D development (25). This protection was more significant when mice were
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374 treated at the prenatal stage (Figure 1, adapted from reference (25))
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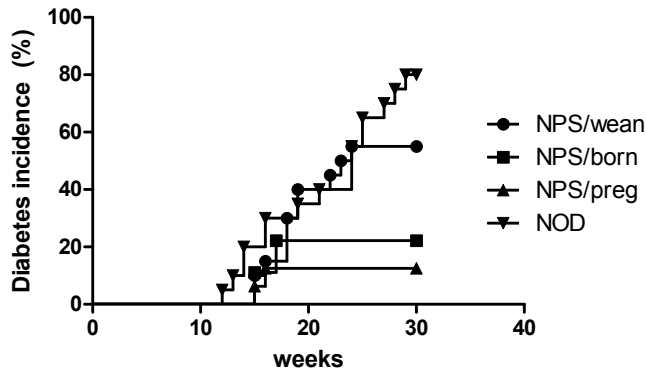


Figure 1: Maternal antibiotic treatment protects offspring from diabetes development in NOD mice. Antibiotic treatment (3 week) starting at different times of life led to a different phenotype of T1D development. NPS (Neomycin/Polymyxin B/Streptomycin); NPS/preg (NOD offspring from mothers treated with NPS during pregnancy); NPS/born (NOD treated with NPS from birth to weaning); NPS/wean (NOD treated with NPS immediately after weaning). The change in diabetes incidence was dependent on the time of antibiotic treatment.

It is clear that the earlier the NOD mice received the antibiotics, the better the protection from diabetes development. In our study, NOD mice treated with NPS at different time points early in life delayed and overall reduced T1D onset. When pregnant mothers were treated with antibiotics, the offspring were most protected from diabetes, while mice receiving antibiotics from birth or weaning were also protected from disease development although this was not statistically significant. Here, not only did the NPS treatment generate a gut bacterial composition that was protective but it was also clear that the timing of treatment was very important in

449 inducing the effects. Overall, these studies suggest that gut microbiota in very early
450 life (prenatal or neonatal) may have the most positive impact on the host immune
451 system. Other studies have demonstrated that microbial exposure during early life is
452 important for development and maintenance of the immune system and germ-free mice
453 are more susceptible to developing T1D (42, 43).

454 Considering which bacteria have protective effects in relation to diabetes development,
455 we have observed that the Gram-positive Firmucutes *Lachnospiraceae* and
456 *Coriobacteriaceae* significantly increased in prenatally NPS treated mice (25). Several
457 studies in mice and human case reports indicated that, in both BioBreeding Diabetes-
458 Prone rats (65-67) and diabetic children (68, 69), decreased *Bacteroidetes*, together with
459 an increase in other Gram-positive Firmicutes such as *Lactobacillus*, *Bifidobacterium*,
460 were found, compared to BioBreeding Diabetes-Resistant rats and healthy children,
461 respectively.

462 SFB (Segmented Filamentous Bacteria) are a group of bacteria within the Genus
463 *Candidatus Arthromitus* which belongs to the Phylum of Firmicutes, Class of *Clostridia*
464 and Family of *Lachnospiraceae*(70). SFB were found to induce intestinal Th17 cells in
465 Lamina Propria (LP) (71) and there are reports showing that colonization of the gut of
466 NOD mice with SFB can induce a substantial population of Th17 cells in the LP and
467 protect the female NOD mice from diabetes development (38). However, we did not
468 find that SFB conferred diabetes protection in NOD mice (27). A study by Yurkovetskiy
469 and colleagues also showed that SFB did not protect GF NOD mice from diabetes
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508 development; however, SFB reduced T1D development in male GF NOD mice after
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510 colonization with other gut bacteria (72). Since SFB are a group of bacteria, genomic
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512 sequencing results from several SFB strains have shown that they are different from
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514 each other (73-76). It is, therefore, conceivable that different strains may have
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516 different biological effects. Nevertheless, it is still important to study the role of SFB in
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518 mice because these gram-positive, anaerobic, commensal bacteria are capable of
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520 inducing the postnatal maturation of homeostatic innate and adaptive immune
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522 responses in the gut. A recent publication also showed that not only can colonization
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524 with SFB induce IL-17A but CXCR2-dependent recruitment of neutrophils in the gut also
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526 occurs (77).
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534 *4. Shaping the immune system via alterations in gut bacteria*

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536 How does alteration of the gut bacteria lead to protection from diseases like type 1
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538 diabetes? There are a number of proposed mechanisms which are associated with
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540 genetics, gut microbiota (related to antibiotic usage, mode of delivery, diet) and
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542 infection in animal models and humans (78, 79). Our recent studies indicate that
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544 protection can be mediated by tolerogenic antigen presenting cells (APCs) originating in
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546 the gut associated lymphoid tissue (GALT), which have reduced ability to stimulate
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548 cytotoxic CD8⁺ T cells. (Figure 2, Cover figure)
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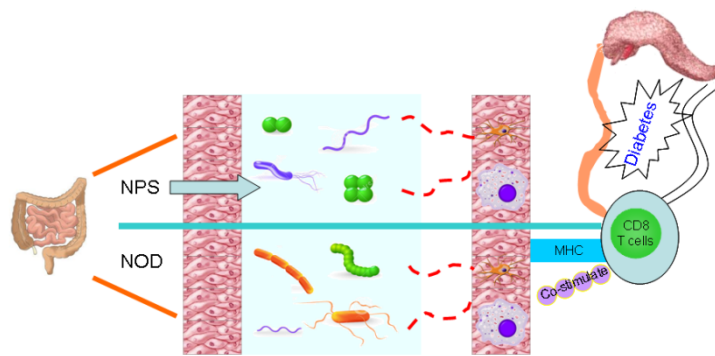


Figure 2: How do antibiotics affect pancreatic beta cell autoimmunity: gut microbiota composition is altered after antibiotic (NPS) treatment and antigen presenting cells exposed to the altered bacteria in the gut display impaired antigen presenting ability to CD8 T cells, which in turn alleviate insulinitis in the pancreas and protect the host from diabetes development.

In supporting the mechanism of tolerogenic APCs as a result of altered gut microbiota due to antibiotic usage, Umenai and coauthors showed hyporesponsiveness of macrophages, a potent subset of APCs, in response to LPS stimulation in mice after Streptomycin treatment (80). Similarly, a recent study demonstrated that dendritic cells became hyporesponsive to LPS stimulation and reduced inflammatory cytokine production upon exposure to the gut bacterium *Lactobacillus reuteri* (81). Dolpady and co-authors reported that administration of a mixture of *Bifidobacteriaceae*, *Lactobacillaceae* and *Streptococcus* in 4-wk old mice promoted tolerogenic CD103⁺ dendritic cells and reduced Th1 and Th17 cells in mucosal and PLN sites (82). Thus, alteration of gut microbiota by different means including antibiotic treatment can

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620 affect APCs lead to acceleration of or protection from diabetes development. The
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622 changes in APC function could be the result of direct contact between APCs and gut
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624 microbiota in the gut or indirect contact, mediated by metabolites from altered gut
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626 microbiota. Due to the immature nature of gut barrier and the changing, maturing
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628 community of gut microbiota in early-life, timing becomes critical.
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634 Reduction in regulatory T cell markers have also been postulated as a mechanism for
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636 protection of NOD mice after treating with a mixture of antibiotics – metronidazole,
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638 streptomycin and polymyxin prenatally (53, 60). These studies demonstrated that the
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640 changes in the immune system occurred when antibiotic treatment was given prenatally,
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642 a particularly important time for immune system development. Livanos and coauthors
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644 reported recently that NOD mice receiving antibiotics (penicillin V) from lactation until
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646 the age of 40 days had earlier diabetes onset and overall higher incidence of diabetes
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648 (83). This early-life treatment reduced the percentage of Treg and Th17 cells in lamina
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650 propria (LP), which may have contributed to the accelerated diabetes development. In
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652 the Streptozotocin (STZ)-induced type 1 diabetes model, mice treated with antibiotics
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654 were fully protected from diabetes (84). This was attributed to blocking pro-diabetic
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656 bacteria translocation to PLN (84).
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666 Early-life treatment using vancomycin reduced the incidence of diabetes in one study
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668 (52) but accelerated diabetes in another (27). The discrepancy may be attributed to
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676 different treatment protocols. However, the protection in Hansen's study was
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678 accompanied by an increase in the level of *Akkermansia*, which was later reported to be
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680 correlated with a pro-diabetic effect (85, 86). A gluten-free diet can also reduce
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682 inflammation and diabetes incidence in NOD, with elevated abundance of *Akkermansia*.
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684 Adding gluten to the gluten-free diet reversed the protection, accompanied by a
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686 decreased level of *Akkermansia* (86). More recently, a study showed that *Akkermansia*
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688 mediate glucose tolerance via IFN γ (87) using loss-and-gain-of-*Akkermansia muciniphila*
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690 approaches in IFN γ knock-out mice. In a clinical study, a 4-day treatment with broad-
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692 spectrum antibiotics (Vancomycin, Gentamicin and Meropenem) significantly shifted the
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694 gut microbiota composition but this did not have a clinical impact in respect of
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696 metabolic markers such as glucose tolerance or insulin secretion (88). However, in a
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698 study by Endesfelder and coauthors a change of gut microbiota occurring as a result of
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700 dietary change had an impact on islet autoimmunity. The authors stratified the
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702 children in the study based on the microbial communities identified. They found that it
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704 was possible to detect functional associations between the diet consumed, the
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706 microbiome and development of autoimmunity. They identified a subgroup of
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708 children where *Bacteroides* was dominant, with low *Akkermansia* in the gut microbiota
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710 and this was associated with early introduction of a non-milk diet, lower abundance of
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712 genes for the production of butyrate and early autoantibody development (89, 90).
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714 They postulated that low butyrate generation by the bacteria contributed to increased
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716 risk of the development of islet autoantibodies (90).
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732 It is clear that the effect of gut microbiota on host immune responses early in life is
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734 much stronger than later in life. This was observed in human studies demonstrating
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736 that the development of type 1 diabetes was closely related to gut microbiota, gut
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738 permeability and immune system in early-life. Amarri and colleagues have previously
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740 found that gut bacteria and gut permeability as well as other immune markers were
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742 significantly altered in breast-fed infants (89). Other early influences have been
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744 demonstrated in a Danish study where the authors found that antibiotics used in early-
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746 life increased the incidence of type 1 diabetes in young children: however, this was also
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748 related to the mode of birth delivery, which has major effects on determining the
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750 composition of gut microbiota at the time of birth (91).
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758 Antibiotic treatment in adult mice is not as effective as prenatal or neonatal treatment
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760 in altering the course of type 1 diabetes. However, later life antibiotic treatment can
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762 significantly alter the gut bacteria and it can have clear impact on diseases other than
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764 T1D, including Crohn's disease (92), colitis (93), obesity and type 2 diabetes (94).
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770 In addition to antibiotics, it is known that innate immune system can also alter gut
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772 microbiota, which affect both type 1 and type 2 diabetes development (11, 30).
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774 However, it was not clear which type of bacteria contribute to the protection or
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776 promotion of diabetes. Using a MyD88-deficient NOD mouse that has a defined T cell
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778 receptor repertoire, we recently found that one type of bacterium, *Leptotrichia*
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788 *goodfellowii*, in the gut can trigger diabetes development in islet-specific glucose-6-
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790 phosphatase catalytic subunit-related protein (IGRP)-reactive CD8 T cell receptor NY8.3
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792 transgenic NOD model. *L. goodfellowii* is a member of *Fusobacteria*, and expresses a
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794 protein peptide sharing a homologous sequence with IGRP and its abundance is
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796 correlated with the progression of diabetes (46). This is the first evidence of molecular
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798 mimicry that relates to a bacterial antigen in initiation of diabetes onset, due to T cell
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800 cross-reactivity to the microbial peptide and leading to an activated autoimmune
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802 response.
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809 810 *5. Fecal microbiota transfer (FMT) as a potential therapy*

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812 Gut bacterial composition may be altered by diet, antibiotics, probiotics, or by direct
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814 microbiota transfer. Fecal microbiota transfer (FMT) has proven to be a effective way
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816 to transfer “healthy” microbiota, that may have beneficial effects on metabolism and
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818 the immune system in recipient mice (30, 95). FMT has also been used in patients with
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820 colitis (96-98) and recently in cancer patients (99, 100). We recently demonstrated
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822 that FMT could restore the gut microbiota and rebalance the gut hemeostasis, which in
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824 turns delayed diabetes onset in NOD mice (26, 29). In other disease models, including
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826 obesity, the gut microbiota have been shown to contribute to generation of the
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828 metabolic syndrome (30). FMT from lean mice to obese mice can ameliorate the
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830 metabolic syndrome in the obese mice due to the rebalancing of gut microbiota (101).
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836 Treating metabolic syndrome with FMT has also been tested in humans [98].
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844 The beneficial effects of FMT have been particularly used in patients suffering from
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846 *Clostridium difficile* infection (102, 103). Although antibiotic treatment has been the
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848
849 mainstay of treatment of *Clostridium difficile* infection, some patients have developed
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851 recurrent infection later on and importantly antibiotic treatment for other medical
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853 conditions can lead to persistent infection with *Clostridium difficile* (104-106). Since
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855 the clinical trial on FMT in patients with recurrent *Clostridium difficile* infection (103),
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857 considerable progress has been made in this field, including formulation of the
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859 microbiota and delivery route of the bacteria. Oral administration is very common
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861 using capsules containing the gut microbiota of healthy donors after pathogen-free
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863 screening, stool purification and preservation processes (107). In addition, FMT has
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865 also been effective in treating diseases like ulcerative colitis (108) and chronic pouchitis
866
867 (109) in clinical practice. Although FMT has not been used in T1D treatment, if specific
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869 combinations of beneficial bacteria could be identified, this could be a promising
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871 therapy, given that fecal filtrate has a similar efficacy to fecal microbiota in treating
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873 other medical conditions (110). However, more comprehensive studies need to be
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875 performed to understand which what bacterium (or bacteria) and metabolites can
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877 trigger the imbalance of immune system at early-life in humans at high-risk of
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879 developing T1D.
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889 6. Perspectives and conclusion

890 It is clear that many factors can affect the composition of gut microbiota, which
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900 modulate the immune system contributing to health or disease including T1D. Birth
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902 delivery mode (cesarean section vs. vaginal delivery), breast vs. formulation feeding and
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904 antibiotic usage, all can have a strong impact on the health of newborn infants (111-
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906 113). The period in utero, as well as post-partum, is a crucial time for establishment of
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908 the immune system of the babies. Intervention during early life may lead to novel and
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910 more effective therapeutic strategies in treating T1D. There is no doubt that gut
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912 microbiota are associated with particular metabolites and can influence both metabolic
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914 pathways and the development of the immune system. Many studies have shown that
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916 depletion or alteration of intestinal microbiota has a significant impact on gut mucosal
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918 and epithelial gene expression, as well as immune responses (114). Gram-negative and
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920 Gram-positive bacteria can stimulate different immune responses and induce Th1 or Th2
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922 type cytokines, as well as proinflammatory or anti-inflammatory mediators (115-118).

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929 **We summarized some of the bacteria studied in association with T1D (Table 1).**
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933 APC dysfunction has been found to be associated with T1D development (119, 120), but
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935 previously this had not been correlated with alteration of gut bacteria. Our recent
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937 studies provide evidence that tolerogenic APC are generated as the results of altered gut
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939 microbiota (25, 27). Interestingly, the tolerogenic APCs can confer T1D protection and
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941 this protection can also be transferred to a second host and to the offspring.
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945 Probiotics and other strategies altering the gut microbiota, including FMT, could be a
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947 promising approach to modulate gut microbiota and rebalance the homeostasis of
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953 mucosal and systemic immune systems. This “bug for drug” approach, also called
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956 “Bacteriotherapy” is being tested in different clinical trials for other medical conditions
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959 (121). The prospect of this approach for T1D could be on the horizon. Type 1
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962 diabetes has not only a genetically inherited component that determines susceptibility
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965 to disease, but the interaction with the environment to precipitate disease is a very
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968 important part of the pathogenesis. Identifying modifiable environmental factors, such
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971 as the gut microbiota would provide a therapeutic target that could be modified by
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974 treatments that could be easy to administer. It would be important to identify safe
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977 means of doing this that could potentially be administered very early in life. Probiotics
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980 have already been tested in infants and young children but the appropriate composition
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983 should be identified. Whether gut bacteria are involved in the initiation of the process
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986 leading to T1D or in the progression of β -cell autoimmunity is still unclear (122). A
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989 pure, culturable bacterial cocktail would need to be identified before this kind of
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992 regimen could be used in clinical practice.

992 A number of immunotherapeutic treatments have been trialed in human type 1
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995 diabetes, some of which have had a transient effect, but none as yet have been long-
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998 lasting (123). It has been argued that for a treatment to be successful, the innate
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1001 immune system should present antigens in a tolerogenic manner to T regulatory cells
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1004 (123) and this is potentially one of the ways in which altering the gut microbiota could
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1007 work if the right combination could be identified. Ideally, this type of treatment would
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1012 be combined with others that could be synergistic in maintaining a tolerogenic
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1014 environment.
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1019 In conclusion, both probiotic and antibiotic treatment can significantly alter the gut
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1021 bacteria, as well as the bacteria composition in other sites of the body including the oral
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1023 cavity. The altered bacteria interact with the host immune system and reduce
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1025 inflammatory cytokines as well as the expression of costimulatory molecules in APCs,
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1027 inducing a tolerogenic environment. However, further investigation is required and, in
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1029 particular, more work done to identify less disease- promoting gut microbiota and how
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1031 to maintain this type of gut microbiome.
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Table 1: Summary of bacteria studied in association with T1D

Bacteria	Gram-positive or -negative	Mode of action	Protection from or acceleration of T1D	References
SFB (Segmented filamentous bacteria), <i>Candidatus Arthromitus</i>	positive	Induce Th17 response	Protection No effect	38 27
<i>Lactobacillus johnsonii</i> & <i>L. reuteri</i>	positive	Healthy probiotics	Protection	39, 81
<i>Klebsiella pneumoniae</i>	negative	Pathogen causing pneumonia	Protection	41
<i>Bifidobacterium</i>	positive	Often as healthy probiotics	Acceleration	67-69
<i>Streptococcus</i>	positive	Some pathogens, some commensal bacteria in mouth, skin, intestine and upper respiratory tract	Protection	40, 82
<i>Akkermansia muciniphila</i>	negative	Mucin-degrading, often anti-inflammatory effect	Protection	52, 90
<i>Bacteroides</i>	negative	processing complex molecules to simpler compounds in the host intestine	Acceleration	89, 90
<i>Leptotrichia goodfellowii</i>	negative	Oral commensal, pathogen in immune compromised patients	Acceleration in NY8.3 NOD	46