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Involvement of Hedgehog Pathway in Early Onset, Aggressive Molecular Subtypes and Metastatic Potential of Breast Cancer

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Abstract

Purpose: Dysregulation of hedgehog pathway is observed in numerous cancers. Relevance of hedgehog pathway genes in cancer cohort and inhibition of its downstream effector (GLI1) towards metastasis in cell lines are explored in the study.

Method: 150 fresh tumours of breast cancer patients were collected for the study. Based on differential expression, panel of 6 key regulators of the pathway (SHH, DHH, IHH, PTCH1, SMO and GLI1) in microarray datasets were identified. Expressional profiles of aforementioned genes were later correlated with clinico-pathological parameters in Pakistani breast cancer cohort at transcript and protein levels. In addition, GLI1 over expressing breast cancer cell lines (MDA-MB-231 and MCF-7) were treated with GANT61 to explore its probable effects on metastasis.

Result: SHH, DHH, PTCH1 and GLI1 were significantly over-expressed in tumours as compared with respective normal mammary tissues. A significant correlation of SHH, DHH and GLI1 expression with advanced tumour size, stages, grades, nodal involvement and distant metastasis was observed ($p < 0.05$). Over-expression of SHH, DHH and GLI1 was significantly related with patients having early onset and pre-menopausal status. Of note, hedgehog pathway was frequently up regulated in luminal B and triple breast cancer negative subtypes affected women. In addition, positive correlations were observed among aforementioned members of pathway and Ki67 emphasizing their role towards disease progression. Exposure of GANT61 (inhibitor for GLI1) significantly restricted cell proliferation, reduced cell motility and invasion.

Conclusion: Role of activated hedgehog pathway in breast cancer metastasis provides a novel target for cancer therapy against aggressive cancer subtypes.

Introduction

Breast cancer is among the most common malignancies accounting for 25% of total cancer burden and 15% of cancer related deaths [1]. Recently, reactivation of early developmental pathways in promoting solid tumour growth has been observed [2]. Pivotal role of hedgehog pathway in embryonic patterning and later mammary gland development has been reported [3]. Briefly, binding of three ligands Sonic hedgehog (SHH), Indian hedgehog (IHH) and Desert hedgehog (DHH) with Patched1 (PTCH1) results in release of Smoothed (SMO). Once relieved SMO activates GLI ((glioma-associated oncogene homolog 1, transcription factor) responsible for activation of several downstream growth effectors [3].

Numerous studies suggested dysregulation of hedgehog pathway has an important impact on carcinogenesis [4–6]. Exploring any probable correlation of hedgehog pathway with clinico-pathological parameters is a step forward towards development of prognostic and predictive markers for future therapeutics. Earlier, SHH and GLI1 over-expression have been associated with poor prognosis in different types of cancers including breast [6,7], colon [8], glioma [9] and prostate [10]. However, prognostic significance of hedgehog pathway with early disease onset and different molecular subtypes of breast cancer remain contradictory. Furthermore, its contribution towards metastasis by using cancer cell lines is yet to be established. Aim of the study was initially to observe hedgehog pathway activation both at mRNA and protein levels in the cohort. Based on these findings, significance of hedgehog pathway in breast cancer metastasis was explored using GANT61 (GLI antagonist). Blockade of GLI1 via GANT61 will be helpful to provide novel insight for involvement of hedgehog pathway in disease progression.

Results

Clinical and molecular characterization of cohort

In Oncomine, 2 datasets TCGA Breast and Finak Breast were assessed for expression variation of hedgehog pathway molecules in deposited data. SHH was significantly over-expressed in TCGA

(F.C=1.5, $p<0.05$) and Finak dataset (F.C=1.2, $p<0.0001$) as compared to normal tissues. Expression of PTCH1 (F.C=2.3, $p<0.0001$) and GLI1 (F.C=2.2, $p<0.0001$) was also significantly elevated in Finak dataset (Suppl. Fig.1). Elevated expression of DHH, PTCH1, and GLI1 in the data deposited for TCGA provisional dataset in cBioPortal was also observed.

Demographic data of present study cohort comprised of 52% of patients having early disease onset and were ≤ 45 yrs of age at the time of diagnosis. Around 60% of cohort represented moderately differentiated tumours. Two third of total cases were in advance stages of breast carcinogenesis. Distribution of these details along with clinical findings and assessment of hedgehog pathway are presented in the table 1. For transcriptional analysis, fold change was evaluated relative to normal control tissues whose mean was calculated as 1.

Significant over expression of SHH (R.E=4.4 \pm 3.32), DHH (R.E=4.7 \pm 3.46), PTCH1 (R.E=3.4 \pm 3.08) and GLI1 (R.E=3.8 \pm 2.49) were detected in the breast tumour biopsies as compared with controls. In contrast, IHH (R.E=0.04 \pm 0.16) expression was significantly reduced in cancerous tissues. Lack of any significant association of SMO expression among tumour and controls were observed. PTCH1, SHH, DHH, and GLI1 over-expression were observed in 74%, 95%, 92% and 98% tumour specimens of the cohort. Expression patterns of these genes at mRNA and protein levels along with heatmap of qPCR data are shown in figure 1.

Correlation of Hedgehog Pathway with patients having early disease onset and pre-menopausal status

In both datasets of Oncomine (TCGA breast and Finak breast) expression of SHH, DHH and GLI1 was higher in patients having early disease onset. Expression of SHH and GLI1 was also elevated in patients having pre-menopausal status while DHH was higher in post-menopausal cases. In the current study, cohort elevated expression of SHH (R.E=5.6 \pm 3.9, $p<0.05$), DHH (R.E=5.5 \pm 4.1, $p<0.05$), SMO

(R.E=1.9±2.2, p<0.05) and GLI1 (R.E=6.3±4.5, p<0.05) correlated significantly with patients having early disease onset. Similarly, expression of SHH (R.E=6.8±4.8, p<0.05), DHH (R.E=5.5±3.9, p<0.05) and GLI1 (R.E=7.9±4.4, p<0.01) was significantly elevated in pre-menopausal patients while PTCH1 (R.E=3.9±3.3, p<0.01) was higher in post-menopausal (Fig. 2).

Correlation of Hedgehog Pathway with markers of poor prognosis

Over expression of SHH, DHH and GLI1 were associated with cell grades in Finak dataset and also related with poorly differentiated tumours. Similarly, transcript copy numbers of SHH and DHH were also found related with advanced stages (III, IV). Elevated transcriptional profiles of SHH, DHH and GLI1 were associated with almost all indicators of poor prognosis in this cohort. Expression of SHH (p<0.05) was 9 folds higher and GLI1 (p<0.05) was 4 folds higher in patients presented with poorly differentiated histological grade. Mean mRNA values of SHH, DHH and GLI1 showed gradual increase with tumour size, nodal spread and distant metastasis. Significant correlation was observed between SHH (R.E=12.76±5.12, p<0.0001), DHH (R.E=9.8±5.1, p<0.05) and GLI1 (R.E=12.3±3.9, p<0.0001) and invasive tumour size (Fig. 2). Expression variation of hedgehog pathway and its association with clinico-pathological parameters is illustrated in table 1. SHH, DHH and GLI1 were also evaluated at protein level using IHC. Microscopic features demonstrated cytoplasmic pattern of ligands and nuclear localization of GLI1. Tumour sections showed 54%, 65% and 75% positive staining against DHH, SHH, and GLI1 also mentioned in the figure 1. High intensity of protein expression was observed in patients with advanced pTNM stages and poorly differentiated nuclear grade. Hence a consistency of transcriptional profiling with protein profiling of SHH, DHH and GLI1 were also established. High expression of all pathway components especially GLI1 (HR=1.79 (1.25-2.56), p<0.05) demonstrated significant association with shorter DMFS (distant metastasis free survival) in grade III patients (suppl. Fig. 2). This information was coherent with expression profile of our dataset rendering hedgehog pathway as a marker of poor prognosis.

Correlation of Hedgehog Pathway with metastasis and nodal involvement

Elevated expression of SHH and DHH correlated with N1+ nodal involvement in both datasets. Increase in mean copy numbers of SHH and DHH were associated with shorter overall survival in TCGA dataset. Similarly increase in mean copy number of SHH (R.E=8.2±3.7, p<0.0001), DHH (R.E=6.5±4.4, p<0.05) and GLI1 (R.E=6.6±3.9, p<0.05) were associated with presence of lymph node involvement in Pakistani cohort. Similarly, patients having distant metastasis significantly demonstrated enhanced SHH, DHH and GLI1 expression with highest levels of DHH (R.E=17.1±2.2, p<0.01) shown in the figure 2.

Association of hedgehog pathway with ER, PR, Ki67 and HER2

In Pakistani cohort, all hedgehog molecules were strongly related to hormonal receptors (ER and PR) (r-value ranging from 0.71 to 0.87, p<0.05). However, majority of hedgehog molecules did not show any significant correlation with HER2 as shown in supplementary table 2. A strong positive correlation was established between Ki-67 proliferation index and SHH, DHH, PTCH1 and GLI1 (r-value ranging from 0.60 to 0.78, p<0.05) indicating their putative role in cell proliferation.

Correlation of Hedgehog Pathway with Molecular Subtypes of Breast Cancer

In TCGA dataset, triple negative subtype of patients showed significant over expression of DHH, PTCH1 and SMO. Following St. Gallen International Expert Consensus System, Pakistani cohort retains highest proportion of luminal-B (53%), followed by triple negative (18%), luminal-A(15%) and HER2 (14%) individuals. Both luminal B and triple negative subtypes representing patients showed significantly increase of SHH, DHH, PTCH1 and GLI1 as shown in the fig 3. Furthermore, prognostic role of hedgehog pathway was assessed using the online available Kaplan Meier Plotter dataset (n=626 patients, follow-up threshold=15yrs) [17]. Elevated expression of hedgehog pathway members was also associated with shorter distance metastasis free survival in patients diagnosed with luminal-B breast cancer patients as shown in the supplementary figure 3.

High expression of SHH, DHH and GLI1 is poor predictor of overall survival

Due to prospective nature of study design, follow-up data for overall survival was collected for 36 to 40 months. Kaplan-Meier plots were generated using high and low expression of SHH, DHH and GLI1 (Fig. 3). Log-rank test was used to assess the effect of expression of SHH (HR=2.97 (1.6-5.6), p=0.0007), DHH (HR=2.25 (1.2-4.4), p=0.02) and GLI1 (HR= 4.06(2.2-7.5), p< 0.0001) which were all associated with poor predicted mortality. Based on univariate and multivariate Cox's proportional regression analyses, elevated expression of SHH (HR=3.94 (1.7-9.4), p=0.002), DHH (HR=2.72 (1.2-6.5), p=0.02) and GLI1 (HR=4.5 (2.2-9.4), p<0.0001) were found to be independent predictor of overall survival. Furthermore patients having ER positive tumours (HR=3.05 (1.36-6.89), p=0.007) and distant metastasis (HR=3.47 (1.2-9.7), p=0.02) were found to be associated with shorter overall survival. Age (HR=0.18 (0.07-0.44), p=0.0003) was also found to be a significant predictor of mortality having association of older age with less hazard ratio and good prognosis (table 2). Overall high expression of SHH, DHH and GLI1 was found to be an independent predictor of poor mortality in the present cohort.

GANT61 mediated metastasis restriction by decreasing cell proliferation

Earlier, effect of GANT61 in suppressing hedgehog pathway was assessed by measuring SHH, PTCH1 and GLI1 expressions using qPCR and western blot. GANT61 effectively inhibited GLI1 (R.E=0.2±0.1 in MDA-MB-231 and R.E=0.2±0.17 in MCF-7, p<0.05). Being targets of GLI1, expression of SHH (R.E=0.18±0.2 in MDA-MB-231 and R.E=0.3±0.4 in MCF-7, p<0.05) and PTCH1 (R.E=0.09±0.14 in MDA-MB-231 and R.E=0.3±0.4 in MCF-7, p<0.05) were also repressed after treatment with GANT61 as shown in the figure 4. GANT61 significantly inhibited proliferation of MDA-MB-231 and MCF-7 cells in a dose and time-dependent manner from their respective control groups (p<0.0001) as mentioned in figure 5. IC50 calculated for GANT61 (10µM) was used for migration and invasion assays.

GANT61 reduced metastatic potential by decreasing cell migration and invasion

Furthermore, effect of hedgehog signaling on motility of breast cancer cells using an *in vitro* wound-healing assay. MDA-MB-231 and MCF-7 cells were treated with pre-conditional medium containing GANT61 (10 μ M) and control medium containing DMSO. GANT61 inhibited cell migration in both cell lines in contrast to their respective control cell lines ($p < 0.0001$) as shown in the fig.5. Moreover, the effect of hedgehog signaling on the invasive ability of breast cancer cells was also evaluated using matrigel invasion assay. The ability of breast cancer cells to invade matrigel was markedly reduced in cells that were treated with GANT61 ($p < 0.0001$), suggesting that hedgehog signaling has an essential role in the migration and invasiveness of breast cancer cells.

Discussion

Expression pattern of all aforementioned genes of hedgehog pathway are in line with earlier studies in different cancers [18–21]. Interestingly, lack of IHH association with clinical parameters is also observed in breast and prostate cancers [22,23] in line with the given cohort. However, up regulation of IHH observed among Chinese breast cancer cohort is contradictory to our findings [24]. This may be attributed to population specific genotypic variations, multiple hedgehog ligands targeting PTCH1 and sensitivity of detection technique.

Over-expression of SHH, DHH and GLI1 was frequently observed in triple negative and luminal-B subtypes of the given cohort. In triple negative patients, activated hedgehog pathway was significantly related with advanced tumour stages and poor prognosis as has already been reported [25,26]. Hence suppression of hedgehog pathway may provide a novel therapeutic strategy for treating TNBC. On the other hand, association of hedgehog pathway with luminal subtypes can also be explained based on positive correlation of different hedgehog molecules with ER in the cohort. Co-expression of ER and SHH was present among 60% of the cohort patient's, indicating molecular crosstalk as mentioned previously [27]. Similarly, strong positive correlation between hedgehog members (DHH, SHH, GLI1) and high Ki-67 index with values 75, 80 and 85% respectively was observed. These findings corroborate

interplay of hedgehog pathway in tumour growth of luminal B subtype patients. Earlier it has been shown that inhibition of GLI1 expression via RNAi mediated depletion restricted cell growth in ER positive tamoxifen resistant cells [28]. Furthermore, role of hedgehog pathway towards tumourigenesis in endocrine therapy resistant patients has also been established [29].Based on these findings, inclusion of hedgehog pathway as novel therapeutic target in treating patients belonging to both TNBC and luminal-B subtypes is strongly recommended.

Generally, aggressive molecular subtypes noted among young women are associated with worse outcome [30]. Hence, identification of alternate predictive and therapeutic biomarker is a necessity for these young patients. In the given study, alteration of hedgehog pathway was frequently observed among young Pakistani patients. Keeping in view increasing incidence of breast cancer in Pakistani women, devising novel therapeutic strategies are an absolute requirement. In the given cohort, patients<45yrs showed frequent over expression of SHH, DHH and GLI1 contributing towards aggressive tumourigenesis. Interestingly earlier findings also indicate that pre-menopausal patients showed more aggressive tumour progression in comparison to post menopausal population [31]. Hence elevated expression of SHH, DHH and GLI1 was also observed in pre-menopausal patients signifying their association with poor prognosis. Thus it is necessary to adapt a molecular screening driven approach to refine treatment for younger breast cancer patients including hedgehog inhibitors in adjuvant therapy regimen.

Over-expression of SHH, DHH and GLI1 was significantly correlated with advanced stages and tumour grades of the cohort. Activation of hedgehog pathway in advance cancer stages has been reported previously in breast [25,32], ovarian [18], renal [19] and prostate cancer [10].Concomitantly, dysregulation of SHH, DHH and GLI-1 in the cohort has also been related with nodal involvement and metastasis. In another study, significant relevance of hedgehog ligands with metastasis and breast cancer specific death has been reported [33]. Similar results were obtained at both mRNA and protein levels in

Taiwanese and German populations [6,34]. These findings strongly suggest role of hedgehog pathway in breast cancer progression and metastatic spread.

High expression of SHH, DHH and GLI1 was also found to be an independent predictor of overall survival in this cohort along with age, distant metastasis and ER expression. However, SMO has a limited clinical relevance as observed in this cohort. No significant association of SMO with patient's age, metastasis or other relevant parameters has been established in this study. Although, numerous SMO inhibitors like GDC-0449 (Vismodegib) have entered clinical trials for triple negative breast cancer patients but their efficacy still remains uncertain. Benvenuto *et al.* have recently proven that targeting hedgehog pathway using antagonists like GANT61 that act downstream of SMO is a more efficient strategy than using antagonists against SMO in breast cancer [35]. Furthermore non-canonical transcriptional activation of hedgehog pathway has also been extensively documented [36]. Hence, role of GANT61 as potential inhibitor of GLI1 against metastatic cascade has been deciphered. In this study, IC50 determined for both MCF-7 (ER+ve) and MDA-MB-231 (ER-ve) cell lines, was 10 μ M. It showed maximum reduction of cell proliferation after 48hrs. This concentration of GANT61 was enough to suppress expression of GLI1 and transcriptional activation of PTCH1 and SHH as mentioned earlier [35]. Furthermore, GANT61 significantly reduced both motility and invasion thus efficiently demonstrating its potential to restrict cancer metastasis.

Expression analysis of hedgehog pathway at early stage is highly recommended especially for young patients. Furthermore, this is the first ever report regarding evaluation of GANT61 potential role on metastasis suppression. Clinical and *in-vitro* functional assays findings strongly emphasize on hedgehog pathway's putative role as potential cancer therapeutic target in breast cancer patients.

Materials and Methods

Publicly available expression data assessment

Pre-evaluation of hedgehog pathway in breast cancer datasets of two publicly available repositories namely, Oncomine and cBioportal were performed. Oncomine (Compendia Bioscience, Ann Arbor, MI; www.oncomine.org) is an online database consisting of earlier published 715 microarray datasets. RNA expression status of SHH, DHH, IHH, PTCH1, SMO and GLI1 genes were evaluated in 593 TCGA breast cancer samples and 59 Finak breast cancer samples available in Oncomine database. In order to score gene as positive (high expression) or negative (low expression), the Log2 Median-Centered ratio, as reported in the Oncomine database, was used to evaluate differential expression of these genes in different clinical groups.

Similarly, cBioportal for Cancer Genomics (www.cbioportal.org) an online publicly available database, was screened in extended TCGA dataset of 1105 breast cancer patients[11]. RNAseq data of TCGA cohort was explored to validate expression pattern of SHH, DHH, IHH, PTCH1, SMO and GLI1.

Clinical characteristics of the cohort

The study proceeded with prior approvals from biosafety and ethical committees of both COMSATS Institute of Information Technology and Holy Family Hospital Rawalpindi, Pakistan. Informed consents from the participants were collected after a thorough briefing of the proposed research. Tumour biopsies (n=150) along with adjacent normal mammary tissues (n=150) were collected immediately after surgery and snap frozen till further usage. Data regarding clinical and pathological findings were obtained in subsequent follow-up from respective laboratory reports and consultation.

RNA isolation and cDNA synthesis

RNA isolation was performed with TRIzol® (15596-018, Invitrogen, USA) from the respective biological specimens following manufacturer's instructions. Briefly, 1cm³ tissue section was homogenized using a hand held homogenizer in ice-cold RNA extraction buffer. Concentration of RNA was determined using nanodrop (Nanophotometer® Pearl, Implen, Germany). cDNA was synthesized using Revert Aid First

Strand cDNA Synthesis Kit (K1622, Thermo Scientific, USA).

Quantitative Real-time PCR

Primers for hedgehog pathway genes (SHH, DHH, IHH, PTCH1, SMO and GLI-1) were designed and synthesized from Integrated DNA Technology. β -actin was used to normalize the data. Sequences along with their amplicon sizes are provided in the supplementary table 1. Furthermore, primers for biomarkers ER (Estrogen receptor), PR (Progesterone receptor), HER-2 (Human Epidermal growth factor Receptor 2) and Ki-67 (Proliferative marker) used for molecular subtyping of breast cancer were also synthesized from IDT. VeriQuest SYBR Green qPCR Master Mix (75600200RXN, Thermo Scientific, USA) was used for qPCR in *Step One plus* (Applied Biosystem, USA). Reaction conditions included an initial denaturation at 95°C for 15min, followed by 40 cycles of denaturation at 95°C for 15 sec and annealing at 55°C for 1min in each cycle. Relative mRNA expression was calculated using the $2^{-\Delta\Delta C_t}$ method. QCanvas was used for the hierarchical clustering and visualizing heatmap of SHH, DHH, IHH, PTCH and SMO based on relative expression values [12].

Immunohistochemistry

Immunohistochemical staining was done using both tumour and normal frozen sections of 4 μ m thickness of given cohort bio-specimens. Initially, these slices were mounted on Super Frost Plus microscopic slides and air-dried for 30min. These fixed tissue sections were treated with 50% methanol and 50% acetone for 15 min. Sections were then air dried for 10 min and stored at -20°C (wrapped in foil) till further usage. Briefly, these slides were placed in PBS for 5 min to rehydrate, followed by blocking with 10% horse serum. These sections were later exposed with primary antibodies for SHH, DHH and GLI1. Antibodies used were rabbit anti-SHH polyclonal antibody (H-160; sc-9024, dilution 1:200; Santa Cruz Biotechnology, USA), mouse monoclonal anti-DHH (sc-271168, dilution 1:200; Santa Cruz Biotechnology, USA) and mouse anti-GLI1 monoclonal antibody (D1; sc-271075, diluted 1:200; Santa

Cruz Biotechnology, USA). Exclusion of primary antibody acted as a negative control. Immunostaining was performed as previously described [13]. Immuno-Reactive-Scores (IRS) were evaluated as the product of % of cells positively stained for each molecule categorized from 1 to 4 (1= $<25\%$, 2= $25-50\%$, 3= $>50\%$). Final IRS scores were ranked as high or low based on mean of IRS. Expression of pathway molecules was correlated with demographic and clinico-pathological findings of the cohort.

Breast cancer cell lines maintenance and culture condition

Breast cancer cell lines (MCF-7 and MDA-MB-231) were generously provided by Dr Yi Li (Breast centre Li, Baylor College of Medicine, USA). These lines were cultured and maintained as recommended by ATCC. Role of hedgehog pathway towards metastasis was assessed in these cell lines by inhibiting downstream effector GLI via GANT61 (G9048-5MG, Sigma, Germany).

Western blot analysis

Cells were homogenized in lysis buffer containing protease inhibitor. Concentration of the harvested proteins was determined using the Pierce BCA (Bicinchoninic Acid) protein assay kit (23225, Thermo Scientific, USA). Western blotting was performed following protocol described previously [14]. Both primary and secondary antibodies used for this study were purchased from Santa Cruz Biotechnology (USA) unless stated otherwise. Human SHH (H-160; sc-9024, dilution 1:1000), human PTCH-1 (H-267; sc-9016, dilution 1:500), human GLI-1 (H-300; sc-20867, dilution 1:1000) were used. Rabbit polyclonal antibody was used as secondary antibody for these blots. Mouse monoclonal antibody targeting against human Actin (C-4; sc-47778, dilution 1:3000) was used as control. The experiments were repeated three times independently.

Cell proliferation assay

The cell proliferation assay was assessed with CCK-8 (CK04-05, Dojindo, Japan) as reported [15]. Briefly, triplicate sets of 96 well plates retaining 5×10^3 cells from both MDA-MB-231 and MCF-7 cell

lines were prepared. After exposure with variable concentration (0, 5 μ M, 10 μ M, 15 μ M and 20 μ M) of GANT61, these plates were incubated at 37°C retaining 5% CO₂ for 48hrs. Assay was repeated at least three times to validate IC₅₀ value for GANT61. Furthermore, time dependent effect of GANT61 was evaluated at different time points (0, 24, 48, 72 and 96hrs).

Cell invasion assay

Using inserts of 8 μ m size placed in 24' well plate, invasion assays were performed. Briefly, each insert was initially coated with Matrigel (356234, BD Matrigel™ Basement Membrane Matrix, BD Biosciences, UK). After rehydration, 5x10⁴ cells of respective cell lines were seeded separately in the insert. Two set of inserts were used for each cell line. Cells in one insert were exposed to DMSO (Dimethyl sulfoxide) as control while in other insert GANT61 (10 μ M) was added. After 24hrs of incubation, inserts were fixed with formalin and stained in crystal violet [14]. Cells were counted under microscope to observe any possible invasion as mentioned (14).

Cell migration assay

A wound of ~300 μ m was artificially introduced into monolayer of breast cancer cell line and cells movement towards the wound was measured over span of 48hrs. Effect of GANT61 exposure towards cells motility was also calculated in these cell lines. Images were taken through a phase-contrast microscope (10 \times objective lens) at 0, 12, 24, 36 and 48hrs. The wound area was quantified with the Image J software [16]. Data was expressed as the mean of three independent experiments \pm standard deviation.

Statistical analysis

Wilcoxon signed rank test was used to evaluate difference between tumours and adjacent controls. Mann Whitney and Kruskal-Wallis Anova were applied to assess the association of molecules with different clinico-pathological parameters. Spearman correlation was also used to evaluate relationship between all

molecules in tumourigenesis. Unpaired t-test was applied to assess statistical differences in cell proliferation, migration and invasion assays. Kaplan-Meier plots were generated and survival was compared using log rank test. Cox's proportional hazard regression analyses were performed to evaluate hazard ratio of covariates for overall survival of patients. Statistical analyses were performed using Graphpad Prism 5 (GraphPad Software, Inc. CA, USA) and StatsDirect (<http://www.statsdirect.com>. StatsDirect Ltd. 2013, England).The value for $p < 0.05$ was considered significant.

List of Abbreviations

Sonic hedgehog (SHH), Indian hedgehog (IHH), Desert hedgehog (DHH), Patched1 (PTCH1), Smoothed (SMO), GLI1 (glioma-associated oncogene homolog 1), ER (Estrogen receptor), PR (Progesterone receptor), HER-2 (Human Epidermal growth factor Receptor 2) and Ki-67 (Proliferative marker), IRS (Immuno-Reactive-Scores), BCA (Bicinchoninic Acid), DMSO (Dimethyl sulfoxide).

Author's contributions

SKR designed the project, liaison with hospital surgeons and pathologists, collected the samples and clinical records, interviewed patients, performed lab experiments, analyzed and interpreted the data, wrote the paper and prepared figures and tables, JSK performed surgeries, contributed towards pathology studies and collected patient follow-up record, STAS contributed towards statistical analysis of data and preparation of tables, FW, LY and WGJ analyzed and interpreted the data and revised the manuscript of paper, MFAM designed the project, liaison with pathologist, analyzed the data, drafted and revised the paper manuscript.

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Competing interests

The authors of the study have no conflict of interest.

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References

1. Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012. *CA Cancer J Clin.* 2015; 65: 87–108. doi: 10.3322/caac.21262.
2. Gupta S, Takebe N, Lorusso P. Targeting the Hedgehog pathway in cancer. *Ther Adv Med Oncol.* 2010; 2: 237–50. doi: 10.1177/1758834010366430.
3. Lewis MT, Veltmaat JM. Next stop, the twilight zone: hedgehog network regulation of mammary gland development. *J Mammary Gland Biol Neoplasia.* 2004; 9: 165–81. doi: 10.1023/B:JOMG.0000037160.24731.35.
4. Ge X, Lyu P, Gu Y, Li L, Li J, Wang Y, Zhang L, Fu C, Cao Z. Sonic hedgehog stimulates glycolysis and proliferation of breast cancer cells: Modulation of PFKFB3 activation. *Biochem Biophys Res Commun.* 2015; 464: 862–8. doi: 10.1016/j.bbrc.2015.07.052.
5. Merchant AA, Matsui W. Targeting Hedgehog--a cancer stem cell pathway. *Clin Cancer Res.* 2010; 16: 3130–40. doi: 10.1158/1078-0432.CCR-09-2846.
6. Jeng K-S, Yu M-C, Hsiau H-I, Chang F-Y, Sheen I-S, Jeng W-J. High expression of Sonic Hedgehog signaling pathway genes indicates a risk of recurrence of breast carcinoma. *Oncotargets Ther.* 2013; 7: 79. doi: 10.2147/OTT.S54702.
7. Bièche I, Lerebours F, Tozlu S, Espie M, Marty M, Lidereau R. Molecular profiling of

- inflammatory breast cancer: identification of a poor-prognosis gene expression signature. *Clin Cancer Res.* 2004; 10: 6789–95. doi: 10.1158/1078-0432.CCR-04-0306.
8. Xu M, Li X, Liu T, Leng A, Zhang G. Prognostic value of hedgehog signaling pathway in patients with colon cancer. *Med Oncol.* 2012; 29: 1010–6. doi: 10.1007/s12032-011-9899-7.
 9. Li Q, Zhang Y, Zhan H, Yuan Z, Lu P, Zhan L, Xu W. The Hedgehog signalling pathway and its prognostic impact in human gliomas. *ANZ J Surg.* 2011; 81: 440–5. doi: 10.1111/j.1445-2197.2010.05585.x.
 10. Kim T-J, Lee JY, Hwang T-K, Kang CS, Choi Y-J. Hedgehog signaling protein expression and its association with prognostic parameters in prostate cancer: a retrospective study from the view point of new 2010 anatomic stage/prognostic groups. *J Surg Oncol.* 2011; 104: 472–9. doi: 10.1002/jso.21988.
 11. Gundem G, Perez-Llamas C, Jene-Sanz A, Kedzierska A, Islam A, Deu-Pons J, Furney SJ, Lopez-Bigas N, Heuer ML, Larsson E, Antipin Y, Reva B, Goldberg AP, et al. IntOGen: integration and data mining of multidimensional oncogenomic data. *Nat Methods.* American Association for Cancer Research; 2010; 7: 92–3. doi: 10.1038/nmeth0210-92.
 12. Kim N, Park H, He N, Lee HY, Yoon S. QCanvas: An Advanced Tool for Data Clustering and Visualization of Genomics Data. *Genomics Inform.* Korea Genome Organization; 2012; 10: 263–5. doi: 10.5808/GI.2012.10.4.263.
 13. Malik FA, Sanders AJ, Jones AD, Mansel RE, Jiang WG. Transcriptional and translational modulation of KAI1 expression in ductal carcinoma of the breast and the prognostic significance. *Int J Mol Med.* 2009; 23: 273–8.
 14. Malik FA, Sanders AJ, Kayani MA, Jiang WG. Effect of expressional alteration of KAI1 on breast cancer cell growth, adhesion, migration and invasion. *Cancer Genomics Proteomics.* International Institute of Anticancer Research; 2009; 6: 205–13.

15. Lin Z, Sheng H, You C, Cai M, Zhang Y, Yu LS, Yu X, Lin J, Zhang N. Inhibition of the CyclinD1 promoter in response to sonic hedgehog signaling pathway transduction is mediated by Gli1. *Exp Ther Med*. Spandidos Publications; 2017; 13: 307–14. doi: 10.3892/etm.2016.3969.
16. Schindelin J, Rueden CT, Hiner MC, Eliceiri KW. The ImageJ ecosystem: An open platform for biomedical image analysis. *Mol Reprod Dev*. 2015; 82: 518–29. doi: 10.1002/mrd.22489.
17. Szász AM, Lánckzy A, Nagy Á, Förster S, Hark K, Green JE, Boussioutas A, Busuttill R, Szabó A, Gyórfy B. Cross-validation of survival associated biomarkers in gastric cancer using transcriptomic data of 1,065 patients. *Oncotarget*. 2014; 7: 49322–33. doi: 10.18632/oncotarget.10337.
18. Chen X, Horiuchi A, Kikuchi N, Osada R, Yoshida J, Shiozawa T, Konishi I. Hedgehog signal pathway is activated in ovarian carcinomas, correlating with cell proliferation: it's inhibition leads to growth suppression and apoptosis. *Cancer Sci*. 2007; 98: 68–76. doi: 10.1111/j.1349-7006.2006.00353.x.
19. Jäger W, Thomas C, Fazli L, Hurtado-Coll A, Li E, Janssen C, Gust KM, So AI, Hainz M, Schmidtmann I, Roos FC, Thüroff JW, Brenner W, et al. DHH is an independent prognosticator of oncologic outcome of clear cell renal cell carcinoma. *J Urol*. 2014; 192: 1842–8. doi: 10.1016/j.juro.2014.07.013.
20. Im S, Choi HJ, Yoo C, Jung J-H, Jeon Y-W, Suh YJ, Kang CS. Hedgehog related protein expression in breast cancer: gli-2 is associated with poor overall survival. *Korean J Pathol*. 2013; 47: 116–23. doi: 10.4132/KoreanJPathol.2013.47.2.116.
21. Cui W, Wang L-H, Wen Y-Y, Song M, Li B-L, Chen X-L, Xu M, An S-X, Zhao J, Lu Y-Y, Mi X-Y, Wang E-H. Expression and regulation mechanisms of Sonic Hedgehog in breast cancer. *Cancer Sci*. 2010; 101: 927–33. doi: 10.1111/j.1349-7006.2010.01495.x.
22. Mukherjee S, Frolova N, Sadlonova A, Novak Z, Steg A, Page GP, Welch DR, Lobo-Ruppert SM,

- Ruppert JM, Johnson MR, Frost AR. Hedgehog signaling and response to cyclopamine differ in epithelial and stromal cells in benign breast and breast cancer. *Cancer Biol Ther.* 2006; 5: 674–83.
23. Azoulay S, Terry S, Chimingqi M, Sirab N, Faucon H, Gil Diez de Medina S, Moutereau S, Maillé P, Soyeux P, Abbou C, Salomon L, Vacherot F, de La Taille A, et al. Comparative expression of Hedgehog ligands at different stages of prostate carcinoma progression. *J Pathol.* 2008; 216: 460–70. doi: 10.1002/path.2427.
24. Xuan Y, Lin Z. Expression of Indian Hedgehog signaling molecules in breast cancer. *J Cancer Res Clin Oncol.* 2009; 135: 235–40. doi: 10.1007/s00432-008-0451-x.
25. Tao Y, Mao J, Zhang Q, Li L. Overexpression of Hedgehog signaling molecules and its involvement in triple-negative breast cancer. *Oncol Lett.* 2011; 2: 995–1001. doi: 10.3892/ol.2011.357.
26. Noman AS, Uddin M, Rahman MZ, Nayeem MJ, Alam SS, Khatun Z, Wahiduzzaman M, Sultana A, Rahman ML, Ali MY, Barua D, Ahmed I, Islam MS, et al. Overexpression of sonic hedgehog in the triple negative breast cancer: clinicopathological characteristics of high burden breast cancer patients from Bangladesh. *Sci Rep. Nature Publishing Group;* 2016; 6: 18830. doi: 10.1038/srep18830.
27. Koga K, Nakamura M, Nakashima H, Akiyoshi T, Kubo M, Sato N, Kuroki S, Nomura M, Tanaka M, Katano M. Novel link between estrogen receptor alpha and hedgehog pathway in breast cancer. *Anticancer Res.* 2008; 28: 731–40.
28. Diao Y, Azatyan A, Ferdous-Ur Rahman M, Zhao C, Zhu J, Dahlman-Wright K, Zaphiropoulos PG, Diao Y, Azatyan A, Ferdous-Ur Rahman M, Zhao C, Zhu J, Dahlman-Wright K, et al. Blockade of the Hedgehog pathway downregulates estrogen receptor alpha signaling in breast cancer cells. *Oncotarget. Impact Journals;* 2016; 7: 71580–93. doi: 10.18632/oncotarget.12259.
29. Ramaswamy B, Lu Y, Teng K -y., Nuovo G, Li X, Shapiro CL, Majumder S. Hedgehog Signaling

- Is a Novel Therapeutic Target in Tamoxifen-Resistant Breast Cancer Aberrantly Activated by PI3K/AKT Pathway. *Cancer Res.* 2012; 72: 5048–59. doi: 10.1158/0008-5472.CAN-12-1248.
30. Azim HA, Michiels S, Bedard PL, Singhal SK, Criscitiello C, Ignatiadis M, Haibe-Kains B, Piccart MJ, Sotiriou C, Loi S. Elucidating prognosis and biology of breast cancer arising in young women using gene expression profiling. *Clin Cancer Res.* 2012; 18: 1341–51. doi: 10.1158/1078-0432.CCR-11-2599.
 31. Azim HA, Partridge AH. Biology of breast cancer in young women. *Breast Cancer Res.* 2014; 16: 427. doi: 10.1186/s13058-014-0427-5.
 32. Souzaki M, Kubo M, Kai M, Kameda C, Tanaka H, Taguchi T, Tanaka M, Onishi H, Katano M. Hedgehog signaling pathway mediates the progression of non-invasive breast cancer to invasive breast cancer. *Cancer Sci.* 2011; 102: 373–81. doi: 10.1111/j.1349-7006.2010.01779.x.
 33. O'Toole S a., Machalek D a., Shearer RF, Millar EK a, Nair R, Schofield P, McLeod D, Cooper CL, McNeil CM, McFarland A, Nguyen A, Ormandy CJ, Qiu MR, et al. Hedgehog overexpression is associated with stromal interactions and predicts for poor outcome in breast cancer. *Cancer Res.* 2011; 71: 4002–14. doi: 10.1158/0008-5472.CAN-10-3738.
 34. ten Haaf A, Bektas N, von Serenyi S, Losen I, Arweiler EC, Hartmann A, Knüchel R, Dahl E. Expression of the glioma-associated oncogene homolog (GLI) 1 in human breast cancer is associated with unfavourable overall survival. *BMC Cancer.* 2009; . doi: 10.1186/1471-2407-9-298.
 35. Benvenuto M, Masuelli L, De Smaele E, Fantini M, Mattera R, Cucchi D, Bonanno E, Di Stefano E, Frajese GV, Orlandi A, Screpanti I, Gulino A, Modesti A, et al. In vitro and in vivo inhibition of breast cancer cell growth by targeting the Hedgehog/GLI pathway with SMO (GDC-0449) or GLI (GANT-61) inhibitors. *Oncotarget. Impact Journals, LLC;* 2016; 7: 9250–70. doi: 10.18632/oncotarget.7062.

36. Gu D, Xie J. Non-Canonical Hh Signaling in Cancer-Current Understanding and Future Directions. *Cancers (Basel)*. Multidisciplinary Digital Publishing Institute (MDPI); 2015; 7: 1684–98. doi: 10.3390/cancers7030857.

List of Figure Legends

Figure 1: Expression Profiling of Hedgehog Pathway Members in Pakistani Cohort.

a). Scatter plots showing mean relative mRNA expression of SHH, DHH, IHH, PTCH1, SMO and GLI1 in tumor samples as compared to normal tissues. b). Heatmap showing differential expression of analyzed molecules in tumors on left side and adjacent normal mammary tissues on right side. c). Immunostaining of SHH (A, B), DHH (C, D) and GLI1 (E, F) in normal tissues (A, C, E) and tumor tissues (B, D, F). (Wilcoxon Signed Rank Test, $p < 0.0001$).

Figure 2: Transcript Profiling of Hedgehog Pathway using Box Whisker Plots.

Association of relative expression of hedgehog pathway components with a). Nuclear grades, b). Tumor size, c). Nodal involvement, d). Metastasis, e). Age at disease onset, f). Menopausal status. (Kruskal-Wallis Anova (a, b and c) and Mann-Whitney U test (d, e and f), * $p < 0.05$, ** $p < 0.001$, *** $p < 0.0001$).

Figure 3: Correlation of Hedgehog molecules with Intrinsic Molecular Subtypes and Overall Survival of patients.

a). Expression variation of hedgehog pathway in molecular subtypes (Luminal A, Luminal B, HER2 and Triple negative) of breast cancer patients evaluated using qPCR. (Kruskal-Wallis Anova, * $p < 0.05$, *** $p < 0.0001$). b). Kaplan Meier plots showing association of SHH, DHH and

GLI1 with Overall survival in Pakistani population (red = high expression, blue = low expression, significant $p < 0.05$).

Figure 4: Effect of GANT61 exposure on SHH, PTCH1 and GLI1 using qPCR and Western blot.

Inhibition of expression of hedgehog pathway after treatment with GANT61 (10 μ M) for 48hrs. Relative mRNA expression of SHH, PTCH1 and GLI1 using qPCR in a). MDA-MB-231, b). MCF-7 (Mann Whitney U test, * $p < 0.05$, ** $p < 0.001$, *** $p < 0.0001$). c). Protein expression of SHH, PTCH1 and GLI1 using western blot having β -actin as internal control in both cell lines. All experiments were conducted in triplicates and were performed thrice.

Figure 5: Effect of GANT61 on proliferation, motility and invasion of breast cancer cells.

Inhibitory effect of GANT61 on proliferation, motility and invasion of breast cancer cells was tested using *in vitro* models. Cell viability assays were conducted using CCK-8, to observe the effect of GANT61 on cell proliferation in MDA-MB-231 and MCF-7 in a). Dose dependent (5, 10, 15, 20 μ M) and b). Time dependent manner (24, 48, 72, 96hrs). IC₅₀ of GANT61 was determined to be 10 μ M. MDA-MB-231 and MCF-7 were treated with the conditional medium containing 10 μ M GANT61 and the control medium. c). Migration was assessed using scratch assay and readings were taken after every twelve hour for 48hrs, upper panel of both cell lines represent 0hr and the lower panel represents 48hr. d). Graph showing difference of distance migrated between untreated and treated cells at each time point of scratch assay. e). Invasion was assessed using Boyden chamber transwell assays and difference between treated and untreated cells was evaluated after 24hrs. d). Graphical representation of number of cells invaded as mean

± S.D. between untreated and treated cells in both cell lines. (Unpaired t test, ***, ### p<0.0001). All experiments were conducted in triplicates and were performed thrice.

Tables

Table 1: Clinical correlation of hedgehog pathway with demographic and pathological variables in Pakistani breast cancer cohort.

Variable	Total	Mean±SD											
		SHH	p-value	DHH	p-value	IHH	p-value	PTCH	p-value	SMO	p-value	GLI-1	p-value
Tumour	150	4.4±3.3	< 0.0001	4.7±3.5	< 0.0001	0.04±0.2	< 0.0001	3.4±3.1	< 0.0001	1.4±2.1	0.61	3.8±2.5	< 0.0001
Control	150	1.0±0.9		1.0±1.4		1.0±1.1		1.0±1.0		1.0±1.3		1.0±0.8	
Disease onset age													
Age ≤ 45 years	76	5.6±3.9	0.03	5.5±4.2	0.04	0.2±0.8	0.65	3.2±3.1	0.26	1.9±2.2	0.03	6.3±4.5	0.03
Age ≥ 45 years	74	4.2±3.3		3.9±2.9		0.04±0.2		3.6±3.0		1.2±1.8		4.2±2.7	
Menopausal status													
Pre-Menopausal	80	6.8±4.8	0.0065	5.6±3.9	0.01	0.23±0.8	0.45	0.2±0.8	< 0.0001	2.8±3.6	0.12	7.9±4.3	< 0.0001
Post-Menopausal	70	4.5±3.5		3.9±3.0		0.05±0.2		3.9±3.3		2.1±3.1		3.9±2.5	
Laterality													
Right	75	4.2±2.8	0.01	5.8±4.0	0.001	0.2±0.8	0.13	3.4±2.9	0.97	2.5±2.9	0.26	4.5±2.9	0.02
Left	75	6.1±4.1		3.8±3.0		0.1±0.4		3.4±3.2		2.1±3.1		6.4±4.5	

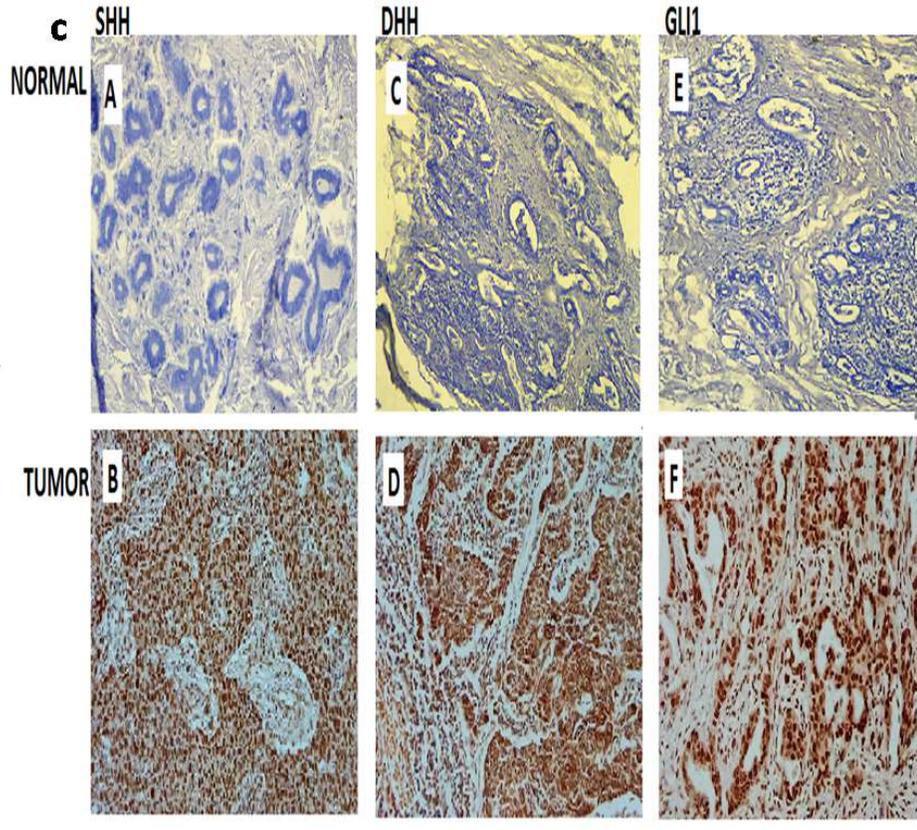
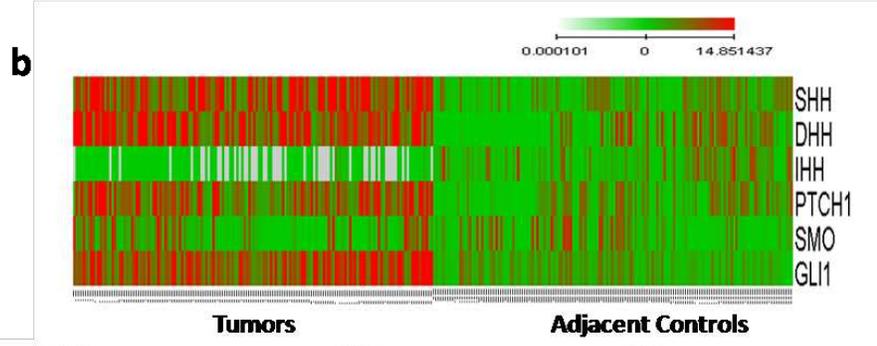
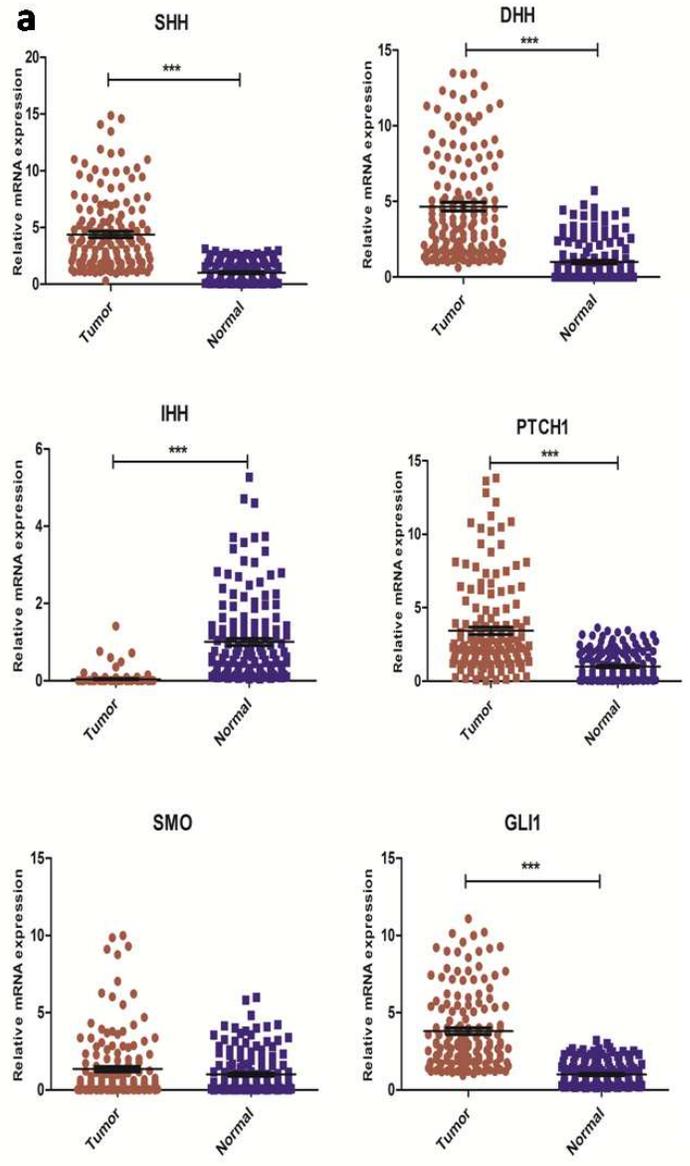
Grade-wise distribution													
Grade I	14	3.5±2.7	0.0001	3.4±2.4	0.48	0.2±0.9	0.2	2.9±2.9	0.68	2.4±3.5	0.11	2.8±1.9	0.01
Grade II	90	5.0±3.8		4.9±3.7		0.2±0.6		3.4±3.2		2.6±3.2		3.2±1.8	
Grade III	46	9.4±6.5		4.9±3.6		0.09±0.3		3.6±3.2		2.1±3.6		5.6±4.3	
Stage-wise distribution													
Stage I	34	4.2±3.3	< 0.0001	6.2±3.6	< 0.0001	0.1±0.5	0.03	3±2.8	0.35	2.6±3.3	0.09	6.7±3.9	0.005
Stage II	68	7.1±4.1		8.3±3.7		0.2±0.7		3.3±3.2		2.3±2.7		7.2±3.6	
Stage III	43	8.7±3.9		8.6±3.8		0.1±0.2		3.6±2.9		1.3±1.7		7.9±3.8	
Stage IV	5	14.6±2.9		17±2.2		0.06±0.1		4.9±2.8		2.3±1.9		14.5±3	
Nodal Involvement													
N0 (none)	48	4.3±3.2	< 0.0001	3.3±2.8	0.01	0.2±0.5	0.02	2.9±3	0.34	2.6±3.2	0.92	4.1±2.7	0.003
N1(lymph node<4)	78	6.1±4.1		5.8±4.2		0.2±0.7		3.6±3.2		2.3±3.3		5.9±3.6	
N2(lymph node>4)	24	8.2±3.7		6.5±4.4		0.006±0		3.6±2.8		2.7±3.9		6.6±3.9	
Distant Metastasis													
M0	145	4.4±3.4	0.0003	4.5±3.4	0.0002	0.14±0.6	0.09	3.3±3.0	0.12	2.4±3.4	0.34	3.8±2.5	0.0002
M1	5	14.6±2.9		17±2.2		0.06±0.1		4.9±2.8		2.3±1.8		14.5±3	
Molecular subtypes													
HER-2	21	2.7±2.4	< 0.0001	2.2±1.1	< 0.0001	0.03±0.1	0.73	4.3±4.5	0.04	2.5±3	0.63	3.7±2.1	0.0007
Luminal A	23	5.3±2.8		1.9±1.6		0.06±0.2		2.1±1.9		1.9±2.7		2.7±1.6	
Luminal B	79	6.7±2.7		4.9±3.5		0.1±0.4		3.7±3		2.6±3.4		5.2±2.9	
Triple Negative	27	5.9±3.2		2.9±2.6		0.3±0.8		4.1±3.8		1.8±2.8		3.8±2.5	

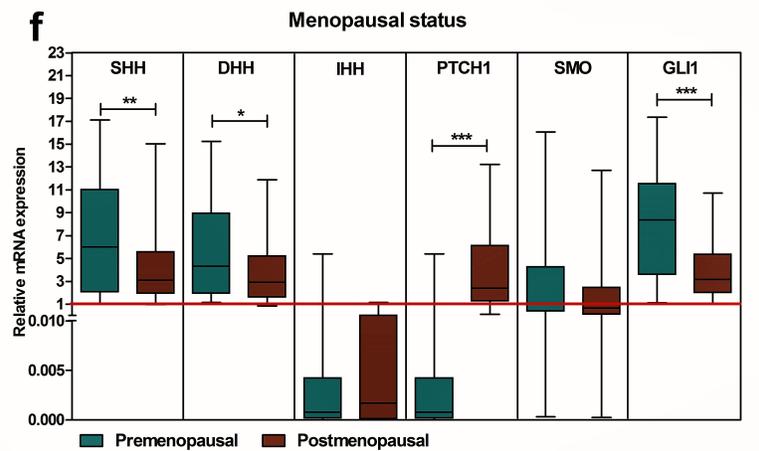
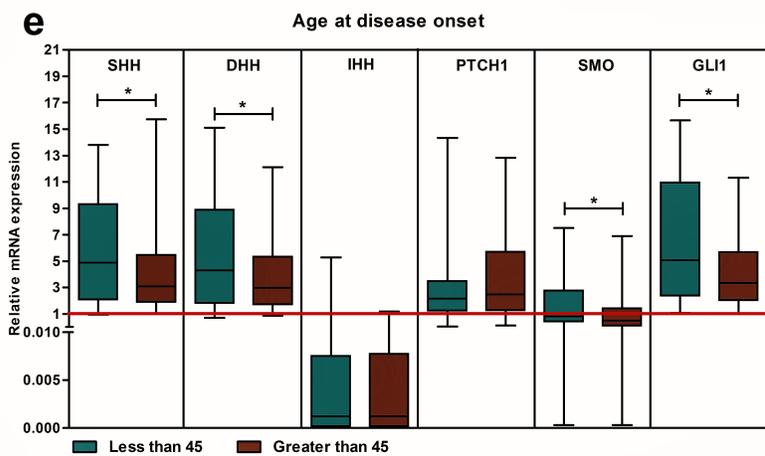
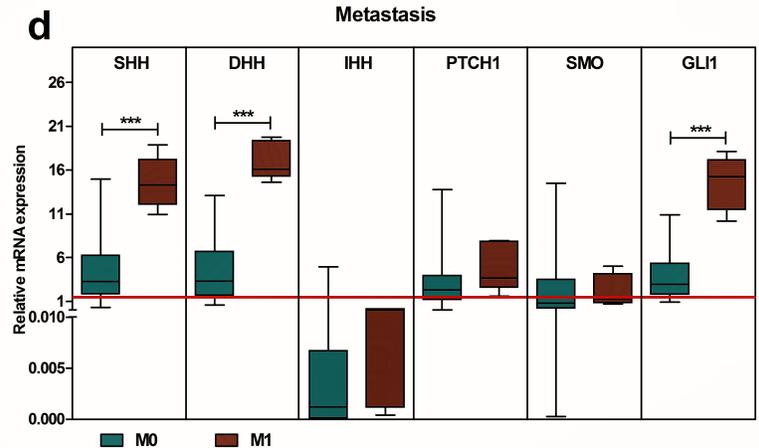
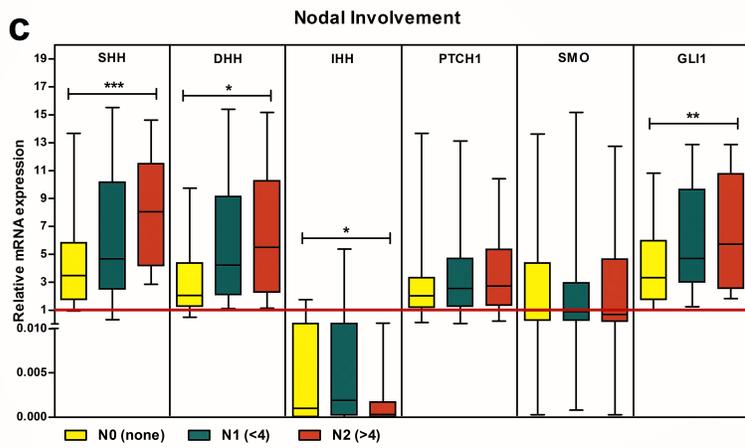
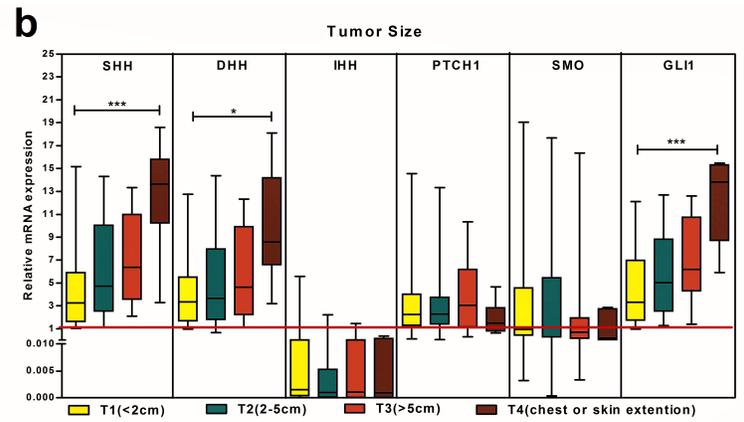
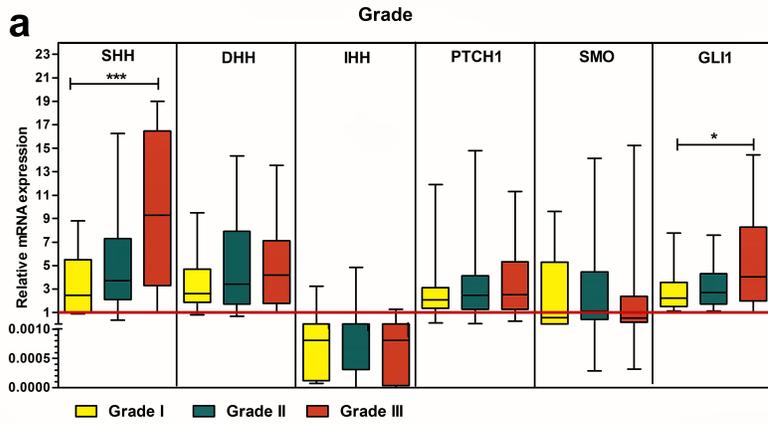
Table 2: Univariate and multivariate Cox's proportional hazard regression analyses of Pakistani breast cancer cohort for potential predictors of overall survival

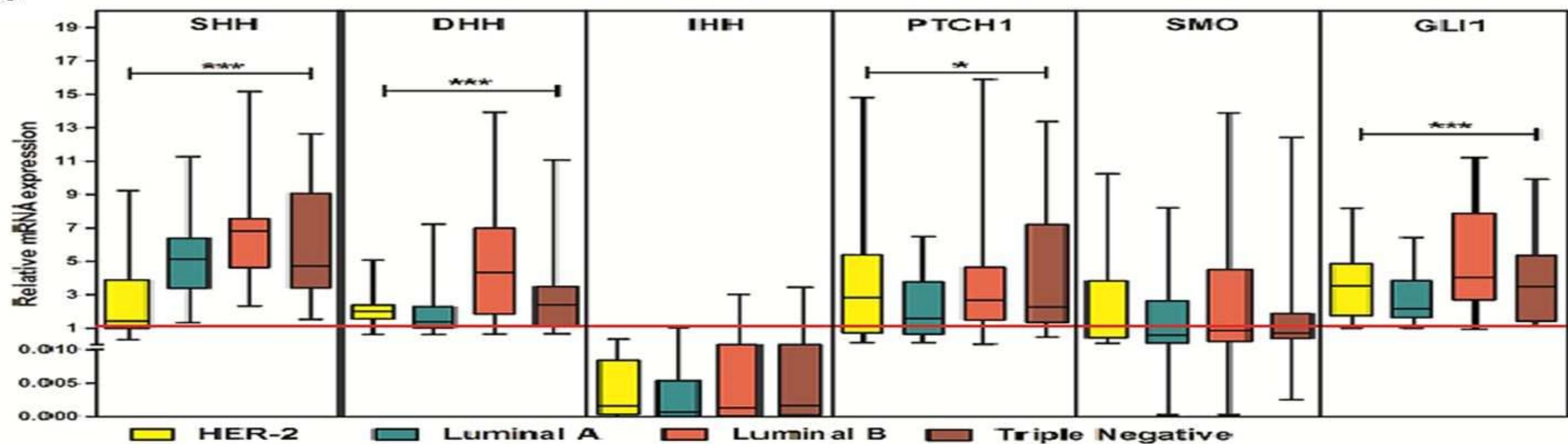
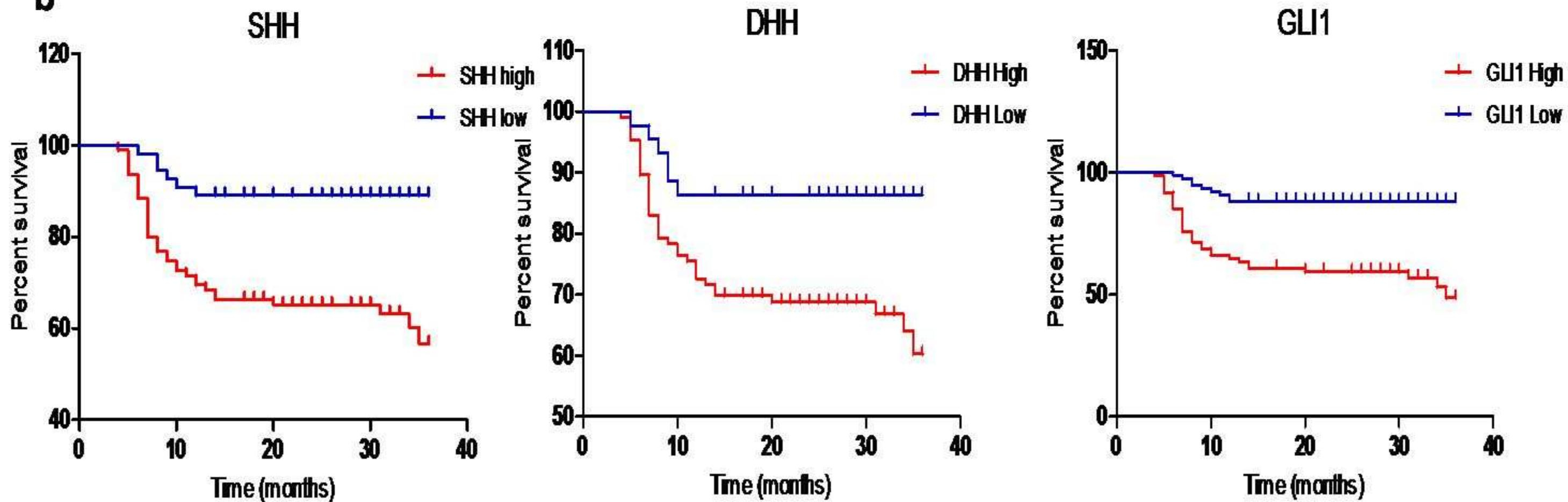
Characteristics	Univariate analysis		Multivariate analysis	
	HR(95% CI)	p value	HR(95% CI)	p value
SHH			3.14(1.3-7.7)	0.01
Age	0.18(0.06-0.45)	0.0003	0.2(0.08-0.56)	0.002
SHH	3.89(1.6-9.2)	0.002		
ER	2.67(1.2-6.0)	0.02	2.26(0.97-5.25)	0.05
SHH	3.53(1.5-8.4)	0.004		
PR	1.76(0.9-3.5)	0.09	-	-
SHH	3.58(1.5-8.6)	0.004		
HER-2	1.16(0.6-2.1)	0.6	-	-
SHH	3.93(1.7-9.3)	0.002		
Metastasis	2.96(1.1-8.3)	0.03	2.62(1.3-5.3)	0.007
SHH	3.79(1.6-9.0)	0.002		
DHH			2.83(1.2-6.8)	0.02

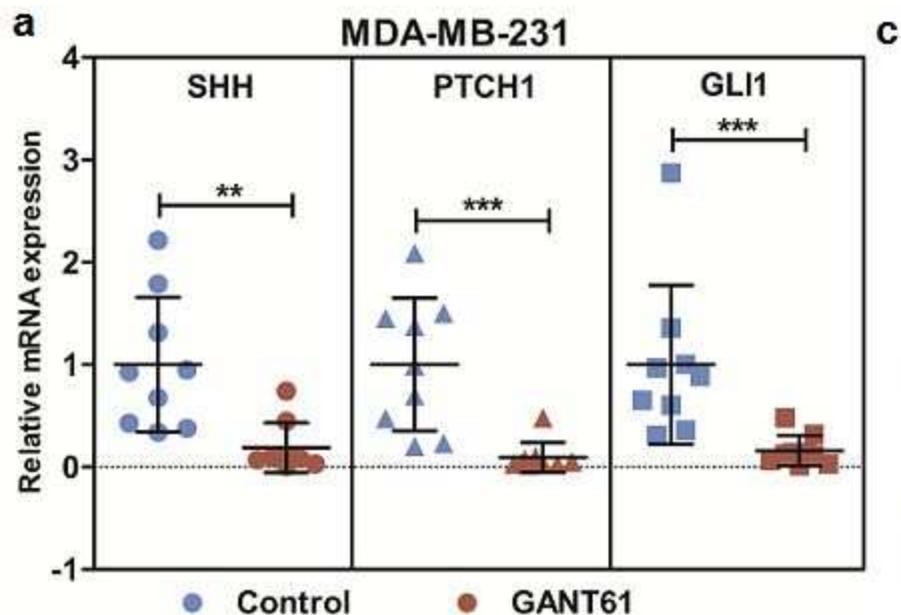
Age	0.16(0.07-0.43)	0.0002	0.17(0.1-0.5)	0.0003
DHH	2.94(1.2-6.9)	0.01		
ER	2.78(1.2-6.3)	0.01	2.00(0.8-4.9)	0.13
DHH	2.42(1.0-5.8)	0.04		
PR	1.97(1.0-3.85)	0.05	2.04(0.97-4.3)	0.06
DHH	2.56(1.1-6.1)	0.03		
HER-2	1.16(0.6-2.1)	0.62	-	-
DHH	2.72(1.2-6.5)	0.02		
Metastasis	3.97(1.9-7.9)	0.0001	2.9(1.4-5.9)	0.003
DHH	3.16(1.3-7.5)	0.009		
GLI1			3.67(1.7-7.9)	0.0008
Age	0.17(0.07-0.43)	0.0002	0.18(0.1-0.5)	0.0004
GLI1	4.67(2.2-9.8)	<0.0001		
ER	2.43(1.1-5.5)	0.03	2.45(1.1-5.6)	0.03
GLI1	3.98(1.9-8.4)	0.0003		
PR	1.26(0.6-2.6)	0.52	-	-
GLI1	4.15(1.9-9.1)	0.0004		
HER-2	1.01(0.5-1.9)	0.98	-	-
GLI1	4.51(2.2-9.5)	<0.0001		
Metastasis	2.54(1.3-5.1)	0.009	1.85(1.9-3.8)	0.009
GLI1	4.01(1.9-8.5)	0.0003		

Bold values show significance at $p < 0.05$, covariates significant in univariate analyses were considered for multivariate analyses.





a**Intrinsic molecular subtypes****b**



c

