- 1 Disentangling the biological pathways involved in early features of
- 2 Alzheimer's disease in the Rotterdam Study

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- 28 **Abstract**
- 29 **INTRODUCTION**,
- 30 Exploring the role of Alzheimer's disease (AD) implicated pathways in pre-dementia phase may
- 31 provide new insight for preventive and clinical trials targeting disease specific pathways.
- 32 **METHODS**,
- 33 We constructed weighted Genetic Risk Scores, first based on 20 genome-wide significant AD
- risk variants, second clustering these variants within pathways. Risk scores were investigated
- for their association with AD, mild cognitive impairment (MCI) and brain magnetic resonance
- imaging phenotypes including white matter lesions, hippocampal volume and brain volume.
- 37 **RESULTS**,
- 38 The risk score capturing endocytosis pathway was significantly associated with MCI (P =
- 39 1.44x10⁻⁴). Immune response (P = 0.016) and clathrin/AP2 adaptor complex pathway (P = 0.016)
- 3.55x10 $^{-3}$) excluding apolipoprotein E (APOE) also showed modest association with white
- matter lesions but did not sustain Bonferroni correction ($P = 9.09 \times 10^{-4}$).
- 42 **DISCUSSION**,
- 43 Our study suggests that the clinical spectrum of early AD pathology is explained by different
- 44 biological pathways in particular, the endocytosis, immune response and clathrin/AP2 adaptor
- 45 *complex* pathways that are independent of *APOE*.
- 46 **Keywords:** Genetic Risk Score, Alzheimer's disease, White matter lesions, Mild cognitive
- 47 impairment, Endocytosis, Immune response

Introduction

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Alzheimer's disease (AD) is heterogeneous and genetically complex disease with a high heritability (56-79 %) [1]. It has been known since the end of the previous century that a polymorphism in the apolipoprotein E (APOE) gene is the strongest common genetic risk factor [2-4]. This finding fueled speculations on the role of lipid metabolism and cholesterol transport pathway in AD in addition to the amyloid cascade and tau phosphorylation mechanism [5, 6]. Furthermore, large-scale genome-wide association studies (GWAS) have discovered over 20 novel common genetic variants that influence the risk of late-onset AD [7-13]. These common genetic variants have been mapped to eight biological pathways including immune response, endocytosis, cholesterol transport, hematopoietic cell lineage, protein ubiquitination, hemostasis, clathrin/AP2 adaptor complex and protein folding, each having a distinct biological function [14-16]. These eight pathways are not independent in that genes may be part of more than one biological pathway. For instance, APOE is part of four of the eight pathways namely cholesterol transport, hematopoietic cell lineage, clathrin/AP2 adaptor complex and protein folding pathways; clusterin (CLU) encoding for apolipoprotein J is involved in six pathways; phosphatidylinositol binding clathrin assembly protein(PICALM) and complement factor 1 (CR1) are involved in 2 pathways [14-16]. These diverse biological pathways may be responsible for the clinically heterogeneous manifestation of AD [17-19], which include endophenotypes such as changes in structural and functional magnetic resonance imaging (MRI) phenotypes, most notably hippocampal volume, total brain volume and white matter lesions [20, 21]. Furthermore, these biological pathways may also modulate the prodromal stages of AD such as mild cognitive impairment (MCI) [2224]. Owing to heterogeneity during pre-dementia phase, one important unanswered question is whether the different biological pathways that are implicated in AD relate to the pleiotropy of clinical endophenotypes. We hypothesized that some biological pathways are involved in distinct clinical endophenotypes while others may be involved in multiple or even all.

Disentangling the connection of biological pathways to various aspects of AD related early pathology may be a crucial step towards improving our understanding of the pathogenesis of AD and a first step towards a more informative and powerful read-out for preventive and therapeutic trials targeting specific pathways.

The current study aims to capture the different biological pathways involved in AD using genetic risk scores to evaluate their role in AD and pre-dementia endophenotypes including MCI, white matter lesion, total brain and hippocampal volume.

Methodology

Study population

This study included samples from the Rotterdam study (RS). RS is a prospective population based study [25] designed to investigate the etiology of age related disorders. At the baseline examination in 1990-93, study recruited 7983 subjects ≥ 55 years of age from the Ommoord district of Rotterdam (RS-I). At the baseline entry and after every 3 to 4 years, all the study participants were extensively interviewed and physically examined at the dedicated research center. During 2000 to 2001, the baseline cohort (RS-I) was expanded by adding 3011 subjects ≥55 years of age, who were not yet part of RS-I (RS-II). Second expansion of RS was performed by recruiting 3932 persons having ≥45 years of age during 2006-2008 (RS-III). The study has

been approved by the Medical Ethical Committee of Erasmus Medical Center and by the Ministry of Health, Welfare and Sport of the Netherlands. Written Informed consents were also obtained from each study participant to participate and to collect information from their treating physicians. Details of AD, dementia and MCI diagnosis is provided elsewhere [26, 27]. In the current analyses, we included in total 1270 late-onset AD cases and 7623 controls (age at follow-up ≥ 65 years and dementia free) from RS-I (1118 cases, 4736 controls), RS-II (134 cases, 1928 controls) and RS-III (18 cases, 959 controls) cohorts, from follow-up conducted during 2009-2013. 10370 dementia free (Normal) participants were also included in study from all three RS baseline cohorts and followed for an average of 11 years to analyze their progression into incident AD. Further, we included 360 MCI cases and 3245 cognitively normal controls from RS-I (235 cases, 1943 controls) and RS-II (125 cases, 1302 controls) who were first time assessed during 2002-2005 in RS (Table 1).

Genotyping

Blood was drawn for genotyping from participants of RS cohort during their first visit and DNA genotyping was performed at the internal genotyping facility of Erasmus Medical Center, Rotterdam. All samples were genotyped with the 550K, 550K duo, or 610K Illumina arrays. Genotyping quality control criteria include, call rate < 95%, Hardy-Weinberg equilibrium $P < 1.0 \times 10^{-6}$ and Minor Allele Frequency (MAF) < 1%. Moreover, study samples with excess autosomal heterozygosity, call rate < 97.5%, ethnic outliers and duplicate or family relationships were excluded during quality control analysis. Genetic variants were imputed from the Haplotype Reference Consortium (HRC) reference panel (version 1.0) [28], using the Michigan imputation server [29]. The server uses SHAPEIT2 (v2.r790) [30] to phase the

genotype data and performs imputation with Minimac 3 software [31]. For this study we used only genetic variants that had imputation quality (R-squared) > 0.5.

MRI scanning

Image acquisition

MRI scanning is assessed on a 1.5-T MRI unit with a dedicated eight-channel head coil (Signa HD platform, GE Healthcare, Milwaukee, USA) since the induction of a dedicated MRI machine in the Rotterdam Study in 2005. The MRI protocol was based on several high-resolution axial sequences, including a T1-weighted (slice thickness 0.8 mm), T2-weighted (1.6 mm), and fluid attenuated inversion recovery sequence (2.5 mm). A detailed description of the MRI protocol is described previously [32].

Image processing

we excluded 251 persons with stroke and/or dementia from the total 5899 subjects who came for MRI, since this may affect image processing. All T1 images were segmented into supratentorial gray matter, white matter and cerebrospinal fluid using a k-nearest neighbor (kNN) algorithm [33]. White matter lesions were segmented based on T1 tissue maps and an automatically detected threshold for the intensity of fluid-attenuated inversion recovery (FLAIR) scans [34]. After visual inspection of all segmentations, an additional 313 subjects were excluded due to poor quality, leaving 5335 for the analysis. The hippocampus was segmented using a fully automated method, described previously [35]. Semi-quantitative MRI post-processing software were used to measure intracranial volume and brain volume which included Elastix and custom-built software [36]. To calculate intracranial volume, non-brain tissues (skull, eyes, dura) were removed by non-linearly registering all brain

scans to a manually created template in which nonbrain tissues were masked. In all scans, visual checks were performed and if needed any segmentation errors manually corrected [36-38]. After excluding subjects whose genotyping information was not available, we ended up with 4527 cognitively normal subjects collectively from RS cohorts including RS-I (968), RS-II (1074) and RS-III (2485) cohorts for our current analyses.

Statistical analysis

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Genetic Risk Score computation

To construct the genetic risk score, we selected late-onset AD associated single nucleotide polymorphisms (SNPs) reaching genome-wide significance level ($P < 5.0 \times 10^{-8}$; Supplementary Table 1), including one rare TREM2 variant [7, 39]. In common variants, we considered only variants identified by the International Genomics of Alzheimer's Project (IGAP) meta-analyses. Additionally, we considered APOE*4 (rs429358) variant for genetic risk score construction. From a total of 21 SNPs, HLA-DRB1-HLA-DRB5 (rs9271192) variant was excluded from GRS calculation because of its low imputation quality (R-squared = 0.31) in RS. This led to a final selection of 20 independent genome-wide significant AD associated variants. Weighted genetic risk score was constructed using the effect sizes (log of OR) of the genome-wide significant variants from IGAP meta-analysis [7] as weights and their respective allele dosages from imputed genotype data of our study cohorts. Genetic risk score was constructed as the sum of the products of SNP dosages and their corresponding weights in R software (https://www.Rproject.org/). We constructed genetic risk score in two ways; 1) Combining all 20 selected variants and 2) Clustering the variants into their respective pathways.

1-Combined Genetic Risk Score (GRS1)

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GRS1 was constructed in two ways, i.e., 1) using all the 20 selected SNPs and 2) excluding the *APOE*4* variant to identify the joint independent effect of all other genome-wide significant SNPs.

2-Pathway-specific Genetic Risk Score (GRS2)

For GRS2, the genome-wide significant AD SNPs were divided into pathways (immune response, endocytosis, cholesterol transport, hematopoietic cell lineage, protein ubiquitination, hemostasis, clathrin/AP2 adaptor complex and protein folding pathway) identified by Jones et al. 2014 [16] (Supplementary Table 2). Classifying genome-wide significant AD SNPs into pathways, we also utilized information from Guerreiro et al, 2013 [14], in which the authors reviewed the possible division of known AD associated genes into biological pathways [14]. Further, Gene Network database (http://129.125.135.180:8080/GeneNetwork/) was used to confirm the allocated pathways. Of the 20 SNPs 14 could be clustered into 7 non mutually exclusive pathways (Supplementary Table 2). Similar to GRS1, we also constructed GRS2 with and without the APOE*4 variant. APOE*4 variant was grouped under four pathways including cholesterol transport [14], hematopoietic cell lineage, clathrin/AP2 adaptor complex and protein folding [16]. GRS2 was constructed for only those pathways, which could be assigned at least two SNPs, therefore protein ubiquitination pathway, which contained only one SNP, was excluded from all analyses, while hematopoietic cell lineage and protein folding pathways were also not considered in the analyses excluding APOE*4 variant.

Association analyses of GRS1, GRS2

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To test the association of AD and MCI with the risk scores we used logistic regression analysis in R software (<u>www.R-project.org</u>), using disease status as the outcome, <u>risk scores</u> as predictor and age and sex as covariates. In order to assess the possible inflation of association results between AD and risk scores, we repeated the association analysis excluding 625 AD cases which were part of IGAP meta-analysis [7] from total 1270 AD cases of the RS cohort. Further, we performed prospective analysis using Cox-proportional hazards model (N=1057 incident AD cases) in R software using 'survival' package [40] and reported results as hazard ratio (HR) per 1 standard deviation (SD) increase in risk score and 95% confidence interval. The association of single variants with AD and MCI in a logistic regression model adjusted for age and sex. Results of association analyses were reported as unstandardized regression coefficient and P values. To test the association of MRI phenotypes including total brain volume, white matter lesions and hippocampal volume with the risk scores we used linear regression adjusted for age, sex and intracranial volume at MRI scan. Single variant association analysis was also performed for MRI phenotypes. Bonferroni correction (0.05/(11 risk scores x 5 phenotypes); $P = 9.09 \times 10^{-04}$ was used to correct for multiple testing.

Results

Association of the **GRS1** with AD, MCI and MRI endophenotypes

The risk score containing all SNPs i.e., GRS1 both including APOE*4 (effect = 0.73, $P = 6.53 \times 10^{-74}$) and excluding APOE*4 (effect = 0.69, $P = 1.12 \times 10^{-11}$) was significantly associated with an increased risk of AD (Table 2). This association remained significant (APOE excluding; effect =

0.66, $P = 8.47 \times 10^{-7}$) after removing the patients that were included in the IGAP meta-analysis 197 198 [7] (Supplementary Table 3). GRS1 was also significantly associated with progression of normal subjects into incident AD including (HR = 1.69, $P = 6.64 \times 10^{-83}$) and excluding APOE*4 (HR = 1.27, 199 $P = 4.88 \times 10^{-15}$; Supplementary Table 4). GRS1 was associated with MCI when APOE*4 was 200 201 included (effect = 0.19, P = 0.012) but the association was stronger with MCI when APOE*4 was excluded from the analysis (effect = 0.59, $P = 9.51 \times 10^{-4}$; Table 3), but these associations did not 202 203 pass multiple testing correction. No association of GRS1 was observed with any of the MRI 204 phenotypes: white matter lesions, hippocampal volume, and total brain volume (Table 4). Association of the GRS2 with AD 205 Among GRS2 of which APOE*4 is a part, cholesterol transport, hematopoietic cell lineage, 206 clathrin/AP2 adaptor complex and protein folding were significantly associated with AD (effect 207 \geq 0.71, $P < 3.22 \times 10^{-64}$) only when APOE*4 was included in the risk scores (Table 2). Among the 208 non-APOE pathways, AD was significantly associated with GRS2 capturing immune response 209 210 (effect = 0.69, $P = 3.20*10^{-5}$) and endocytosis pathway (effect = 0.75, $P = 1.28 \times 10^{-5}$) and association sustained (Immune response; effect = 0.68, $P = 2.22 \times 10^{-3}$ and endocytosis; effect = 211 212 0.79, $P = 5.37 \times 10^{-4}$) after removing the patients that were included in the IGAP meta-analysis [7] (Supplementary Table 3). GRS2 capturing immune response (HR = 1.14, $P = 1.19 \times 10^{-5}$), 213 endocytosis (HR = 1.19, $P = 5.16 \times 10^{-8}$) and APOE*4 excluded clathrin/AP2 adaptor complex (HR 214 215 = 1.09, $P = 5.98 \times 10^{-3}$) pathway showed association with conversion risk from normal into 216 incident AD. Both Immune response and endocytosis pathways were significant after multiple testing. GRS2 including APOE*4 were also significantly associated with normal to AD conversion 217 (HR \geq 1.60, $P \leq$ 1.44x10⁻⁶⁹; Supplementary Table 4). Comparatively, except for APOE*4, CR1 and 218

BIN1, no single variant showed significant evidence of association (Supplementary Table 5). 219 220 BIN1 as a part of endocytosis pathways also partially explains the association of GRS2 capturing endocytosis with AD. 221 Association of the **GRS2** with MCI 222 In GRS2, only endocytosis pathway showed significant evidence for association (effect = 1.16, P 223 = 1.44x10⁻⁴; Table 3) with MCI and it retained significance after multiple testing. Although the 224 significance of the association is similar to that of the overall risk score (GRS1), the effect 225 estimate is considerably higher (1.16 versus 0.59 overall). In the single variant analysis, the 226 strongest association of MCI was observed with rs6733839 in the BIN1 gene (effect = 0.262, P = 227 1.12x10⁻³; Supplementary Table but this association lost significance after Bonferroni 228 correction. Whereas BIN1 is part of the endocytosis pathway, which partially explains the 229 association between MCI and GRS2 capturing endocytosis. 230 231 Association of the **GRS2** with MRI phenotypes White matter lesions were associated with GRS2 capturing immune response (effect = 0.15, P = 232 0.016), and clathrin/AP2 adaptor complex excluding APOE*4 (effect = 0.26, $P = 3.55 \times 10^{-3}$). If we 233 consider multiple testing, both these associations loses significance after accounting for all 234 235 tested phenotypes and risk scores. Of note is that the association of the GRS2 capturing the clathrin/AP2 adaptor complex loses its association when APOE*4 is included in the GRS2 (effect 236 = 0.011, P = 0.507). We did not observe association of GRS2 with hippocampal volume and total 237 brain volume. In the single variant analysis association of white matter lesions is seen with 238 239 variants in *PICALM*, *CLU* genes ($P \le 0.05$). Hippocampal volume shows association with variants

in BIN1 and CELF1 genes (P < 0.05; Supplementary Table 6). None of the single variant association sustained Bonferroni correction.

Discussion

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Combined GRS1 including and excluding APOE*4 is significantly associated with AD but not with MCI and MRI phenotypes in our study. Our study shows that the GRS2 capturing immune response and endocytosis pathways are not only significantly associated with AD, normal to AD conversion but also its endophenotypes, for instance, the GRS2 capturing the endocytosis pathway also associates significantly with MCI, a group at high risk of developing AD [41, 42], while the GRS2 capturing Immune response and clathrin/AP2 adaptor complex showed modest association with the presence of white matter lesions at MRI in cognitively normal subjects in the RS cohort (Supplementary Figure 1). In our study, the association of GRS1 with AD is consistent with other similar studies on AD [43-45]. GRS1 association with MCI did not pass Bonferroni correction while other studies observed significant association of combined risk score with MCI [46, 47]. We did not find association of GRS1 with any of the studied MRI endophenotypes. These findings are consistent with those of Mormino et al. 2016 [48] and Lupton et al. 2016 [49], both studies did not find association of hippocampal volume with combined GRS1 based on genome-wide signficant AD variants but Mormino et al. 2016 [48] observed this association only with risk score based on non-genome wide significant AD variants. The largest study so far that included RS, however, reported significant evidence of association of risk score based on all genome-wide significant AD variants with hippocampal volume and total brain volume [27].

This is the first study that addressed the role of specific pathways in AD and its early clinical manifestations i.e., MCI and MRI phenotypes. Our study shows that GRS2 based on immune response pathway was significantly associated with AD, normal to AD conversion. We also observed evidence of association of *immune response* with white matter lesions at MRI but this did not pass Bonferroni correction, therefore should be considered carefully while interpreting results. These findings are converging with studies showing enrichment of immune system pathway with non-genome wide significant AD variants [16, 50]. The genes clustered in immune response pathway (CLU, CRI, INPP5D, MS4A6A, TREM2, MEF2C, EPHA1) are mainly expressed in microglial cells and play a part in the innate immune response in central nervous system [51-55]. Microglial cells are also thought to play a role in amyloid plaque clearance [56, 57]. It has been hypothesized that activation of immune system and the subsequent inflammatory response are involved in neuronal damage including axonal loss and white matter pathology due to demyelination [58]. This may explain the association of the AD genes involved in the immune response with white matter lesions that we observed in the present study [59]. Whit matter lesions are associated with increased risk of cognitive decline, developing dementia [21] and AD [60, 61]. White matter lesions are also more frequently observed in AD patients than controls [62, 63]. The present study reveals further that the genes capturing the *endocytosis pathway* not only strongly associate to AD but also to MCI. We also showed that *endocytosis* pathway is also associated with progression from normal (dementia free) to AD in average 11 years of followup. This pathway is independent of APOE and includes the BIN1, PICALM, CD2AP, SORL1 genes. We show that the association of GRS1 with MCI status is mainly attributed to the genes

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involved in the *endocytosis* pathway. Omitting the AD genes not related to the *endocytosis* pathway makes the association of the pathway with MCI even stronger. The association suggests that the endocytosis pathway plays a critical role in an early prodromal phase of AD and converges with studies suggesting activation of endocytic pathway is the earliest reported intracellular manifestation of AD [41, 42, 64]. Based on the effect estimates of association of endocytosis pathway with AD and MCI (0.79 vs 1.59), we also speculated that endocytosis is more strongly associated to MCI than to AD. MCI is considered a prodromal stage in AD patients that suggest endocytosis pathway is associated with early pathology of AD. The endocytosis pathway is involved in neuronal uptake of macromolecules and secretory vesicles during synaptic transmission. As efficient uptake of extracellular cholesterol is critical for neuronal functions such as repair, synapse formation and exon elongation [65], normal neuronal work needs smooth functioning of endocytosis pathways [66]. Post-mortem studies have also demonstrated reduced brain cholesterol levels in the brain areas responsible for memory and learning, among late-onset AD cases and age matched controls [67]. These facts suggest that defects in endocytosis which derive the cholesterol uptake could lead to impaired neurotransmitter release and synaptic function [68]. Dysfunction in endocytosis can also contribute to accumulation of abnormal Aß peptide [69]. Based on this finding, we can suggest that *endocytosis* pathway is a common molecular mechanism between MCI and AD that starts manifesting at early stages of disease. Risk contributed by variants clustered in this pathway at various stages of AD progression can possibly provide clue about disease trajectory. Similar to the *immune response* pathway, the *clathrin/AP2 adaptor complex* pathway modestly

associated with white matter lesions. Although, the association failed to pass the multiple

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testing, but combined AD risk score did not capture the association with white matter lesions in our and even in large studies [27]. Capturing this association in small sample indicates the importance of this pathway in explaining the white matter lesions pathology. Two variants tagging PICALM and CLU genes cluster in the clathrin/AP2 adaptor complex pathway. Each variant independently shows nominally significant association with white matter lesions in our analyses but combining their effect are additive and improve the strength of association. There is a strong evidence that the two protein encoded by the genes interact at molecular level [70, 71]. PICALM is involved in VAMP2 trafficking that is a crucial process to maintain functional integrity of synapses which are crucial to cognitive function [72, 73]. PICALM is also found to be expressed in the white matter and, immune-labeling of human brain tissue shows that PICALM is mainly found in blood vessel walls [74]. CLU clustered in clathrin/AP2 adaptor is involved in efflux of free insoluble amyloid beta (Aβ) peptide through blood brain barrier [75]. Increased plasma levels of *CLU* were found to be associated with increased burden of Aß peptide in healthy elderly population and brain atrophy in AD [76, 77] and decreased integrity of white matter in young adults [78]. Demyelination of white matter is reported to occur even before the accumulation of AB plaques and neurofibrillary tangles [79]. The findings of the present study suggest that the increased genetic burden of risk variants in clathrin/AP2 adaptor complex (clathrin mediated endocytosis) and immune response pathway may play a role in early pathogenesis of AD through white matter pathology. Among pathways including APOE (Cholesterol transport, hematopoietic cell lineage,

clathrin/AP2 adaptor complex, protein folding), significant association with AD and normal to

AD conversion suggests that APOE*4 appear to be the driving genetic factor for these

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associations. Only the clathrin/AP2 adaptor complex shows evidence of association (P = 0.036) 327 328 to AD and normal to AD conversion ($P = 5.98 \times 10^{-3}$) when APOE*4 variant is excluded from the analysis but did not pass multiple testing correction. Our study provides a readout of pathway based risk score association with AD and its pre-330 dementia endophenotypes. Main clinical significance of our findings is that they will allow to 331 determine whether a certain biological pathway is involved in an individual patient. This will 332 permit targeted interventions based on predicted pathological pathways. Similar as the case of 333 cardiovascular diseases [80], a heterogeneous disease treatment can be followed based on 334 335 pathway biomarkers (e.g., glucose level, total cholesterol and high density lipid levels, liver enzymes in case of cardiovascular disease) [81] but rather on genetic basis. This require 336 337 reference pathways and treatment portfolio. In the meantime, the pathway based genetic risk score will allow stratification of the high risk patients in clinical trials based on causal pathways 338 involved in patients. This may improve both the power and efficiency of future clinical and 339 preventive trials. 340 341 Our study is a step forward to use known genetic and pathway information for disentangling the mechanisms of AD but it has one major limitation that pathway information is based on 342 known AD variants identified so far. This will further improve in future with improved genetic 343 risk information that can better capture the underlying pathways. Another possible limitation of 344 our study is that 625 cases of RS-I was a part of meta-analysis performed by IGAP [7] which can 345 346 contribute to possible inflation in our results of association of risk score with AD. However, excluding these patients, the results of this study largely remained unchanged.

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In conclusion we found different pathways are implicated in different endophenotypes of AD. *Endocytosis* pathway is involved in MCI and AD, while *immune response* associates with AD. Further, *Immune response* and *clathrin/AP2 adaptor complex* pathways are involved in white matter lesions but their association should be carefully considered due to multiple testing limitation. Interestingly, all the observed associations with early AD pathology are shown by *APOE* excluding pathways. Future findings from genomic research will improve the quality of the pathway-specific genetic scores.

Acknowledgements

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This study was funded by the PERADES Program (Defining Genetic, Polygenic and 357 Environmental Risk for Alzheimer's disease, using multiple powerful cohorts, focused 358 359 Epigenetics and Stem cell metabolomics), Project number 733051021. The Rotterdam Study is 360 funded by Erasmus Medical Center and Erasmus University, Rotterdam, Netherlands Organization for the Health Research and Development (ZonMw), the Research Institute for 361 Diseases in the Elderly (RIDE), the Ministry of Education, Culture and Science, the Ministry for 362 363 Health, Welfare and Sports, the European Commission (DG XII), and the Municipality of 364 Rotterdam. The authors are grateful to the study participants, the staff from the Rotterdam 365 Study and the participating general practitioners and pharmacists. The generation and management of GWAS genotype data for the Rotterdam Study (RS-I, RS-II, RS-III) was executed 366 by the Human Genotyping Facility of the Genetic Laboratory of the Department of Internal 367 Medicine, Erasmus MC, Rotterdam, The Netherlands. The GWAS datasets are supported by the 368 Netherlands Organization of Scientific Research NWO Investments (Project number 369 175.010.2005.011, 911-03-012), the Genetic Laboratory of the Department of Internal 370 Medicine, Erasmus MC, the Research Institute for Diseases in the Elderly (014-93-015; RIDE2), 371 372 the Netherlands Genomics Initiative (NGI)/Netherlands Organization for Scientific Research 373 (NWO) Netherlands Consortium for Healthy Aging (NCHA), project number 050-060-810. We thank Pascal Arp, Mila Jhamai, Marijn Verkerk, Lizbeth Herrera and Marjolein Peters, MSc, and 374 Carolina Medina-Gomez, MSc, for their help in creating the GWAS database, and Karol Estrada, 375 376 PhD, Yurii Aulchenko, PhD, and Carolina Medina-Gomez, MSc, for the creation and analysis of 377 imputed data.

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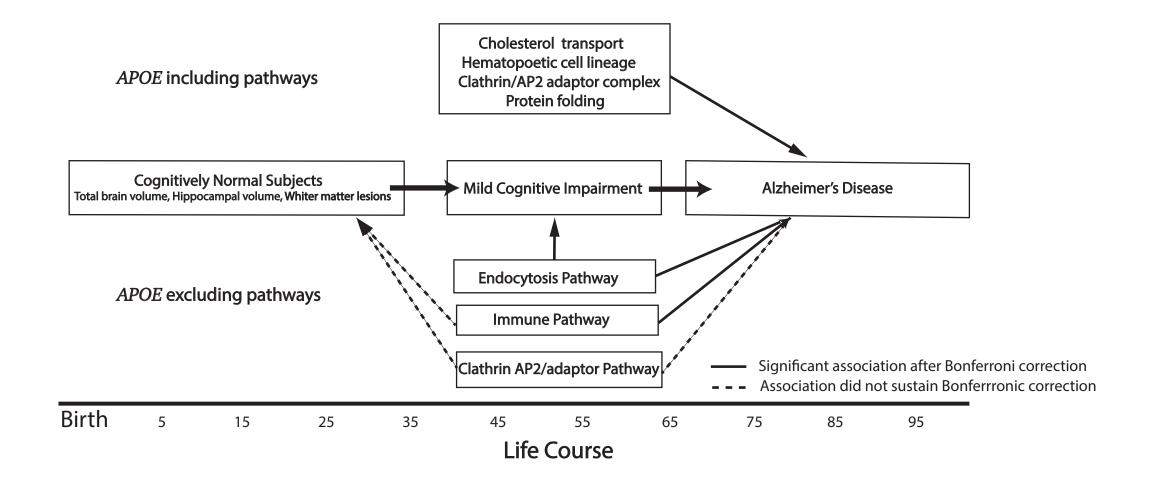
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Figure caption:

590	Supplementary Figure 1 : Diagram showing the association results of AD, MCI and WML in
591	cognitively normal subjects with pathways-specific GRS2 both including and excluding
592	APOE*4. Bonferroni correction threshold ($P = 9.09 \times 10^{-4}$)
593	



Supplementary Material

Supplementary Table 1: List of Genome-wide significant variants associated with AD

Chromosome	Position (BP)	SNP	Gene	Coding Allele	Effect	MAF	RSI_Rsq	RSII_Rsq	RSIII_Rsq	Effect origin
1	207692049	rs6656401	CR1	G	-0.157	0.197	<mark>0.953</mark>	<mark>0.948</mark>	<mark>0.950</mark>	Lambert et al. (2013)[1]
2	127892810	rs6733839	BIN1	Т	0.188	0.409	<mark>0.960</mark>	<mark>0.911</mark>	<mark>0.962</mark>	Lambert et al. (2013)
2	234068476	rs35349669	INPP5D	Т	0.066	0.488	<mark>0.975</mark>	<mark>0.973</mark>	<mark>0.976</mark>	Lambert et al. (2013)
5	88223420	rs190982	MEF2C	Α	0.08	0.408	<mark>0.979</mark>	<mark>0.934</mark>	<mark>0.978</mark>	Lambert et al. (2013)
6	41129252	rs75932628	TREM2	Т	0.889	0.0016	<mark>0.762</mark>	<mark>0.726</mark>	<mark>0.668</mark>	Ruiz et al. (2014)[2]
6	32578530	rs9271192	HLA-DRB5-HLA-DRB1 [†]	Α	-0.108	0.276	<mark>0.314</mark>	<mark>0.312</mark>	<mark>0.314</mark>	Lambert et al. (2013)
6	47487762	rs10948363	CD2AP	G	0.098	0.266	<mark>0.998</mark>	<mark>0.998</mark>	<mark>0.998</mark>	Lambert et al. (2013)
7	100004446	rs1476679	ZCWPW1	Т	0.078	0.287	<mark>0.995</mark>	<mark>0.996</mark>	<mark>0.995</mark>	Lambert et al. (2013)
7	143110762	rs11771145	EPHA1	Α	-0.102	0.338	<mark>0.998</mark>	<mark>0.998</mark>	<mark>0.999</mark>	Lambert et al. (2013)
7	37841534	rs2718058	NME8	G	-0.07	0.373	1.000	1.000	<mark>1.000</mark>	Lambert et al. (2013)
8	27195121	rs28834970	PTK2B	С	0.096	0.366	<mark>0.993</mark>	<mark>0.990</mark>	<mark>0.994</mark>	Lambert et al. (2013)
8	27467686	rs9331896	CLU	Т	0.146	0.379	<mark>0.902</mark>	<mark>0.974</mark>	<mark>0.901</mark>	Lambert et al. (2013)
11	121435587	rs11218343	SORL1	С	-0.27	0.039	<mark>0.998</mark>	<mark>0.995</mark>	<mark>0.998</mark>	Lambert et al. (2013)
11	47557871	rs10838725	CELF1	С	0.075	0.316	<mark>0.998</mark>	<mark>0.998</mark>	<mark>0.998</mark>	Lambert et al. (2013)
11	59923508	rs983392	MS4A6A	G	-0.108	0.403	<mark>0.989</mark>	<mark>0.990</mark>	<mark>0.991</mark>	Lambert et al. (2013)
11	85867875	rs10792832	PICALM	G	0.13	0.358	<mark>0.999</mark>	<mark>0.999</mark>	<mark>0.999</mark>	Lambert et al. (2013)
14	53400629	rs17125944	FERMT2	С	0.122	0.092	1.000	1.000	1.000	Lambert et al. (2013)
14	92926952	rs10498633	SLC24A4-RIN3	Т	-0.104	0.217	<mark>0.999</mark>	<mark>0.999</mark>	1.000	Lambert et al. (2013)
19	45411941	rs429358	APOE*4	С	1.3503	0.148	<mark>0.949</mark>	<mark>0.944</mark>	<mark>0.947</mark>	Lambert et al. (2013)
19	1063443	rs4147929	ABCA7	G	-0.135	0.19	<mark>0.916</mark>	<mark>0.917</mark>	<mark>0.991</mark>	Lambert et al. (2013)
20	55018260	rs7274581	CASS4	С	-0.139	0.083	<mark>0.990</mark>	<mark>0.989</mark>	<mark>0.990</mark>	Lambert et al. (2013)

Abbreviations: AD ~ Alzheimer's disease, MAF ~ minor allele frequency, Rsq = R-squared (HRC imputation quality)

^{*}Effect (log of odds ratio) is based on Coding Allele column

[†]Variants have low imputation quality in HRC imputation in our cohorts therefore excluded from genetic risk score calculation .

Supplementary Table 2: Clustering of genome-wide significant variants into their respective 8 biological pathways.

			Genes repo	rted in pathway	Constructed GRS(Yes/No)		
Pathway	Gene*	Assigned SNP	Jones et al	Guerreiro et al	Including APOE	Excluding APOE	
	CLU	rs9331896	Yes	Yes			
	CR1	rs6656401	Yes	Yes			
	INPP5D	rs35349669	Yes	Yes			
Immune Response	EPHA1	rs11771145	-	Yes	No	Yes	
	MS4A6A	rs983392	-	Yes			
	TREM2	rs75932628	-	Yes			
	MEF2C	rs190982	-	Yes			
	CD2AP	rs10948363		Yes			
For de autoria	PICALM	rs10792832	Yes	Yes	N1 -	Yes	
Endocytosis	BIN1	rs6733839	Yes	Yes	NO	Yes	
	SORL1	rs11218343	-	Yes			
	CLU	rs9331896	Yes	Yes			
Cholesterol transport†	ABCA7	rs4147929	Yes	Yes	W	W	
•	SORL1	rs11218343	-	Yes	Yes	Yes	
	APOE*4	rs429358	Yes	Yes			
	CR1	rs6656401	Yes	-	.,		
Hematopoietic cell lineage†	APOE*4	rs429358	Yes	-	No Yes Yes No No Yes	No	
Protein ubiquitination	CLU	rs9331896	Yes	-	No	No	
	CLU	rs9331896	Yes	_			
Hemostasis	INPP5D	rs35349669	Yes	-	No	Yes	
	CLU	rs9331896	Yes	-			
Clathrin/AP2 adaptor complex†	PICALM	rs10792832	Yes	-	Yes	Yes	
	APOE*4	rs429358	Yes	-			
	CLU	rs9331896	Yes	-			
Protein folding†	APOE*4	rs429358	Yes	-	Yes	No	

^{*}SNPs classification into pathways is based on the information from Jones et al. [3] and Guerreiro et al[4].

[†]APOE*4 variant (rs429358) is grouped under these pathways. Protein folding and Hematopoietic cell lineage pathways are left with one SNP after excluding APOE*4 variant therefore were not considered for APOE excluding analysis.

Supplementary Table 3: Results of association of AD with risk scores

		Including A	POE	Excluding APOE					
SNP Cluster*	β	SE	P-value	β	SE	P-value			
GRS1 (Combined)	0.72	0.052	1.80x10 ⁻⁴³	0.66	0.135	8.45x10 ⁻⁷			
Immune response	-	-	-	0.68	0.221	2.22x10 ⁻³			
Endocytosis	-	-	-	0.79	0.228	5.37x10 ⁻⁴			
Cholesterol Transport	0.70	0.054	3.48x10 ⁻³⁸	0.41	0.294	0.159			
Hematopoietic cell lineage [†]	0.72	0.055	7.19x10 ⁻⁴⁰		-	-			
Hemostasis			-	0.35	0.390	0.364			
Clathrin/AP2 Adaptor complex	0.70	0.055	6.47x10 ⁻³⁸	0.27	0.314	0.383			
Protein folding†	0.71	0.055	5.18x10 ⁻³⁸		-	-			

Abbreviations: GRS1 \sim Combined genetic risk score, SNP \sim single nucleotide polymorphism, $\beta \sim$ regression coefficient, SE \sim Standard error.

Supplementary Table 4: Results of longitudinal analysis from normal (dementia free) to AD conversion

		Including AP	OE	Excluding APOE					
SNP Cluster*	HR	95% CI	P-value	HR	95% CI	P-value			
GRS1 (Combined)	1.69	1.61-1.79	6.64x10 ⁻⁸³	1.27	1.19-1.34	4.88 x10 ⁻¹⁵			
Immune response	-	-	-	1.14	1.07-1.21	1.19 x10 ⁻⁵			
Endocytosis	-	-	-	1.19	1.12-1.26	5.16 x10 ⁻⁸			
Cholesterol Transport	1.60	1.52-1.68	1.44x10 ⁻⁶⁹	1.07	1.01-1.14	2.45 x10 ⁻²			
Hematopoietic cell lineage [†]	1.60	1.52-1.68	4.29x10 ⁻⁷¹	-	-	-			
Hemostasis	-	-	-	1.08	1.01-1.14	1.62 x10 ⁻²			
Clathrin/AP2 Adaptor complex	1.61	1.52-1.69	4.21x10 ⁻⁷¹	1.09	1.02-1.15	5.98 x10 ⁻³			
Protein folding†	1.60	1.52-1.68	7.66x10 ⁻⁷⁰	-	-	-			

Note: Multiple testing correction by Bonferroni 0.05/ (5 phenotypes x 11 risk scores); $P < 9.09 \times 10^{-4}$ was considered significant

Abbreviations: GRS1 ~ Combined genetic risk score, SNP ~ single nucleotide polymorphism,

HR ~ Hazard ratio per 1 standard deviation of risk score, Cl ~ Confidence interval

*Cox proportional hazards model adjusted for age at baseline and sex in RS (N = 10370 normal at baseline)

[†]Only one SNP available in excluding APOE GRS2

^{*}Logistic regression model adjusted for age and sex in RS (N=645) excluding 625 cases included in IGAP meta-analysis

[†]Only one SNP available in excluding APOE GRS2

Supplementary Table 5: Results of single variant association with AD and MCI

Phenotype ->		Alzheimer's	Disease		Mild	Cognitive In	npairment
SNP	Gene	β	SE	P-value	β	SE	P-value
rs429358	APOE*4	0.949	0.057	8.78x10 ⁻⁶³	0.137	0.114	0.229
rs75932628	TREM2	0.816	0.470	0.083	1.293	0.639	0.043
rs6656401	CR1	-0.180	0.055	1.11x10 ⁻³	0.072	0.105	0.496
rs11218343	SORL1	-0.161	0.110	0.142	-0.193	0.206	0.351
rs10838725	CELF1	0.062	0.048	0.192	0.069	0.086	0.424
rs983392	MS4A6A	-0.081	0.045	0.072	-0.127	0.081	0.115
rs10792832	PICALM	0.085	0.045	0.063	0.181	0.083	0.028
rs17125944	FERMT2	0.128	0.070	0.067	-0.016	0.131	0.901
rs10498633	SLC24A4-RIN3	-0.092	0.053	0.081	0.049	0.094	0.601
rs4147929	ABCA7	-0.003	0.061	0.961	-0.064	0.107	0.552
rs6733839	BIN1	0.149	0.045	9.74x10 ⁻⁴	0.262	0.081	1.12x10 ⁻³
rs35349669	INPP5D	0.055	0.044	0.216	-0.081	0.080	0.315
rs7274581	CASS4	-0.120	0.083	0.149	-0.034	0.143	0.810
rs190982	MEF2C	0.010	0.046	0.833	0.061	0.083	0.458
rs10948363	CD2AP	0.079	0.049	0.102	0.030	0.088	0.732
rs1476679	ZCWPW1	0.005	0.047	0.918	-0.037	0.085	0.662
rs11771145	EPHA1	-0.075	0.047	0.107	-0.117	0.085	0.168
rs2718058	NME8	-0.051	0.045	0.258	0.054	0.081	0.507
rs28834970	PTK2B	0.071	0.045	0.111	0.034	0.081	0.679
rs9331896	CLU	0.049	0.047	0.290	0.026	0.084	0.762

Note: Multiple testing correction by Bonferroni 0.05/ (5 phenotypes x 20 variants); $P < 5 \times 10^{-4}$ was considered significant

Abbreviations: SNP ~ Single nucleotide polymorphism, β ~ regression coefficient, SE ~ Standard error

Supplementary Table 6: Single variant association with MRI phenotypes

		White matter	lesions		Hip	opocampal vol	ume		Total Brain volume				
SNP	Gene	β	SE	P-value	β	SE	P-value	β	SE	P-value			
rs429358	APOE*4	0.002	0.023	0.919	-0.001	0.023	0.973	0.004	0.010	0.676			
rs75932628	TREM2	0.276	0.221	0.212	-0.016	0.221	0.942	-0.050	0.093	0.591			
rs6656401	CR1	-0.003	0.022	0.893	-0.004	0.022	0.846	-0.012	0.009	0.187			
rs11218343	SORL1	-0.001	0.040	0.978	-0.045	0.040	0.265	-0.021	0.017	0.208			
rs10838725	CELF1	-0.005	0.018	0.765	0.036	0.018	0.041	0.008	0.007	0.275			
rs983392	MS4A6A	-0.004	0.017	0.815	0.012	0.017	0.460	0.002	0.007	0.803			
rs10792832	PICALM	0.036	0.017	0.030	-0.007	0.017	0.667	-0.003	0.007	0.668			
rs17125944	FERMT2	0.009	0.027	0.723	-0.047	0.027	0.084	0.001	0.011	0.901			
rs10498633	SLC24A4-RIN3	-0.016	0.019	0.399	0.006	0.020	0.774	0.004	0.008	0.636			
rs4147929	ABCA7	0.023	0.022	0.285	0.010	0.022	0.664	0.006	0.009	0.488			
rs6733839	BIN1	0.008	0.017	0.621	-0.039	0.017	0.022	-0.002	0.007	0.814			
rs35349669	INPP5D	0.015	0.017	0.380	-0.016	0.017	0.328	-0.010	0.007	0.132			
rs7274581	CASS4	-0.012	0.030	0.676	-0.012	0.030	0.702	0.021	0.012	0.098			
rs190982	MEF2C	0.013	0.017	0.431	1.67x10 ⁻⁴	0.017	0.992	-0.011	0.007	0.127			
rs10948363	CD2AP	-0.013	0.018	0.472	0.031	0.018	0.094	1.87x10 ⁻⁴	0.008	0.980			
rs1476679	ZCWPW1	-0.012	0.017	0.487	0.001	0.018	0.948	-0.005	0.007	0.513			
rs11771145	EPHA1	-0.024	0.017	0.157	-0.012	0.017	0.502	0.003	0.007	0.638			
rs2718058	NME8	0.009	0.017	0.608	-0.029	0.017	0.089	-0.001	0.007	0.912			
rs28834970	PTK2B	-0.029	0.017	0.082	0.018	0.017	0.274	0.007	0.007	0.288			
rs9331896	CLU	0.034	0.017	0.051	-0.006	0.018	0.749	0.004	0.007	0.597			

Note: Multiple testing correction by Bonferroni 0.05/ (5 phenotypes x 20 variants); $P < 5 \times 10^{-4}$ was considered significant

Abbreviations: MRI $^{\sim}$ Magnetic resonance imaging, SNP $^{\sim}$ Single nucleotide polymorphism, β $^{\sim}$ regression coefficient, SE $^{\sim}$ Standard error

References

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- [3] International Genomics of Alzheimer's Disease C. Convergent genetic and expression data implicate immunity in Alzheimer's disease. Alzheimers Dement. 2015;11:658-71.
- [4] Guerreiro R, Bras J, Hardy J. SnapShot: genetics of Alzheimer's disease. Cell. 2013;155:968- e1.

Table 1: Cohort characteristics

Characteristics	Rotterdam Study
AD data set	
Total	8893
Late-onset AD	1270
AD free controls	84.30 (6.8)
Age-of-onset (SD)	5228 (59%)
Females (%)	
MCI data set	
Total	3605
MCI cases	360
Controls	71.9 (7.2)
Age (SD)	2063 (57%)
Females (%)	
MRI data set	
Total	4527
Age (SD)	64.74 (10.8)
Females (%)	2516 (56%)

Abbreviation: SD ~ Standard deviation, AD ~ Alzheimer's disease, MCI ~ Mild cognitive impairment, MRI ~ Magnetic resonance imaging

Table 2: Results of association of AD with risk scores GRS

		Including A	Excluding APOE					
SNP Cluster*	β SE		P-value	β	SE	P-value		
GRS1 (Combined)	0.73	0.040	6.53x10 ⁻⁷⁴	0.69	0.101	1.12x10 ⁻¹¹		
Immune response	-	-	-	0.69	0.166	3.20x10 ⁻⁵		
Endocytosis	-	-	-	0.75	0.171	1.28x10 ⁻⁵		
Cholesterol Transport	0.71	0.042	3.22x10 ⁻⁶⁴	0.39	0.219	0.077		
Hematopoietic cell lineage [†]	0.73	0.042	5.16x10 ⁻⁶⁶	-	-	-		
Hemostasis			-	0.50	0.292	0.090		
Clathrin/AP2 Adaptor complex	0.72	0.042	4.68x10 ⁻⁶⁵	0.50	0.236	0.036		
Protein folding [†]	0.72	0.042	2.96x10 ⁻⁶⁴	-	-	-		

Abbreviations: GRS1 $^{\sim}$ Combined genetic risk score, SNP $^{\sim}$ Single nucleotide polymorphism, β $^{\sim}$ Regression coefficient, SE $^{\sim}$ Standard error.

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 $[\]ensuremath{^*}$ Logistic regression model adjusted for age and sex in RS (N=1270 cases)

[†]Only one SNP available in excluding APOE GRS2

Table 3: Results of association of MCI with risk scores GRS

		Including A	POE		Excluding APOE				
SNP Cluster*	β	SE	P-value	β	SE	P-value			
GRS1 (combined)	0.19	0.075	0.012 0.59 0.179		9.51x10 ⁻⁴				
Immune response	-	-	-	0.46	0.295	0.116			
Endocytosis	-	-	-	1.16	0.305	1.44x10 ⁻⁴			
Cholesterol Transport	0.11	0.082	0.164	0.39	0.392	0.322			
Hematopoietic cell lineage [†]	0.09	0.084	0.269	-	-	-			
Hemostasis	-	-	-	-0.08	0.524	0.872			
Clathrin/AP2 Adaptor complex	0.12	0.082	0.128	0.72	0.423	0.089			
Protein folding [†]	0.10	0.083	0.218	-	-	-			

Note: Multiple testing correction by Bonferroni 0.05/ (5 phenotypes x 11 risk scores); P < 9.09x10-4 was considered significant

Abbreviations: GRS1 $^{\sim}$ Combined genetic risk score, SNP $^{\sim}$ Single nucleotide polymorphism, $\beta \sim$ Regression coefficient, SE $^{\sim}$ Standard error.

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^{*} Logistic regression model adjusted for age and sex in RS (N=360 cases)

[†] Only one SNP available in excluding APOE pathway based GRS2

Table 4: Results for association of <u>risk scores-GRS</u> with MRI phenotypes

Including APOE									Excluding APOE									
SNP cluster *	White matter lesions Hippocampal volume			Brai	in volu	ıme	White matter lesions Hippocampal volume				Bra	Brain volume						
	β	SE	Р	β	SE	P	β	SE	P	β	SE	P	β	SE	P	β	SE	P
GRS1 (combined)	0.012	0.016	0.448	-0.001	0.016	0.929	0.002	0.007	0.806	0.059	0.037	0.114	-0.009	0.038	0.810	-0.006	0.016	0.724
Immune response	-	-	-	-	-	-	-	-	-	0.149	0.062	0.016	-0.024	0.062	0.706	-0.010	0.026	0.692
Endocytosis	-	-	-	-	-	-	-	-	-	0.071	0.062	0.254	-0.046	0.063	0.462	0.004	0.026	0.865
Cholesterol Transport	0.005	0.017	0.785	0.001	0.017	0.964	0.004	0.007	0.574	0.063	0.080	0.434	0.013	0.080	0.875	0.023	0.033	0.497
Hematopoietic cell lineage [†]	0.002	0.017	0.901	0.001	0.017	0.976	0.004	0.007	0.556	-	-	-	-	-	-	-	-	-
Hemostasis	-	-	-	-	-	-	-	-	-	0.228	0.108	0.034	-0.077	0.109	0.479	-0.009	0.045	0.835
Clathrin/AP2 Adaptor complex	0.011	0.017	0.507	-0.002	0.017	0.924	0.003	0.007	0.658	0.258	0.088	3.55x10 ⁻³	-0.077	0.109	0.479	-0.009	0.045	0.835
Protein folding [†]	0.007	0.017	0.700	-0.001	0.017	0.970	0.004	0.007	0.619	-	-	-	-	-	-	-	-	-

Note: Multiple testing correction by Bonferroni 0.05/ (5 phenotypes x 11 risk scores); P < 9.09x 10^{-4} was considered significan

Abbreviations: GRS1 $^{\sim}$ Combined genetic risk score, MRI $^{\sim}$ Magnetic resonance imaging, SNP $^{\sim}$ Single nucleotide polymorphism, β $^{\sim}$ Regression coefficient, SE $^{\sim}$ Standard error, P $^{\sim}$ P-value

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^{*} Linear regression model with MRI phenotype as outcome and <u>risk score-GRS</u> as predictor, adjusted for age at MRI scan, sex in RS (N=4527)

[†] Only one SNP available in excluding APOE pathway based GRS2