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Exploiting the Anti-HIV-1 Activity of Acyclovir: Suppression of Primary and Drug-Resistant HIV Isolates and Potentiation of the Activity by Ribavirin

Christophe Vanpouille,a Andrea Lisco,abcd Andrea Introini,a Jean-Charles Grivel,a Arshi Munawwar,b Melanie Merbah,a Raymond F. Schinazi,c Marco Derudas,d Christopher McGuigan,d Jan Balzarini,e and Leonid Margolis,a

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Multiple clinical trials have demonstrated that herpes simplex virus 2 (HSV-2) suppressive therapy using acyclovir (ACV) or valacyclovir in HIV-1/HSV-2-infected persons increased the patient’s survival and decreased the HIV-1 load. It has been shown that the incorporation of ACV-monophosphate into the nascent DNA chain instead of dGMP results in the termination of viral DNA elongation and directly inhibits laboratory strains of HIV-1. We evaluated here the anti-HIV activity of ACV against primary HIV-1 isolates of different clades and coreceptor specificity and against viral isolates resistant to currently used drugs, including zidovudine, lamivudine, nevirapine, a combination of nucleoside reverse transcriptase inhibitors (NRTIs), a fusion inhibitor, and two protease inhibitors. We found that, at clinically relevant concentrations, ACV inhibits the replication of these isolates in human tissues infected ex vivo. Moreover, addition of ribavirin, an antiviral capable of depleting the pool of intracellular dGTP, potentiated the ACV-mediated HIV-1 suppression. These data warrant further clinical investigations of the benefits of using inexpensive and safe ACV alone or in combination with other drugs against HIV-1, especially to complement or delay highly active antiretroviral therapy (HAART) initiation in low-resource settings.

Compelling clinical evidence has demonstrated that the treatment of HIV-1-infected persons with acyclovir (ACV) or its prodrug valacyclovir (vACV) reduces HIV-1 load and delays HIV-1 disease progression (20). In particular, ACV/vACV treatments are associated with a reduction of HIV-1 load in plasma, semen, cervico-vaginal secretions, and rectal swabs (3, 9, 11, 20, 27, 45, 46) and in plasma during pregnancy (10). Also, several clinical studies, as well as two meta-analyses of a total of 15 randomized clinical studies, demonstrate that ACV/vACV treatment delays progression to AIDS and prolongs the patient’s survival (14, 22).

Understanding the mechanisms of ACV/vACV anti-HIV-1 activity is crucial to assessing its potential clinical use. It is hypothesized that ACV/vACV, an acyclic guanosine analogue, reduces HIV-1 load indirectly by suppressing HSV-2-mediated inflammation (41). However, we and others have shown that upon phosphorylation by kinases expressed by coinfecting human herpesviruses (HHVs) (including, but not limited to, herpes simplex virus 2 [HSV-2]), ACV directly inhibits HIV-1 reverse transcriptase (RT) (21, 25). These findings suggest that the anti-HIV activity of ACV may not be restricted to subjects coinfected with HSV-2, as other HHVs or yet unknown host enzymes (24) are also able to phosphorylate ACV. In consideration of its wide availability, safety, and low cost compared to other antiretroviral agents, ACV/vACV could be utilized to complement or delay highly active antiretroviral therapy (HAART) initiation in low-resource settings.

Here, we further investigate the antiviral activity of ACV against HIV-1 in experiments designed to mimic clinically relevant situations: (i) we evaluated its anti-HIV-1 activity not only against laboratory HIV-1 strains but also against primary HIV-1 clinical isolates of different subtypes and against multidrug-resistant variants; (ii) we evaluated the effect of ribavirin, a drug that is used against hepatitis C virus (HCV), on the anti-HIV-1 activity of ACV, since ribavirin is known to deplete the pool of the intracellular counterpart of ACV-triphosphate (ACV-TP), dGTP.

METHODS AND MATERIALS

Cultures and viral stocks. Tonsillar tissues obtained from the Children’s National Medical Center (Washington, DC), in accordance with an IRB-approved protocol, were dissected and cultured as described elsewhere (13). The MT-4 T-cell line was obtained from ATCC and maintained in RPMI with 10% fetal calf serum (FCS). The analysis of tonsillar tissue samples from 38 different donors showed that all of them contain HHVs of different types (21).

Virus stocks. HIV-1_LAI04 viral stocks were obtained from the Rush University Virology Quality Assurance Laboratory (Chicago, IL). Primary isolate viruses (HIV-1_AUS931, HIV-1_AUS936, HIV-1_AUS952, and HIV-1_AUS97), multi-nucleoside reverse transcriptase inhibitor (NRTI)-resistant viruses (HIV-1_3246, HIV-1_1020, HIV-1_1406, HIV-1_17303-3, HIV-1_1020, HIV-1_1406, and HIV-1_17303-3), nevirapine-resistant virus (HIV-1_N119), fusion inhibitor-resistant virus (HIV-1_N2A3, gp41(M136/538A/N842)), and protease inhibitor-resistant virus (HIV-1_L10R/N46K/L63P/V82T/I84V) and

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TABLE 1 Inhibition of HIV-1 replication by ACV in tonsillar tissues ex vivo

<table>
<thead>
<tr>
<th>Strain</th>
<th>HIV replication</th>
<th>ACV</th>
<th>% Inhibition by ACV</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Untreated control</td>
<td>3 μM</td>
<td>30 μM</td>
</tr>
<tr>
<td>Primary HIV-1 isolates</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>HIV-1LAI.04</td>
<td>13,052 (4,508–19,484)</td>
<td>7,714 (1,487–12,305)</td>
<td>749 (267–3,238)</td>
</tr>
<tr>
<td>HIV-1pNL4-3 gp41(36G)/V38A/N42D</td>
<td>4,455 (566–27,716)</td>
<td>3,836 (608–16,167)</td>
<td>871 (175–2,031)</td>
</tr>
<tr>
<td>HIV-1LAI.04 gp41(36G)/V38A/N42D</td>
<td>14,950 (4,373–71,055)</td>
<td>12,926 (3,568–46,592)</td>
<td>7,438 (1,404–24,983)</td>
</tr>
<tr>
<td>HIV-1LAI.04 gp41(36G)/V38A/N42D</td>
<td>5,365 (3,351–10,700)</td>
<td>4,832 (2,017–10,142)</td>
<td>2,059 (596–5,403)</td>
</tr>
<tr>
<td>HIV-1LAI.04 gp41(36G)/V38A/N42D</td>
<td>4,462 (2,166–6,919)</td>
<td>2,263 (1,137–3,287)</td>
<td>261 (215–336)</td>
</tr>
<tr>
<td>AZT-resistant variant HIV-1AZT.4X</td>
<td>18,006 (15,618–30,906)</td>
<td>10,325 (9,033–15,529)</td>
<td>6,946 (6,573–9,967)</td>
</tr>
<tr>
<td>Lamivudine-resistant variant HIV-1M184V</td>
<td>33,750 (27,305–44,612)</td>
<td>37,165 (28,921–43,976)</td>
<td>10,103 (8,118–14,408)</td>
</tr>
<tr>
<td>Multi-NRTI-resistant variants</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>HIV-1SI234-4</td>
<td>1,904 (752–4,139)</td>
<td>791 (203–1,782)</td>
<td>67 (0–598)</td>
</tr>
<tr>
<td>HIV-119076-4</td>
<td>4,664 (513–11,393)</td>
<td>1,415 (360–13,650)</td>
<td>29 (13–627)</td>
</tr>
<tr>
<td>HIV-14653–13</td>
<td>793 (577–16,866)</td>
<td>782 (550–17,295)</td>
<td>33 (17–3,359)</td>
</tr>
<tr>
<td>HIV-12303–3</td>
<td>3,389 (1,940–10,550)</td>
<td>2,524 (867–7,047)</td>
<td>1,054 (253–4,206)</td>
</tr>
<tr>
<td>HIV-11617–1</td>
<td>12,559 (1,693–34,307)</td>
<td>9,125 (2104–23,525)</td>
<td>1,861 (216–4,273)</td>
</tr>
<tr>
<td>HIV-133764–2</td>
<td>933 (818–9,311)</td>
<td>295 (211–3,349)</td>
<td>98 (91–302)</td>
</tr>
<tr>
<td>Nevirapine-resistant variant HIV-1M119</td>
<td>52,832 (18,027–139,405)</td>
<td>8,019 (6,935–29,579)</td>
<td>16,599 (6,152–29,579)</td>
</tr>
<tr>
<td>Fusion inhibitor-resistant variant HIV-1</td>
<td>1,043 (778–2,881)</td>
<td>619 (600–1,174)</td>
<td>48 (44–56)</td>
</tr>
<tr>
<td>Protease inhibitor-resistant HIV-1 variants</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>HIV-1LAI04/M46I/L63P/V82T/I84V</td>
<td>23,863 (18,986–31,508)</td>
<td>12,590 (11,524–15,546)</td>
<td>902 (614–1,148)</td>
</tr>
<tr>
<td>HIV-1M184V/L63P/V82T/I84V</td>
<td>4,410 (287–10,834)</td>
<td>3,685 (558–8,507)</td>
<td>348 (241–1,095)</td>
</tr>
</tbody>
</table>

*For each experimental condition, 27 tonsillar tissue blocks were inoculated with each HIV-1 isolate from a multi-NRTI-resistant HIV-1 panel and treated for 12 days with 3 or 30 μM ACV.

RESULTS

ACV suppresses replication of primary HIV-1 isolates. Previously, we reported that ACV suppresses replication of four laboratory HIV-1 strains, CXC4-tropic LAL04 (HIV-1LAI04), CCR5-tropic Bal, SF162, and ADB, in ex vivo infected human lymphoid tissues (21). Here, we evaluated the susceptibility of four primary isolates to ACV: HIV-1MS69SN20, HIV-197G520, and clade C, HIV-1MS69SN20 of clade A, and HIV-1M119 of clade B. Isolate HIV-1MS69SN20 uses CCR3, CCR5, and CCR4; HIV-197G520 uses CCR3, CCR5, and CCR4; HIV-1LA04 uses CCR3 and CCR5.

Human lymphoid tissues were inoculated with each of these
isolates and then treated for the entire culture period with ACV at 3 or 30 μM. These concentrations were chosen on the basis of our previous work with this compound (21). The median cumulative release of p24gag in the culture supernatants of tissues infected with each primary HIV-1 isolate with or without ACV is presented in Table 1. ACV suppresses replication of primary isolate HIV-197USNG30, HIV-196USNN20, and HIV-1 ME1, and this suppression was not significantly different from that of laboratory isolate HIV-1LAI.04 at both 3 and 30 μM ACV (P < 0.05) (Fig. 1). At 3 μM, the inhibition of HIV-1 96USNG31 replication was lower than that of the laboratory strain HIV-1LAI.04 (P < 0.03). However, at 30 μM, the ACV inhibitory activity was not statistically different for the two viruses (P > 0.05).

In summary, we found that primary isolates from clade A and B were inhibited by ACV as efficiently as the laboratory isolate HIV-1LAI.04. However, one of the two clade C HIV-1 subtypes tested had lower susceptibility to ACV compared to HIV-1LAI.04. ACV suppresses NRTI-resistant HIV-1 isolates. We assessed the ACV susceptibility of HIV-1 variants that are resistant to one or more licensed NRTIs. First, we investigated the ACV anti-HIV activity against the AZT-resistant isolate AZT.4X (characterized by the D67N, K70R, T215Y, and K219Q mutations in RT) in ex vivo lymphoid tissues. ACV at concentrations of 3 and 30 μM inhibited AZT.4X replication by 57.2% and 73.7% respectively. No statistical difference between the inhibition of AZT.4X and the inhibition of the laboratory strain HIV-1LAI.04 was observed (P > 0.05) (Table 1).

![FIG 1](http://aac.asm.org) Inhibition of HIV-1 replication by ACV in human lymphoid tissues. Tissue blocks (27 from each donor) were infected with HIV-1 97USNG31, HIV-1 96USNN20, HIV-1 97USNG30, or HIV-1 ME1 and treated with 3 or 30 μM ACV. Replication of HIV-1 was evaluated by p24gag core antigen release in pooled medium bathing 27 tissue blocks using a bead-based assay. Each point represents the measurement of medium pooled from three wells, each of which contained nine tissue blocks. The anti-HIV activity of ACV was evaluated by comparing viral replication in drug-treated tissues with that in untreated donor-matched control tissues. The graph represents the typical replication kinetics of 4 to 6 experiments performed with tissues from different donors. For average replication values, see Table 1.

### TABLE 2 Potentiation of the ACV anti-HIV-1 activity by ribavirin in human tonsillar tissues

<table>
<thead>
<tr>
<th>Condition</th>
<th>ACV concn (μM)</th>
<th>HIV-1 replication [median p24gag (IQR)]b</th>
<th>% inhibitionc</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ribavirin (10 μM)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0</td>
<td>7,379 (4,135–10,737)</td>
<td>NA</td>
<td></td>
</tr>
<tr>
<td>0.3</td>
<td>5,020 (3,220–8,837)</td>
<td>18.3 ± 16.8</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>2,284 (1,426–4,497)</td>
<td>65.9 ± 17.0</td>
<td></td>
</tr>
<tr>
<td>10</td>
<td>1,327 (1,098–1,807)</td>
<td>91.8 ± 7.5**</td>
<td></td>
</tr>
<tr>
<td>No ribavirin</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0</td>
<td>8,228 (3,714–12,357)</td>
<td>NA</td>
<td></td>
</tr>
<tr>
<td>0.3</td>
<td>8,528 (5,844–12,112)</td>
<td>6.1 ± 11.7</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>5,869 (3,554–8,257)</td>
<td>41.3 ± 25.2</td>
<td></td>
</tr>
<tr>
<td>10</td>
<td>3,234 (1,554–5,489)</td>
<td>65.2 ± 17.6</td>
<td></td>
</tr>
</tbody>
</table>

a For each condition, 27 tonsillar tissue blocks from each of n donors were inoculated with X4_LAI.04 and treated for a 12-day period with 1, 3, or 10 μM ACV in the absence or presence of 10 μM ribavirin.

b Median cumulative p24gag release (pg/ml) in culture supernatant of infected tissues. n = 10 for all data points except for 10 μM ACV, where n = 6. IQR, interquartile range.

c Percentage (mean ± SD) of inhibition of HIV replication is defined as [1 – (R_{ACV}/R_{Ctl})] × 100, where R_{ACV} and R_{Ctl} are the amounts of p24gag accumulated in the medium over a 12-day culture period in ACV-treated cultures and in donor-matched untreated ACV cultures, respectively. * P value of < 0.05 for HIV-1LAI.04 inhibition in the absence versus in the presence of 10 μM ribavirin at 3 μM ACV. ** P value of < 0.05 for HIV-1LAI.04 inhibition in the absence versus in the presence of 10 μM ribavirin at 30 μM ACV. NA, not available.
ACV suppresses NNRTI-resistant HIV-1 isolates. Furthermore, we evaluated the ACV-mediated suppression of HIV-1N119 in human lymphoid tissue ex vivo. HIV-1N119 has the RT mutation Y181C, which confers resistance to nevirapine, as well as to related nonnucleoside reverse transcriptase inhibitors (NNRTIs). HIV-1N119 replication was suppressed by 54.9% ± 35.9% and by 83.9% ± 6.1% at 3 and 30 μM ACV, respectively, rates similar to that of HIV-1LAI.04 (P > 0.05) (Table 1).

ACV suppresses HIV-1 isolates resistant to protease and fusion inhibitors. We tested the ACV susceptibility of HIV-1 isolates resistant to a fusion inhibitor (HIV-1pNL4-3 gp41(36G)/V36A/N255K) or two protease inhibitors (HIV-1L10R/M46I/L63P/V82T/I84V and HIV-1M46I/L63P/V82T/I84V). HIV-1pNL4-3 gp41(36G)/V36A/N255K is a recombinant virus that contains one or more amino acid substitutions in gp41 which confer resistance to enfuvirtide (T-20). Both HIV-1L10R/M46I/L63P/V82T/I84V and HIV-1M46I/L63P/V82T/I84V are resistant to the structurally diverse protease inhibitors MK-639, XM323, A-80897, Ro31-8959, VX-478, and SC-52151 (7).

We did not observe any difference in ACV-mediated inhibition of HIV-1LAI.04, HIV-1pNL4-3, gp41(36G)/V36A, N255K, HIV-1L10R/M46I/L63P/V82T/I84V, and HIV-1M46I/L63P/V82T/I84V at either 3 or 30 μM ACV (P > 0.05, Table 1). Thus, both fusion and protease inhibitor-resistant HIV-1 variants tested here retained susceptibility to ACV at a level similar to that of the wild-type HIV-1 (HIV-1LAI.04).

Ribavirin potentiates the anti-HIV-1 activity of ACV and of ACV prodrug derivatives. We investigated in T-cell lines and in ex vivo human lymphoid tissue whether ribavirin, an antiviral compound known to deplete the intracellular dGTP pool, potentiates the anti-HIV-1 activity of a dGTP antagonist, ACV. Control experiments did not show a statistically significant difference between the cumulative p24 gag production in HIV-1LAI.04-infected tissues treated and that in tissues not treated with ribavirin (P > 0.05) (Table 2).

Next, in human lymphoid tissue ex vivo, we evaluated the susceptibility of HIV-1 replication to the combination of ribavirin and ACV. Without ribavirin, 3 μM ACV suppressed HIV-1LAI.04 replication by 41.3% ± 8.0%, while in the presence of ribavirin, HIV-1LAI.04 replication was inhibited by 65.9% ± 5.4% (P = 0.008). Similarly, in the absence of ribavirin, 10 μM ACV suppressed HIV-1LAI.04 replication by 65.2% ± 5.6%, while in the presence of ribavirin, HIV-1LAI.04 replication was inhibited by 91.8% ± 2.4% (P = 0.025). Also, 10 μM ribavirin potentiated the anti-HIV-1 activity of 0.3 μM ACV (18.3% ± 5.3% versus 6.1% ± 3.7% inhibition in the absence and presence of ribavirin, respectively) (Fig. 3). However, this potentiation did not reach statistical significance (P > 0.05).

Also, we evaluated the effect of ribavirin on the suppressive activity of ACV in MT-4 cells. Unlike human tissues, which contain various HHVs that activate (phosphorylate) ACV into ACV-MP, MT-4 cells are HHV free; therefore, in these experiments we used ACV-ProTides, which are lipophilic phosphorylated ACV prodrug derivatives (42).

For each experimental condition, 10^5 MT-4 cells infected with HIV-1LAI.04 were treated with ACV ProTides CI2648 or CI2681 at

*FIG 2 Inhibition of replication of multidrug-resistant HIV-1 clones by ACV in human lymphoid tissues. Donor-matched sets of tissue (27 blocks for each experimental condition from each of three to seven donors) were incubated with ACV (3 or 30 μM) for 12 days or used as untreated controls. Some of these sets were infected with multi-NRTI-resistant clones HIV-1J324–4, HIV-1H343–6, HIV-1A110–6, HIV-1J100–5, HIV-1J346–5, HIV-1F37–4, HIV-1A110–6, and HIV-1H343–6, or with the laboratory strain HIV-1LAI.04. Replication of HIV-1 was evaluated by p24 gag core antigen release in pooled medium bathing 27 tissue blocks using a bead-based assay. The anti-HIV activity of ACV, as a percentage, was evaluated by comparing viral replication in drug-treated tissues with that in untreated donor-matched control tissues. Presented are means ± SEM of HIV-1 inhibition in 27 human tissue blocks from each of three to seven donors, relative to results for matched untreated tissues (n = 3 for HIV-1J324–4, and HIV-1H343–6, n = 4 for HIV-1A110–6, n = 7 for HIV-1H0076–4, HIV-1J303–3, HIV-1J35764–2, and HIV-1LAI.04).
concentrations varying from 0.2 to 50 μM in the absence of ribavirin. When cells were treated with ribavirin at concentrations ranging from 1 to 20 μM, ACV ProTides CI2648 or CI2681 was added at concentrations varying from 0.01 to 20 μM. For both ProTides, ribavirin markedly decreased the 50% effective concentration (EC50) of ACV ProTides at concentrations ranging from 0.01 to 50 μM. For both ACV ProTides, ribavirin markedly decreased the EC50 from 1.3 to 0.018 μM for CI2648 and from 9 to 1.8 μM for CI2681 (Fig. 4).

**DISCUSSION**

The growing evidence of the important role played by sexually transmitted infections in general and by HSV-2 in particular in HIV transmission and pathogenesis has led to the development of new anti-HIV-1 strategies aimed at suppressing coinfected viruses. Highly specific and potent anti-herpetic drugs were developed in the mid-1970s, making it possible to test these strategies in randomized clinical trials of HSV-2 suppressive therapy using ACV/vACV. These trials consistently demonstrated that such therapy resulted in increased survival and a decrease in HIV-1 load by 0.3 to 0.5 log10 in plasma, semen, cervico-vaginal secrections, and rectal swabs (3, 9, 11, 20, 27, 45, 46) and by 0.5 log10 in plasma during pregnancy (10). Although this effect is relatively small compared to that produced by the current potent anti-HIV cocktails, it is important to remember that the effect achieved by ACV monotherapy is similar to the monotherapy effect of zidovudine, which is one of the most efficient components of HAART (17). Moreover, two studies have recently reported that HSV-2 suppressive therapy using ACV (400 mg twice daily [b.i.d.]) delayed HIV-1 disease progression by 16% (20) and 27% (30). This delay may seem modest, but given HSV-2 seroprevalence rates as high as 90% in HIV-infected individuals in sub-Saharan Africa and the limited resources in parts of this region, low-cost interventions with well-tolerated drugs which can slow disease progression are an appealing strategy to delay initiation of HAART or the onset of AIDS in coinfected individuals (6, 38). Deciphering the mechanisms by which ACV suppresses HIV-1 is therefore important both for basic virology and for optimization of anti-HIV-1 therapeutic strategies.

Earlier, we and others (21, 25) showed that ACV, upon metabolic activation, is a direct inhibitor of HIV-1. In particular, we found that human tissues, including the tonsil tissues used in these studies, carry HHVs (not necessarily HSV-2) (21). Furthermore, when these tissues are infected ex vivo with HIV-1, ACV is phosphorylated by HHV-encoded kinases (and/or possibly by yet unidentified host kinases) (24) and acts as an NRTI, directly inhibiting HIV-1 RT (21).

In the studies referenced above, the anti-HIV-1 activity of ACV was tested for well-defined laboratory-adapted strains of HIV-1, which may be different from primary isolates in several important aspects. In the current work, we tested the anti-HIV activity of ACV against primary isolates and drug-resistant isolates that commonly emerge in treated patients. We performed these tests in an ex vivo system of human tissues that reflects many in vivo tissue features.

We found that ACV inhibited all primary and multidrug-resistant HIV-1 variants tested. Moreover, the understanding of the molecular mechanism of ACV-mediated HIV-1 suppression allowed us to potentiate ACV suppressive activity by adding ribavirin, a known antiviral used in HCV treatment, which is capable of depleting the pool of intracellular dGTP (8).

We first tested the susceptibility of four clinical HIV-1 isolates to two concentrations of ACV, 3 and 30 μM, in ex vivo lymphoid tissue. These concentrations of ACV were chosen because they are in the range of those achieved in patients treated with oral ACV/vACV (23, 34) and they have been reported to inhibit the replication of HIV-1 laboratory isolates by approximately 50 and 90%, respectively (21). ACV suppressed the replication of all tested HIV-1 primary isolates, including two of clade C, which is prevalent in Southern and East Africa and India and is responsible for nearly half of all HIV infections worldwide, one of clade A, mainly present in Western/Central Africa and Russia, and one of clade B, commonly found in Europe, the Americas, and Australia. Also, the inhibition was irrespective of HIV-1 coreceptor specificity (CCR5, CXCR4, CCR3, CCR2B, Bob, Bonzo). In general, there...
was no difference in suppression of viral replication in \textit{ex vivo} tonsillar tissues between clinical HIV-1 isolates and laboratory strains, although one of the two clade C isolates (HIV-1\textsubscript{96USA52NCa1}) was not as susceptible to the low concentration of ACV as the laboratory isolate HIV-1\textsubscript{1LAI04}.

This result should be taken into consideration in the interpretation of the current or recently completed clinical trials on the anti-HIV use of ACV. In fact, although average delays of 16% and 27% of disease progression were observed (20, 30), variations in the response to ACV, which might be explained by differences in susceptibilities of particular HIV-1 isolates, especially to low-dose ACV (i.e., 400 mg b.i.d.), were noted. In this regard, the higher doses of ACV in two ongoing clinical trials (43; NCT101859084 [http://clinicaltrials.gov/ct2/show/NCT01059084]
(\textsuperscript{[1]}) (vACV, 500 mg b.i.d. or 1,000 mg thrice daily) may reduce the variability stemming from the intrinsic differences in susceptibility of HIV-1 RT of different isolates, as observed in our experiments in \textit{ex vivo} tissues treated with a higher concentration of ACV (30 \textmu M). Furthermore, it has been recently reported that, among HIV-1/HSV-2-coinfected persons, vACV suppressive therapy results in greater reduction in plasma HIV-1 levels than standard-dose ACV suppression (26).

For the future use of ACV or its derivatives in anti-HIV-1 therapy, it is important that ACV remains active against common drug-resistant HIV-1 variants that evolve in the course of regular anti-HIV therapy. Indeed, in the United States, up to 50% of patients receiving antiretroviral therapy are infected with viruses resistant to at least one of the available antiretroviral drugs. Moreover, mutations conferring resistance to one drug may often confer resistance to another drug. Furthermore, a significant proportion of new HIV infections results from the transmission of strains that are already resistant to one or more antiretroviral drugs (33).

Here, we first tested ACV against HIV-1 variants that are resistant to some of the most common anti-HIV NRTIs, zidovudine (AZT) and lamivudine (3TC). We found that ACV efficiently suppressed replication of NRTI-resistant viruses. However, while ACV suppressed replication of AZT-resistant HIV-1 as efficiently as it suppressed the wild-type variant, 3TC resistance, conferred by the M184V mutation, decreased by \sim 4-fold the susceptibility to ACV.

Second, to further evaluate the ACV susceptibility of NRTI-resistant HIV-1 variants, we tested a panel of six prototypical insecticid-drug-resistant HIV-1 RT molecular clones. Each of the clones carries several mutations that occur most frequently in HIV-infected individuals treated with NRTIs. ACV (30 \textmu M) suppressed equally well the replication of the six multidrug-resistant clones and of HIV-1\textsubscript{1LAI04}. However, 3 \textmu M ACV (the EC\textsubscript{50} for HIV-1\textsubscript{1LAI04}) suppressed by 50% the replication of only four of the six multidrug-resistant clones, indicating that some combinations of the common NRTI mutations may provide a reduced susceptibility to low-dose ACV. Finally, none of the tested viruses (except an HIV-1 variant that carries a K65R RT mutation, which showed an \sim 2-fold decrease in susceptibility to 30 \textmu M ACV), including one virus carrying the V75I RT mutation, were found to be resistant to ACV. It was reported earlier that this mutation was selected under the pressure of ACV or its prodrg derivative in single-cell cultures of CD4\textsuperscript{+} lymphoblasts and the MT-4 cell line (25, 39). However, in \textit{ex vivo} tissues V75I-mutated virus was not resistant to ACV. Also, \textit{in vivo} ACV does not select for this mutation (2, 19). This further underscores the necessity to evaluate viral resistance in \textit{ex vivo} tissues, which represent the \textit{in vivo} more adequately than single-cell cultures. Nevertheless, it would be of interest to investigate which particular features determine the difference between tissues and single-cell cultures in viral resistance.

In resource-limited countries, the WHO recommends a first-line regimen of two NRTIs and one NNRTI. Zidovudine and tenofovir disoproxil fumarate are the preferred NRTIs in this regimen in combination with lamivudine or emtricitabine, while the NNRTI is often nevirapine. Currently, inhibitors of viral protease, integrase, or fusion are also important components of anti-HIV cocktails (37). Although cross-resistance generally occurs predominantly between different drugs of the same class, we tested ACV susceptibility to two protease inhibitors and to one fusion inhibitor in HIV-1 variants resistant to nevirapine. We found that these four resistant HIV-1 isolates were as susceptible to ACV as the wild-type reference laboratory isolate HIV-1\textsubscript{1LAI04}.

Finally, understanding of the molecular mechanism of ACV suppression of HIV-1 allowed us to potentiate its suppressive activity. Indeed, our earlier data demonstrated that termination of HIV-1 DNA elongation results from the incorporation of ACV-TP by HIV-1 RT into the nascent DNA chain instead of its natural counterpart dGTP (21). Therefore, increasing the intracellular ACV-TP/dGTP ratio should increase the probability for RT to incorporate ACV-TP rather than dGTP. This was previously demonstrated in a cell-free system where a 12-fold decrease in the concentration of dGTP resulted in an \sim 30% increase in ACV-TP inhibition of HIV-1 RT (21). In the present work, we used ribavirin to decrease the intracellular dGTP concentration and therefore increase the intracellular ACV-TP/dGTP ratio. Ribavirin inhibits IMP dehydrogenase, the enzyme that converts IMP to XMP, a key step in the \textit{de novo} biosynthesis of GTP and dGTP (8, 35). This strategy was used earlier to potentiate the antiviral activity of ACV, as well as those of ganciclovir and penciclovir, against herpesvirus and hepatitis B virus (8, 28, 44) and also of 2',3'-dideoxyinosine (ddl) and 2',3'-dideoxy-2,6-diaminopurine riboside (ddDAPR) against HIV (4, 5).

Here, we used ribavirin at 10 \textmu M, a concentration which \textit{per se} has no significant effect on HIV replication, and found that ribavirin significantly potentiated the anti-HIV-1 activity of ACV. The ribavirin-mediated potentiation of the ACV anti-HIV activity did not seem to be related to the efficiency of ACV phosphorylation. Indeed, a pronounced potentiation of the anti-HIV activity of ACV by ribavirin was observed both for ACV and for prephosphorylated ACV prodrg derivatives (ACV ProTides). Although both these strategies are based on the assumption that ribavirin acts by decreasing the dGTP pool, other effects of ribavirin cannot be excluded. For example, ribavirin’s carboxamidine group can make the native nucleoside drug resemble adenosine or guanosine, which therefore can be directly incorporated into DNA. Also, ribavirin is known to modulate host T-cell-mediated immunity against viral infection. Nevertheless, we think that the decrease of dGTP is the main mechanism by which ribavirin potentiates ACV anti-HIV activity.

The difference between \textit{ex vivo} tissues and isolated cells in potentiating these effects may be related to the size of the natural pool of dGTP. It is likely that this pool is larger in highly proliferating immortalized cell lines like MT-4 than in tissue cells.

In general, the fact that a drug that reduces the dGTP pool potentiates the anti-HIV activity of ACV emphasizes that ACV-TP and its natural counterpart dGTP compete for incorpo-
ration during DNA elongation by HIV-1 RT and confirms once again the existence of a direct effect of the activated ACV metabolite on HIV-1 RT.

The strategy of potentiating ACV anti-HIV activity may be clinically relevant, as the concentration of 10 μM ribavirin required for potentiation in our experiments is attained in human plasma upon oral dosing of ribavirin in HCV-infected patients (15). Our finding that ribavirin, an HCV drug, potentiates acyclovir, an anti-HSV drug recently reported to have anti-HIV-1 activity, is important in light of frequent coinfection with these three viruses: about 50% of HIV-1-infected patients are coinfected with HSV-2 in the United States (29), and about one quarter of the people infected with HIV are also infected with HCV (40). Our results show that in combination with ACV, ribavirin could have a dual-targeted action in coinfected individuals by inhibiting both HCV and HIV-1. Although both drugs are already used for therapy, only further studies may show whether this strategy could be pursued in vivo.

In summary, we found that in human lymphoid tissue, ACV suppresses the replication of HIV-1 of different clades and of different coreceptor tropisms as well as HIV-1 variants with mutations conferring resistance to the currently used NRTIs, NNRTIs, and inhibitors of viral protease, integrase, and fusion. Together with the reported absence of emergence of HIV variants resistant to ACV in vivo (2, 19) and the well-documented beneficial effect of ACV on disease progression (20), these data warrant further clinical investigation of the use of ACV alone or in combination with anti-HIV drugs against this virus. Moreover, this inexpensive and safe drug may be used to slow disease progression and delay HAART in low-income countries where access to HAART is limited. The combination of ACV with ribavirin may be envisioned also in topical microbicide applications where both HSV-2 and HIV-1 might be suppressed with this drug combination regimen. Further studies are needed to address the feasibility, the potential benefits, and the cost-effectiveness of novel anti-HIV-1 strategies based on ACV and its derivatives.

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REFERENCES


28. Nefts J, Andrei G, De Clercq E. 1998. The novel immunosuppressive agent mycophenolate mofetil markedly potentiates the antiviral effects...


