Reconstructing the Demographic History of Orang-utans using Approximate Bayesian Computation

Alexander Nater1, Maja P. Greminger1, Natasha Arora1, Carel P. van Schaik1, Benoit Goossens2,3,4, Ian Singleton5,6, Ernst J. Verschoor7, Kristin S. Warren8, and Michael Krützen1

(1) Anthropological Institute & Museum, University of Zurich, Zurich, Switzerland
(2) Organisms and Environment Division, School of Biosciences, Cardiff University, Cardiff, UK
(3) Danau Girang Field Centre, c/o Sabah Wildlife Department, Kota Kinabalu, Sabah, Malaysia
(4) Sabah Wildlife Department, Kota Kinabalu, Sabah, Malaysia
(5) Foundation for a Sustainable Ecosystem (YEL), Medan, Indonesia
(6) Sumatran Orangutan Conservation Programme (PanEco-YEL), Medan, Indonesia
(7) Department of Virology, Biomedical Primate Research Centre, Rijswijk, The Netherlands
(8) College of Veterinary Medicine, School of Veterinary and Life Sciences, Murdoch University, Perth, Australia

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Corresponding author:
Alexander Nater
Department of Evolutionary Biology
Evolutionary Biology Centre
Uppsala University
Norbyvägen 18D
752 36 Uppsala, Sweden
Phone number: +46 (0)18 471 2837
E-mail address: alexander.nater@ebc.uu.se
Abstract

Investigating how different evolutionary forces have shaped patterns of DNA variation within and among species requires detailed knowledge of their demographic history. Orang-utans, whose distribution is currently restricted to the Southeast Asian islands of Borneo (Pongo pygmaeus) and Sumatra (Pongo abelii), have likely experienced a complex demographic history, influenced by recurrent changes in climate and sea levels, volcanic activities and anthropogenic pressures. Using the most extensive sample set of wild orang-utans to date, we employed an approximate Bayesian computation (ABC) approach to test the fit of 12 different demographic scenarios to the observed patterns of variation in autosomal, X-chromosomal, mitochondrial and Y-chromosomal markers. In the best-fitting model, Sumatran orang-utans exhibit a deep split of populations north and south of Lake Toba, probably caused by multiple eruptions of the Toba volcano. In addition, we found signals for a strong decline in multiple Sumatran populations ~24 ka, probably associated with hunting by human colonizers. In contrast, Bornean orang-utans experienced a severe bottleneck ~135 ka, followed by a population expansion and substructuring starting ~82 ka, which we link to an expansion from a glacial refugium. Therefore, we showed that orang-utans went through drastic changes in population size and connectedness, caused by the recurrent contraction and expansion of rainforest habitat during Pleistocene glaciations, and probably also by the impact of hunting by early humans. Our findings also emphasize the fact that important aspects of the evolutionary past of species with complex demographic histories might remain obscured when applying overly simplified models.
Introduction

Patterns of DNA variation are the result of both adaptive and non-adaptive processes, and the debate about the relative importance of natural selection and random genetic drift in shaping genetic diversity within and among species is still ongoing (e.g. Hahn 2008; Nei et al. 2010). A common approach to detect signals of selection aims at identifying genomic regions that show marked deviations in DNA variation from a neutral equilibrium model (reviewed in Nielsen 2005). However, in certain demographic scenarios, such as population size changes or population subdivision, random genetic drift can result in similar deviations as selection (e.g. Excoffier et al. 2009; Teshima et al. 2006). Therefore, confounding effects of demographic processes can only be unravelled from selective signals if the demographic history is explicitly taken into account when formulating the expectations under the neutral model against which observed patterns of DNA variation are tested (e.g. Haddrill et al. 2005; Stajich & Hahn 2005). Consequently, methods to reconstruct the demographic history of natural populations have recently generated great interest among evolutionary geneticists, as recent technical advances allow conducting genome-wide studies of selection in a large variety of species (reviewed in Ellegren 2014).

Orang-utans, currently restricted to two distinct species on Borneo (Pongo pygmaeus) and northern Sumatra (Pongo abelii) (Wich et al. 2008), are the only Asian great apes and are phylogenetically most distant to humans (Groves 2001). Their ancestral position in the lineage leading to African great apes and modern humans has evoked great interest in this taxon in the overall effort to reconstruct the adaptive evolutionary history of great apes in general and humans in particular (Locke et al. 2011; Prado-Martinez et al. 2013). However, orang-utans might have experienced a complex demographic history, as their distribution has been subject to major changes during the Pleistocene. The ancestors of extant orang-utans have sequentially colonized the islands of the Sunda archipelago arriving from the Southeast Asian mainland (Delgado & Van Schaik 2000; Rijksen & Meijaard 1999). Since then, their population history was strongly influenced by geological and climatic events: rising and falling sea levels cyclically connected and isolated the islands of Sundaland, allowing for potential terrestrial migration between the islands at certain points in time (Voris 2000).

Major volcanic eruptions, mainly on Sumatra and Java, might have led to the extinction of local orang-utan populations and subsequent re-colonisations (Muir et al. 2000). Of special interest here is the Toba volcano on northern Sumatra, which has seen at least four major

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eruptions during the last 1.2 million years (Chesner et al. 1991). This sequence of eruptions culminated in the Toba supereruption ~73 ka, which is considered to be the most powerful volcanic eruption within the last 25 million years (Chesner et al. 1991) and is thought to have had severe consequences for flora and fauna on Sundaland (Williams et al. 2009). In the Late Pleistocene, all orang-utan populations on the mainland, southern Sumatra and Java went extinct (Delgado & Van Schaik 2000; Rijksen & Meijaard 1999). Climatic changes during the Pleistocene might have been responsible for the southward shift of the distribution and the disappearance of orang-utans from the mainland (Jablonski 1998). Moreover, anthropogenic factors, such as prehistoric hunting by hunter-gatherer societies, are likely to have played a significant role in the decline and extinction of orang-utans populations on insular Southeast Asia (Delgado & Van Schaik 2000).

Genetic signals of these past demographic changes have been found in studies of genetic diversity in extant orang-utan populations on Borneo and Sumatra. Most genetic studies analysing autosomal and mitochondrial DNA (mtDNA) agree that Sumatran orang-utans show a higher level of sequence diversity and corresponding long-term effective population size ($N_e$) (Locke et al. 2011; Muir et al. 2000; Prado-Martinez et al. 2013; Steiper 2006; Zhang et al. 2001), even though Sumatran orang-utans have a much smaller current census size and a more restricted distribution than Borneans (~6,600 vs. ~54,000 individuals, Wich et al. 2008). This large $N_e$ of the Sumatran species was interpreted as a signal of immigration from multiple differentiated populations into the current Sumatran gene pool (Muir et al. 2000; Steiper 2006). However, Y-chromosomal diversity in orang-utans shows the opposite pattern compared to mtDNA and autosomal data, with a smaller $N_e$ on Sumatra than Borneo (Nater et al. 2011). Such contrasting patterns of $N_e$ between species and among genomic regions hint at complex population dynamics that have so far not been properly investigated.

Recently, Locke et al. (2011) used extensive single-nucleotide polymorphism (SNP) data from whole-genome resequencing of five Bornean and five Sumatran orang-utans to model the demographic history of the two species. They found that a model with a population split ~400 ka with subsequent gene flow between Borneo and Sumatra fits the observed data best. Furthermore, Locke and colleagues inferred that Sumatran orang-utans underwent a continuous exponential population growth since the population split, while Bornean orang-utans were subject to a continuous exponential decline. Given the large amount of genetic data, the study by Locke and colleagues is currently regarded as the most accurate reconstruction of demographic history in orang-utans to date. However, the demographic

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modelling approach by Locke and colleagues did not take several idiosyncrasies of orang-utan biology into account, thus severely limiting the conclusions that could be drawn from their findings. First, it has been shown that biased sampling and disregard of population structure will produce misleading results regarding $N_e$ and its temporal changes (Chikhi et al. 2010; Stadler et al. 2009). The study by Locke and colleagues incorporated data from only five captive individuals each from Borneo and Sumatra without further provenance information. This limited genetic sampling is unlikely to represent the entire genetic diversity present on both islands. Second, given this lack of detailed sample provenance, the analyses were restricted to models that treated Bornean and Sumatran orang-utans as single panmictic populations each. Previous work, however, unequivocally showed that both Bornean and Sumatran orang-utans are deeply structured genetically (Arora et al. 2010; Nater et al. 2011; Warren et al. 2001). Especially on Sumatra populations north and south of Lake Toba exhibit high genetic differentiation (Nater et al. 2013; Nater et al. 2011). Third, Locke and colleagues did not test complex demographic models including population bottlenecks or recent declines, as suggested in previous genetic studies. For example, genetic signals of a bottleneck with subsequent population expansion on Borneo might be linked to a glacial refugium or the impact of the Toba supereruption ~73 ka (Arora et al. 2010; Steiper 2006), and patterns of a recent population decline in Sabah, Borneo, are most likely attributable to recent anthropogenic pressures (Goossens et al. 2006).

Reconstructing the demographic history of a species has long been hindered by the fact that full-likelihood methods were restricted to relatively simple demographic models (e.g. Hey & Nielsen 2004; Wilson et al. 2003), which might not capture all relevant processes in complex demographic settings. This restriction is mainly caused by the fact that the computation of the likelihood function of complex demographic models with many parameters is either intractable or computationally too expensive, especially for large data sets (Marjoram et al. 2003). Approximate Bayesian computation (ABC) allows circumventing these problems by approximating the likelihood functions with simulations of genetic data under a given demographic model (Beaumont et al. 2002; Marjoram et al. 2003). In order to estimate the model parameters, parameter values are drawn from predefined prior distributions and used to simulate genetic data matching the observed data in type of markers and number of loci. Both observed and simulated data are then reduced to a set of summary statistics and the Euclidian distance between the observed and the simulated summary statistics is calculated. Based on

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Here we present an ABC modelling approach of the demographic history of orang-utans based on autosomal and sex-linked marker systems. We aim to improve the current knowledge of demographic history by applying three major improvements over previous studies. First, we capitalize on the knowledge base of behavioural ecology and population genetics of orang-utans in order to test realistic demographic models. Second, due to our extensive set of orang-utan samples with detailed and reliable provenance, we are able to investigate models incorporating population substructure in both orang-utan species, which allows us to disentangle changes in population size from confounding effects due to changes in population structure. Third, by combining autosomal and sex-linked markers into a combined demographic analysis, we make use of the specific information content of different marker systems in this species with its heavily sex-biased dispersal. Due to strong female philopatry in orang-utans (Arora et al. 2012; Galdikas 1995; van Noordwijk et al. 2012), mitochondrial markers contain information about population split times without the confounding influence of gene flow. In contrast, Y-chromosomal loci should have more power than autosomal markers to reveal low levels of male-mediated gene flow.

**Materials & Methods**

**Sample Collection and Genetic Markers**

A representative sampling scheme covering the whole range of a species is crucial for accurate reconstruction of demographic history (Stadler et al. 2009). We used an extensive set of samples from wild-born orang-utans from ten sampling locations, covering the entire distribution of the genus (Figure 1, see Supporting Material for detailed information about sample origin). Samples were analysed for several genetic marker systems with different modes of inheritance and effective population sizes (Table 1), thus ensuring representation of both male and female population history, an important aspect in demographic reconstructions in species with strongly sex-biased dispersal (Nater et al. 2011; Nietlisbach et al. 2012).

The autosomal microsatellite data contained genotypes of 25 microsatellite markers from a total of 237 individuals (Arora et al. 2010; Greminger et al. 2014; Nater et al. 2013). We also included sequences from three mtDNA genes with a total length of 1,355 bp from 118
individuals (Nater et al. 2011), and Y-chromosomal haplotypes based on 11 Y-linked microsatellite loci from 129 individuals (Nater et al. 2011). We complemented the data set by additionally sequencing 8,055 bp of the non-coding X-chromosomal region Xq13.3 (Kaessmann et al. 2001) in 36 individuals and four non-coding autosomal regions (Fischer et al. 2006) of a total of 8,238 bp in 22 individuals. Basic summary statistics for all marker systems are provided in Table 2. The primers and cycling conditions used for PCR amplification and sequencing of the X-chromosomal and autosomal regions are described in the Supporting Table S1.

Approximate Bayesian Computation

Model Selection Procedure

We reconstructed the demographic history of orang-utans using an ABC approach implemented in the software package ABCtoolbox v1.1 (Wegmann et al. 2010). To achieve this goal, we first performed a model selection procedure, in which we used a hierarchical approach to test a total of 12 different demographic models (Figure 2) with increasing levels of complexity (see Supporting Tables S3 and S4 for more details about model parameterisation and prior distributions).

We started by testing four relatively simple models that assumed a single population for each of the two orang-utan species (Figure 2A). The first model in this set (I2) assumed constant population sizes and no migration between the two populations. The second model (IM2) incorporated asymmetric migration after the population split, up to a point in the past where migration between Borneo and Sumatra ceased. Gene flow in all models with migration was strictly male-mediated, as recent genetic and behavioural findings showed extreme female philopatric tendencies in orang-utans (Arora et al. 2012; Nater et al. 2011; van Noordwijk et al. 2012). The third model (IM2-GR) additionally allowed the two populations to change size exponentially after the population split and corresponded largely to the favoured model in the genomic study by Locke et al. (2011). In the fourth and most complex 2-population model (IM2-BN-GR), both populations retained a constant size after the population split, with the possibility for a sudden population size rescale followed by exponential growth or decline.

In order to test more biologically relevant demographic scenarios, we designed a series of 10-population models that incorporated the repeatedly reported extensive population substructure in extant orang-utan populations (Arora et al. 2010; Goossens et al. 2005; Kanhaswamy et al. 2006; Nater et al. 2013; Nater et al. 2011; Warren et al. 2001). The use of ten extant Demographic History of Orang-utans (Pongo spp.)
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population units models is justified by previously published data (Arora et al. 2010; Greminger et al. 2014; Nater et al. 2013; Nater et al. 2011), and the combination of patterns of population differentiation in both mtDNA and autosomal microsatellite markers, pointing to six populations on Borneo, one Sumatran population south of Lake Toba, and three Sumatran populations north of Lake Toba (see validation for ten population units in Supporting Figures S1 and S2). For all 10-population models, we assumed equal population sizes and equal symmetric migration rates among all populations within Borneo and among all populations north of Lake Toba, respectively, as well as a separate population size parameter for the population south of Lake Toba. We included asymmetric migration rates between Borneo and south of Lake Toba, and between north of Lake Toba and south of Lake Toba.

To assess to what extent the additional population units improve model fit, we first tested the best-fitting 2-population model against two basic 10-population models (IM10 and IM10-BOSU, Figure 2B). The IM10 model incorporated the population splitting sequence derived from mtDNA data, i.e. the populations north and south of Lake Toba show the oldest split, while Bornean populations diverged after this split (Nater et al. 2011). As this is in discordance with the current species designation (Groves 2001), which assigns a single species each to Sumatra and Borneo, we also tested this model against a model following the species split pattern (IM10-BOSU), i.e. with the oldest split between Sumatra and Borneo, to see if incomplete lineage sorting could be responsible for the particular phylogenetic pattern observed for mtDNA.

We further tested for the presence of population size changes in the demographic history of orang-utans, as suggested by previous studies (Arora et al. 2010; Goossens et al. 2006; Locke et al. 2011; Steiper 2006). First, we tested for signals of recent declines on Sumatra (IM10-DECSU), Borneo (IM10-DECBO) or both islands (IM10-DECALL) (Figure 2C).

In a second test, we evaluated the support for a bottleneck on Borneo (IM10-BNBO-DECSU), possibly linked to a refugium during the penultimate glaciation (Arora et al. 2010) (Figure 2D).

Last, we tested for evidence for a bottleneck on Sumatra linked to the Toba supereruption, either allowing for a broad prior range of the magnitude of decline (IM10-BNBO-TOBA-DECSU) or restricting to a severe bottleneck of less than 100 individuals in each of the four
Sumatran populations (IM10-BNBO-RECOL-DECSU), resembling a founder effect after local extinction and re-colonization events on Sumatra (Figure 2E).

**ABC Data Simulation**

To simulate genetic data under different demographic models, we used the software FASTSIMCOAL v1.1.2 (Excoffier & Foll 2011). Simulations for the different marker systems were run with the same set of parameters, whereby the effective population sizes were scaled 1 to 0.75, 0.25, and 0.25 for autosomal, X-chromosomal, mitochondrial and Y-chromosomal markers, respectively. We then used ARLSUMSTAT v3.5.1.3 (Excoffier & Lischer 2010) to calculate a total of 259 summary statistics for each simulated data set as well as for the observed data set (Supporting Table S5). The summary statistics were chosen in order to capture the information in the genetic data about population differentiation, within population diversity, and population size changes. To avoid problems with unreliable phasing, we only used summary statistics that do not require phased sequence data for X-chromosomal and autosomal loci. Since the number of simulated populations differed between the 2-population and 10-population models, summary statistic would not be directly comparable between the two sets of models. Therefore, when running the 10-population models, we applied a script that pooled the simulated data into a Bornean and a Sumatran group after each simulation step. Summary statistics were then also calculated island-wise, in order to be able to directly compare to the 2-population models.

We first performed an initial run of $2 \times 10^6$ simulations with the standard rejection sampler (Tavare et al. 1997). These simulations were used for both model selection and validation. To reduce the dimensionality of the summary statistics, we performed a principal component analysis (PCA) with the “prcomp” function in R version 2.12.1 (R Development Core Team 2010). A 100,000 random simulations from each of the two compared models were pooled and standardised, and these summary statistics were used to extract the loadings of the first ten principal components. We then transformed both the simulated and the observed data and used it to perform a multinomial logistic regression with the R package “abc” version 1.6. For this, we used the 0.1% of the simulations with the smallest Euclidean distance between the transformed summary statistics and the observed data.

In order to assess model fit, we also calculated the marginal densities and the probability of the observed data under the general linear model (GLM) used for the post sampling regression for each model with ABCtoolbox (Leuenberger & Wegmann 2010). For this, we again transformed both the simulated summary statistics as well as the observed data with the Demographic History of Orang-utans (Pongo spp.)
loadings for the first ten principal components. This time, PCA loadings were obtained for each model separately by using 100,000 random simulations. The GLM was built from the 2,000 simulations closest to the observed data, and we assessed the goodness of fit of all tested models to the observed data by calculating the p-value of the observed data under the GLM (Supporting Table S6). The p-value is representing the proportion of the retained simulations showing a lower or equal likelihood under the inferred GLM as compared to the observed genetic data (Wegmann et al. 2009a). Thus, low p-values indicate that the observed data is unlikely to have been generated under the inferred GLM, implying a bad model fit.

Parameter Estimation

To obtain good estimates of the posterior distributions of the parameters for the best-fitting model (IM10-BNBO-DECSU), we used a MCMC without likelihood method (Wegmann et al. 2009b). To reduce the dimensionality of the data and extract as much information as possible about the model parameters, we used the first 20,000 simulations with the standard sampler to define the first 12 orthogonal components of the summary statistics that maximise the covariance matrix between summary statistics and model parameters. For this, we applied a partial least-squares (PLS) regression approach (Boulesteix & Strimmer 2007) as implemented in the “pls” R package (Mevik & Wehrens 2007) and used the R script provided in the ABCtoolbox package. We defined the optimal number of PLS components by assessing the drop in the root mean squared error for each parameter with the inclusion of additional PLS components. This way, a large set of summary statistics is reduced to a number of independent components, whereby summary statistics that are most informative about the model parameters are weighted more than summary statistics that do not show much response to changing parameter values (Wegmann et al. 2009b). The initial simulations were also used to define the tolerance distance based on a tolerance level of 0.1 and to calibrate the transition kernel of the MCMC run with a rangeProp setting of 1 unit of standard deviation (Wegmann et al. 2009b; Wegmann et al. 2010). We then ran a total of $10^7$ iterations with the MCMC sampler, followed by a ABC-GLM post sampling regression (Leuenberger & Wegmann 2010) on the 10,000 simulations with the smallest Euclidean distance to the PLS components of the observed summary statistics. Finally, we used R to plot the posterior distributions of important model parameters.

ABC Validation

The performance of ABC in model selection and parameter estimation in complex demographic settings inevitably suffers from the loss of information when the observed and Demographic History of Orang-utans (Pongo spp.)
simulated genetic data are reduced to a set of summary statistics (Robert et al. 2011). This necessitates a careful validation of the employed ABC procedure in order to avoid biases in the approximation of posterior probabilities of evaluated models and the estimation of model parameters. Accordingly, we validated our model selection and parameter estimation approach with four different procedures. The first three validation approaches made use of so-called pseudo-observed data sets (pods), whereby parameter combinations are randomly drawn from the prior distributions and the corresponding summary statistics were simulated under a given model. These sets of summary statistics were then treated as if it were real observed data, but since the model and the corresponding parameter values that generated these summary statistics were known, we could use the pods to validate both our model selection and parameter estimation procedure.

In the first validation step, we investigated the model misclassification rate for each pair-wise model comparison by generating 100 pods under each model with parameters randomly drawn from the prior distributions. We then performed the same model selection procedure as with the real observed data and counted the number of assignments to each model. We derived the model misclassification rate by counting all assignments of pods to a model other than the one generating it (Figure 2).

Second, we assessed the accuracy of the parameter estimation, both in terms of different point estimators (mode, average and median) and over the whole posterior distribution under different tolerance levels (proportion of retained simulations). For this, we generated 1,000 pods under the best-fitting model (IM10-BNBO-DECSU) and performed the same parameter estimation procedure on each pods as for the real data. The accuracy of the point estimators was assessed using the average of the root mean squared errors (RMSE) over all 1,000 pods (Supporting Table S7), while the root mean integrated squared error (Leuenberger & Wegmann 2010) was used to assess accuracy over the whole posterior distribution (Supporting Table S8). The results indicated that accuracy of the posterior distributions is little affected by varying tolerance levels and that the mode of the distribution is the most accurate point estimator for parameter estimation.

Third, to increase confidence in the parameter estimates of the best-fitting model, we checked for biased posterior distributions by producing 1,000 pods under the best-fitting model with parameter values drawn from the prior distributions. We used ABCtoolbox to calculate the posterior quantiles of the true parameter values within the estimated posterior distributions for each pods and used a Kolmogorov-Smirnov test for uniformity in R (Wegmann et al. 2009b).
Significant deviation from uniformity after sequential Bonferroni correction (Rice 1989) would indicate biased posterior distributions (Cook et al. 2006). The distribution of posterior quantiles within which the true values of the pods fell did not significantly deviate from the expectation of uniformity for most parameters (Supporting Figure S4). In most cases where the posterior quantiles were not distributed uniformly, data points were overrepresented in the centre of the histogram, indicating that the posterior distributions were estimated too conservatively.

In a last validation approach, we tested if the best-fitting model (IM10-BNBO-DECSU) and the corresponding posterior distributions of the model parameters are able to reproduce the summary statistics of the observed data. For this, we randomly sampled 10,000 parameter sets from the inferred posterior distributions and used these to simulate genetic data under the best-fitting model. We then carried out a PCA transformation of the simulated data and plotted the first 16 principal components to check if the transformed observed data fell within the distribution of the simulated data (Supporting Figure S5). This was the case for all the first 16 principal components, suggesting that the best-fitting model and its inferred parameter estimates are well able to explain the observed data.

Results

Model Selection

We tested 12 demographic models, evaluating the impact of multiple demographic processes on the current genetic makeup of orang-utan populations (Figure 2). We first compared simple models that treated Bornean and Sumatran orang-utans as single populations, but differed in the opportunity for migration after the population split (IM2 vs. I2, Figure 2A). We found substantial support for the model allowing migration after the split (IM2, Bayes factor, i.e. ratio of model posterior probabilities (BF) 5.18). However, this simple isolation with migration model achieved only a very poor fit to the observed data, as shown by the probability of the observed data under the general linear model used for parameter estimation (GLM p-value) of 0.003, indicating that additional processes were involved in shaping the gene pool of orang-utans. Of all four 2-population models tested, we observed a very strong support for a model that allowed a sudden change in population size for both populations followed by exponential growth (IM2-BN-GR vs. I2, IM2, IM2-GR, BF 36.79). Still, this
model did not achieve a good fit to the observed data, as evidenced by a p-value of the observed data under the GLM of only 0.017 (Supporting Table S6).

The poor model fit of all tested 2-population models can be explained by the extensive population substructure within the two orang-utan species (Arora et al. 2010; Kanthaswamy et al. 2006; Nater et al. 2013; Nater et al. 2011; Warren et al. 2001), which differs to a great extent for female- and male-mediated marker systems (Nater et al. 2011; Nietlisbach et al. 2012). Accordingly, the Ne for each marker system varies to a large degree and cannot be described accurately with just one population size parameter per island. In agreement with this notion, we found that a basic model with ten current population units (IM10) achieved a better fit to the observed genetic data (GLM p-value 0.224) than all the 2-population models (Supporting Table S6), and also obtained much stronger statistical support when directly compared against the best 2-population model (IM10 vs. IM2-BN-GR, BF 830.21, Figure 2B). However, in our case, a better fit of the 10-population model compared to the 2-population models was not unexpected, since part of the observed genetic data was used beforehand to derive the number of population units in the 10-population models. When we computed summary statistics for the IM10 model without pooling the genetic data for the Sumatran populations north and south of Lake Toba, the model fit was still poor (GLM p-value 0.019). In order to improve model fit, we first tested whether a population split sequence following the species designation fits the data better than the pattern suggested by mtDNA data (deepest split within Sumatran orang-utans north and south of Lake Toba). This was strongly rejected by the observed data (IM10 vs. IM10-BOSU, BF 45.45, Figure 2B).

We then further tested for recent population declines on Sumatra (IM10-DECSU vs. IM10, BF 57.03), on Borneo (IM10-DECBO vs. IM10, BF 0.48) or on both islands (IM10-DECALL vs. IM10-DECSU, BF 0.94, Figure 2C). Incorporating a population decline on Sumatra considerably improved the model fit (GLM p-value 0.553).

Next, we tested a model incorporating a bottleneck on Borneo together with a recent decline on Sumatra (Figure 2D), which revealed substantial support for a bottleneck on Borneo (IM10-BNBO-DECSU vs. IM10-DECSU, BF 3.60).

Finally, we evaluated the statistical support for a bottleneck on Sumatra associated with the Toba supereruption (Figure 2E). We found substantial support against a bottleneck on Sumatra in general (IM10-BNBO-DECSU vs. IM10-BNBO-TOBA-DECSU, BF 3.29), and...
overwhelming support against a severe bottleneck (less than 100 individuals per population) (IM10-BNBO-DECSU vs. IM10-BNBO-RECOL-DECSU, BF 10,887.60). After performing a series of hierarchical model selection steps, we were able to identify a demographic model (IM10-BNBO-DECSU) capable of reproducing the observed patterns of DNA variation in the two current orang-utan species. Therefore, this model is likely to capture the biologically most relevant processes in the demographic history of orang-utans.

**Parameter Estimation**

We estimated the model parameters for the selected 10-population model (IM10-BNBO-DECSU, Figure 3) based on a total of 10 million simulations (Table 3, Figure 4). The parameter estimates point to a current $N_e$ of ~970 diploid individuals in each of the six Bornean populations. We found support for a bottleneck on Borneo starting ~135 ka and ending ~82 ka, during which $N_e$ on Borneo was reduced from an ancestral $N_e$ of ~17,000 individuals to ~2,600 individuals. The bottleneck on Borneo was followed by population recovery and substructuring, with a current total $N_e$ of all Bornean populations of ~6,150 individuals.

On Sumatra, the three populations north of Lake Toba suffered a decline ~24 ka from a $N_e$ of ~10,500 to currently only ~960 individuals in each of the three populations, corresponding to a total $N_e$ in the meta-population north of Lake Toba of ~38,300 and ~3,300 individuals before and after the decline, respectively. We estimated that population structure north of Lake Toba was established ~860 ka, with an ancestral effective population size of ~14,400 individuals. The population south of Lake Toba also went through a recent decline ~24 ka from a $N_e$ of ~24,200 individuals in the ancestral population to currently only ~1,030 individuals. Thus, Sumatran orang-utan populations first expanded during the Middle Pleistocene before experiencing an island-wide population crash in the Late Pleistocene or Early Holocene.

We inferred the population split time between Borneo and south Toba as ~1.13 Ma, and between north and south of Lake Toba as ~3.39 Ma. Gene flow between Borneo and Sumatra appears to have ceased ~87 ka, but this parameter was associated with a broad posterior distribution. We found no evidence for asymmetric migration rates between Borneo and south of Lake Toba, and between south of Lake Toba and north of Lake Toba. The migration rates between the two islands were comparable to the migration rates over the Toba caldera on...
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Sumatra, while migration rates among the populations on Borneo and among those north of Toba, respectively, were estimated to be about a magnitude higher.

Discussion

Our modelling approach capitalized on the use of multiple genetic marker systems and an extensive set of geographically well-defined samples, in contrast to previous studies, which based their findings on a small number of captive individuals with poorly recorded provenance (Locke et al. 2011; Mailund et al. 2011; Mailund et al. 2012). Thus, our study was able to shed light on important aspects of orang-utan demographic history that so far remained unexamined due to non-representative sampling and dismissal of within-species population structure. For instance, the inferred model by Locke et al. (2011) of a continuously expanding Sumatran orang-utan population with a substantially larger current Ne as compared to Bornean orang-utans was unrealistic in the light of current species distribution and abundance, and did not capture recent population dynamics. Our results indicate that such misleading signals are the result of a recent decline and deep divergence of orang-utan populations on Sumatra, which yields a larger long-term Ne for Sumatran orang-utans as compared to Bornean orang-utans in oversimplified demographic models.

Inference of Best-Fitting Model

We inferred that a model with comprehensive population structure, a bottleneck on Borneo and a recent decline on Sumatra (IM10-BNBO-DECSU) fits the observed data significantly better than a range of simplified models that treat each orang-utan species as a single panmictic population. Estimation of demographic parameters under this model revealed a population split time between Borneo and Sumatran populations south of Lake Toba of just over a million years ago, followed by bidirectional gene flow. This species split time estimate is considerably older than estimates obtained using whole genome data, suggesting a species split time of between 330 and 600 ka (Locke et al. 2011; Mailund et al. 2011; Mailund et al. 2012). Such recent species split estimates are, however, in disagreement with findings based on mitochondrial DNA, which yielded divergence time estimates of island specific mtDNA lineages of 1–5 Ma (Nater et al. 2011; Steiper 2006; Warren et al. 2001; Xu & Arnason 1996; Zhang et al. 2001; Zhi et al. 1996).

The discrepancy between model-based species split estimates using exclusively autosomal data and mtDNA divergence time estimates from phylogenetic methods is owed to two
idiosyncrasies in the biology of orang-utans. First, due to the pronounced philopatric tendencies of female orang-utans (Arora et al. 2012; Galdikas 1995; Nietlisbach et al. 2012; van Noordwijk et al. 2012), mtDNA has likely experienced only little if any gene flow between the two species after the species split. Therefore, the coalescent time of island-specific mitochondrial lineages is expected to predate the population split between Borneo and Sumatra, depending on $N_e$ in the ancestral population (Nichols 2001). Second, due to male-mediated gene flow, model-based approaches using solely autosomal data are likely to underestimate species split times, as disentangling the contributions of migration and split time remains challenging (Hey & Nielsen 2004). The recent split time estimates from autosomal genomic data might reflect the end of an initial period of frequent, but strictly male-driven gene flow after the species split. Such complex temporal fluctuations in migration rates, as expected during glacial cycles for Sundaland species, are so far not properly addressed in any applied demographic model. Still, combining markers with different inheritance patterns as done in this study is likely to improve the estimates of both migration rates and split times in species with sex-biased dispersal such as orang-utans.

Our findings of recent gene flow between Bornean and Sumatran orang-utans are in agreement with previous observations (Becquet & Przeworski 2007; Muir et al. 2000; Verschoor et al. 2004). In their genomic study, Locke and colleagues (2011) found an unexpectedly high incidence of low-frequency mutations shared between Borneo and Sumatra, which also hints at recent gene flow between the two islands. Contrary to studies that indicated the presence of impassable dispersal barriers on the exposed Sunda shelf, either due to large river systems (Harrison et al. 2006) or a putative savannah corridor (Bird et al. 2005; Gathorne-Hardy et al. 2002), it seems that habitat conditions during glacial periods did at least sporadically allow male orang-utans to cross the exposed Sunda shelf. However, given the strict and long lasting separation of mtDNA lineages on both islands (Nater et al. 2011), it appears that the exposed shelf was not covered with forest able to sustain orang-utan populations over prolonged periods. In fact, large parts of the Sunda shelf between Borneo and Sumatra were covered with nutrient-poor sandy soils (Bird et al. 2005; Slik et al. 2011). Forests on such soil types are characterized by low growth and productivity (Paoli et al. 2010). These constraints might explain why orang-utan populations on both islands could not expand onto the exposed shelf to an extent where population admixture and thus exchange of mtDNA lineages was possible.
Glacial Cycles and Population Size Changes

Since we also tested models that incorporated sudden population size changes, we were able to detect signals of a population bottleneck on Borneo. In contrast to Sumatra, the currently observed pattern of strong population differentiation on Borneo (Arora et al. 2010; Warren et al. 2001) seems to have been established only recently, as parameter estimation indicated that Bornean orang-utans were organized at least temporarily as a single panmictic population before ~80 ka. At ~140 ka, the ancient population on Borneo experienced a sudden drop in $N_e$ from ~17,000 to ~2,500 individuals, which then recovered again to the current total $N_e$ of ~6,000 for all Bornean orang-utans. Such a change in both $N_e$ as well as population structure could be explained by a common Bornean refugium during either the penultimate (190–130 ka) or last (110–18 ka) glacial period, when the drier and more seasonal climate might have caused a drastic reduction of rainforest coverage on Borneo (Bird et al. 2005; Gathorne-Hardy et al. 2002; Morley 2000). Population contractions with subsequent expansions likely occurred multiple times on Borneo during Pleistocene glacial and interglacial cycles, but incorporating such complex population dynamics into a demographic model is currently not feasible with the data at hand.

Interestingly, a similar signal of a glacial refugium with subsequent population structuring, as observed in Bornean orang-utans, has been found in western gorillas (Gorilla gorilla). By using a demographic modelling approach comparable to our study, Thalmann et al. (2011) found that the two subspecies of western gorillas (G. g. gorilla and G. g. diehli) diverged only about ~18 ka, thus directly following the last glacial maximum (LGM) 19–26 ka (Clark et al. 2009). Furthermore, the ancient population of western gorillas exhibited a $N_e$ of just ~2,500 individuals as compared to 22,000 and 17,000 individuals in the two subspecies after the population split. Therefore, it seems that western gorillas, similar to Bornean orang-utans, were constrained to a relatively small refugial population during glacial periods from which they subsequently expanded when the climate got warmer and wetter during interglacials.

Geological Processes and Population Size Changes

Linking bottleneck signals to specific environmental processes is difficult due to the large confidence intervals associated with most parameter estimates. For instance, the 90%-highest posterior interval for the estimate of the start of the bottleneck on Borneo (21–348 ka) also overlaps with the Toba supereruption on northern Sumatra ~73 ka (Chesner et al. 1991). It has been hypothesized that this colossal explosive eruption might have had a strong global...
impact, causing a severe bottleneck in humans (Rampino & Ambrose 2000). However, evidence presented here points toward climatic changes during the glacial periods rather than the Toba supereruption as being the main cause for the detected bottleneck on Borneo, as our results showed that the supereruption did not even have a strong impact on the Sumatran populations despite their much closer geographic proximity. Models incorporating a severe bottleneck in the Sumatran populations around the time of the supereruption were clearly rejected and the signal of a recent population decline on Sumatra was considerably younger than the Toba supereruption. Studies indicate that the destruction caused by the Toba supereruption had been geographically limited, as shown by the distribution of rainforest refugia in Southeast Asia (Gathorne-Hardy et al. 2002), including on Mentawai Island around 350 kilometres from the Toba caldera (Gathorne-Hardy & Harcourt-Smith 2003), as well as the similar composition of Southeast Asian fossil sites before and after the date of the supereruption (Louys 2007). Given the proximity of contemporary populations of Sumatran orang-utans to the Toba caldera and the strong dependency of orang-utans on intact rain forest habitat, they are undoubtedly one of the most striking examples illustrating the limited impact of the Toba supereruption on the local flora and fauna in Southeast Asia. However, the lack of bottlenecks signals in the Sumatran populations does not imply that the activity of the Toba volcano did not influence the population history of Sumatran orang-utans at all. Rather, the results of this study, as well as previous findings (Nater et al. 2013; Nater et al. 2011), showed strikingly that the Toba eruptions must have repeatedly caused devastating damage to the local surroundings, which led to a long-lasting separation of gene pools north and south of Lake Toba.

In contrast to Toba as cause for the bottleneck on Borneo, a contraction of rainforests following the colder and drier climate during the last glacial period well explains the absence of a similar bottleneck in the Sumatran population history. During the generally drier glacial periods, large parts of Sumatra experienced considerably more rain fall compared to Borneo (Gathorne-Hardy et al. 2002; Whitten et al. 2000), because the Barisan mountain range running the length of Sumatra acted as a barrier for the wet monsoon winds, causing high precipitation along its western slopes (Whitten et al. 2000). This mountain ridge effect in combination with the close proximity to the sea during glacial periods, when sea levels were low, might have allowed large areas of rainforest to persist on Sumatra during glacial periods (Gathorne-Hardy et al. 2002). Thus, Sumatran orang-utans were almost certainly not forced into glacial refugia to the same extent as Borneans.
Anthropogenic Impacts on Orang-Utan Populations

While Sumatran orang-utans did not seem to go through glacial bottlenecks, we found evidence for recent and drastic declines in population sizes north and south of Lake Toba. These signals of population decline cannot be attributed to the large-scale human-induced habitat degradation that started in the last century (Rijksen & Meijaard 1999), of which genetic signals were found in previous studies of Bornean orang-utans (Goossens et al. 2006; Sharma et al. 2012). Rather, our results point toward an earlier decline in the Late Pleistocene or Early Holocene. In the Late Pleistocene, orang-utans went extinct on the Southeast Asian mainland as well as in many Sundaland regions (Delgado & Van Schaik 2000; Jablonski 1998; Rijksen & Meijaard 1999). Furthermore, the Pleistocene-Holocene boundary is characterized by the disappearance of many large-bodied animals world-wide (Koch & Barnosky 2006), including large parts of the megafauna in Southeast Asia (Louys et al. 2007). The increased occurrence of megafaunal extinctions during this period has been attributed to climatic changes following the LGM, the impact of human hunting and human-induced habitat changes, or the combination of these two factors (reviewed in Koch & Barnosky 2006).

Both climatic and anthropogenic factors might have played a role in the decline and local extinctions of orang-utan populations in the Late Pleistocene. During the LGM, the drier and more seasonal climate caused a shifting of zones of evergreen rainforest toward the equator (Flenley 1998; Jablonski 1998; Morley 2000), likely causing populations in southern China to go extinct. The warmer climate following the LGM was accompanied by rising sea levels, which drastically increased the extent of coastlines in Sundaland (Voris 2000). This enlargement of coastal habitat might have promoted an expansion of early modern humans on Sundaland, leading to increased hunting pressure on large-bodied animals, including orang-utans (Hill et al. 2007; Soares et al. 2008). Such hunting by modern humans might have caused the local extinctions of orang-utans on many Sundaland islands, and led to a strong decline in Sumatran populations north and south of Lake Toba. Bornean orang-utans did not seem to be as strongly affected by human hunting, probably because the large size and low productivity of Borneo left enough inland areas with relatively low human densities (Delgado & Van Schaik 2000).

Our modelling approach revealed that the two recognised orang-utan species experienced drastically different demographic histories. Sumatran orang-utans exhibit a deep and temporally stable population structure, including an old divergence of gene pools north and Demographic History of Orang-utans (Pongo spp.)
south of Lake Toba with limited amount of gene flow over the Toba caldera. The populations on Sumatra recently suffered a strong decline, which, in combination with the strong population structure, explains the high genetic diversity found in recent genomic studies despite their low current census size (Locke et al. 2011; Prado-Martinez et al. 2013). In contrast, we find that the population structure currently observed within Bornean orang-utans has been established only recently and the population went through at least one bottleneck most likely associated with a glacial refugium. These results strongly suggest that special consideration needs to be given to demographic factors when analysing adaptive evolutionary processes in great apes. Due to their strong dependence on intact forest habitat, most great ape taxa were severely affected by the climate shifts during glacial periods, which were accompanied by drastic changes in forest coverage in the tropics (Flenley 1998; Morley 2000). Accordingly, great ape populations experienced population bottlenecks, founder events, population expansions and population structuring as recent as 15,000 years ago (Clark et al. 2009). Given the long generation time of all great apes (18–30 years, Wich et al. 2009), great ape populations will likely not have reached an equilibrium state for most genomic regions. Thus, population expansions and substructuring caused by relatively recent climatic changes might produce erroneous signals of selective sweeps if demography is not taken into account. Our results therefore emphasize the need to further advance the development of tools to jointly estimate demography and selection in order to unravel the convoluted evolutionary history of great apes (Li et al. 2012).

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Demographic History of Orang-utans (Pongo spp.)

References


Demographic History of Orang-utans (Pongo spp.)
Demographic History of Orang-utans (Pongo spp.)


Demographic History of Orang-utans (Pongo spp.)
Demographic History of Orang-utans (*Pongo* spp.)

Data Accessibility

Sequence data not published previously is accessible under GenBank accession numbers XXXX-YYYY.

Author Contributions

AN, MPG, CPvS, and MK designed the study; BG, IS, EJV, and KSW provided samples; AN, MPG, and NA performed laboratory procedures; AN and MPG conducted genetic data analysis; AN performed demographic modelling; AN wrote the manuscript; MPG, NA, CPvS, and MK critically revised the manuscript and provided comments at all stages; BG, EJV, and KSW edited the final manuscript.
Table 1: Sample sizes for the different marker systems in the ten geographic regions

<table>
<thead>
<tr>
<th>Sampling regiona</th>
<th>mtDNA</th>
<th>Y-STRs</th>
<th>Autosomal STRs</th>
<th>Autosomal regions</th>
<th>Xq13.3</th>
</tr>
</thead>
<tbody>
<tr>
<td>North Kinabatangan (NK)</td>
<td>6</td>
<td>10</td>
<td>32</td>
<td>4</td>
<td>3</td>
</tr>
<tr>
<td>South Kinabatangan (SK)</td>
<td>13</td>
<td>15</td>
<td>76</td>
<td>4</td>
<td>3</td>
</tr>
<tr>
<td>East Kalimantan (EK)</td>
<td>7</td>
<td>9</td>
<td>34</td>
<td>4</td>
<td>5</td>
</tr>
<tr>
<td>Sarawak (SR)</td>
<td>8</td>
<td>2</td>
<td>12</td>
<td>2</td>
<td>1</td>
</tr>
<tr>
<td>Central Kalimantan (CK)</td>
<td>9</td>
<td>9</td>
<td>68</td>
<td>2</td>
<td>2</td>
</tr>
<tr>
<td>West Kalimantan (WK)</td>
<td>9</td>
<td>8</td>
<td>32</td>
<td>4</td>
<td>4</td>
</tr>
<tr>
<td>Batang Toru (BT)</td>
<td>8</td>
<td>8</td>
<td>18</td>
<td>4</td>
<td>3</td>
</tr>
<tr>
<td>North Aceh (NA)</td>
<td>7</td>
<td>15</td>
<td>32</td>
<td>6</td>
<td>3</td>
</tr>
<tr>
<td>Langkat (LK)</td>
<td>14</td>
<td>15</td>
<td>66</td>
<td>10</td>
<td>6</td>
</tr>
<tr>
<td>West Alas (WA)</td>
<td>37</td>
<td>38</td>
<td>104</td>
<td>4</td>
<td>7</td>
</tr>
<tr>
<td>Total</td>
<td>118</td>
<td>129</td>
<td>474</td>
<td>44</td>
<td>37</td>
</tr>
</tbody>
</table>

a, sampling regions corresponding to Figure 1. Sample sizes are given as number of sampled chromosomes. The light grey shading refers to Bornean populations, middle grey to Sumatran populations north of Lake Toba, and dark grey to the Sumatran population south of Lake Toba.
Table 2: Summary statistics for the marker systems used in the ABC analysis.

<table>
<thead>
<tr>
<th>Sequences</th>
<th>L-Bases&lt;sup&gt;a&lt;/sup&gt;</th>
<th>Group</th>
<th>N&lt;sub&gt;Ind&lt;/sub&gt;&lt;sup&gt;b&lt;/sup&gt;</th>
<th>N&lt;sub&gt;Seg&lt;/sub&gt;&lt;sup&gt;c&lt;/sup&gt;</th>
<th>π&lt;sup&gt;d&lt;/sup&gt;</th>
<th>θ&lt;sub&gt;W&lt;/sub&gt;&lt;sup&gt;e&lt;/sup&gt;</th>
<th>D&lt;sup&gt;f&lt;/sup&gt;</th>
</tr>
</thead>
<tbody>
<tr>
<td>mtDNA (16S, ND3, CYTB)</td>
<td>1,355</td>
<td>Borneo</td>
<td>52</td>
<td>19</td>
<td>0.0022</td>
<td>0.0031</td>
<td>-0.92</td>
</tr>
<tr>
<td></td>
<td></td>
<td>South Toba</td>
<td>8</td>
<td>1</td>
<td>0.0002</td>
<td>0.0003</td>
<td>-1.05</td>
</tr>
<tr>
<td></td>
<td></td>
<td>North Toba</td>
<td>58</td>
<td>41</td>
<td>0.0100</td>
<td>0.0066</td>
<td>1.79</td>
</tr>
<tr>
<td>Autosomal regions</td>
<td>8,238</td>
<td>Borneo</td>
<td>10</td>
<td>19.50 ± 4.56</td>
<td>0.0033</td>
<td>0.0027</td>
<td>0.68 ± 0.66</td>
</tr>
<tr>
<td>(Chr2a_R17, Chr9_R16, Chr12_R1, Chr19_R7)</td>
<td></td>
<td>South Toba</td>
<td>2</td>
<td>13.50 ± 7.79</td>
<td>0.0037</td>
<td>0.0036</td>
<td>0.08 ± 0.49</td>
</tr>
<tr>
<td></td>
<td></td>
<td>North Toba</td>
<td>10</td>
<td>28.75 ± 4.66</td>
<td>0.0046</td>
<td>0.0040</td>
<td>0.51 ± 0.52</td>
</tr>
<tr>
<td>Xq13.3</td>
<td>8,055</td>
<td>Borneo</td>
<td>18</td>
<td>6</td>
<td>0.0001</td>
<td>0.0002</td>
<td>-1.11</td>
</tr>
<tr>
<td></td>
<td></td>
<td>South Toba</td>
<td>3</td>
<td>13.50 ± 7.79</td>
<td>0.0037</td>
<td>0.0036</td>
<td>0.08 ± 0.49</td>
</tr>
<tr>
<td></td>
<td></td>
<td>North Toba</td>
<td>15</td>
<td>28.75 ± 4.66</td>
<td>0.0046</td>
<td>0.0040</td>
<td>0.51 ± 0.52</td>
</tr>
<tr>
<td>Microsatellites</td>
<td>N&lt;sub&gt;Loci&lt;/sub&gt;&lt;sup&gt;g&lt;/sup&gt;</td>
<td>Group</td>
<td>N&lt;sub&gt;Ind&lt;/sub&gt;</td>
<td>N&lt;sub&gt;A&lt;/sub&gt;&lt;sup&gt;h&lt;/sup&gt;</td>
<td>H&lt;sub&gt;O&lt;/sub&gt;&lt;sup&gt;i&lt;/sup&gt;</td>
<td>H&lt;sub&gt;E&lt;/sub&gt;&lt;sup&gt;j&lt;/sup&gt;</td>
<td>G-W&lt;sup&gt;k&lt;/sup&gt;</td>
</tr>
<tr>
<td>Autosomal-STR</td>
<td>25</td>
<td>Borneo</td>
<td>127</td>
<td>3.16 ± 4.13</td>
<td>0.53 ± 0.22</td>
<td>0.61 ± 0.25</td>
<td>0.90 ± 0.15</td>
</tr>
<tr>
<td></td>
<td></td>
<td>South Toba</td>
<td>9</td>
<td>3.84 ± 1.18</td>
<td>0.60 ± 0.23</td>
<td>0.62 ± 0.16</td>
<td>0.72 ± 0.22</td>
</tr>
<tr>
<td></td>
<td></td>
<td>North Toba</td>
<td>101</td>
<td>6.32 ± 3.11</td>
<td>0.61 ± 0.16</td>
<td>0.65 ± 0.16</td>
<td>0.82 ± 0.17</td>
</tr>
<tr>
<td>Y-STR</td>
<td>11</td>
<td>Borneo</td>
<td>53</td>
<td>3.18 ± 2.48</td>
<td>-</td>
<td>0.31 ± 0.33</td>
<td>0.90 ± 0.14</td>
</tr>
<tr>
<td></td>
<td></td>
<td>South Toba</td>
<td>8</td>
<td>1.27 ± 0.65</td>
<td>-</td>
<td>0.08 ± 0.19</td>
<td>0.88 ± 0.18</td>
</tr>
<tr>
<td></td>
<td></td>
<td>North Toba</td>
<td>68</td>
<td>1.91 ± 1.64</td>
<td>-</td>
<td>0.12 ± 0.24</td>
<td>0.91 ± 0.17</td>
</tr>
</tbody>
</table>

Statistics are provided as average and standard deviation for marker systems with multiple independent loci; <sup>a</sup>, sequence length in base pairs; <sup>b</sup>, number of sampled individuals; <sup>c</sup>, number of segregating sites; <sup>d</sup>, nucleotide diversity; <sup>e</sup>, Watterson’s θ per base pair; <sup>f</sup>, Tajima’s D (Tajima 1989); <sup>g</sup>, number of loci; <sup>h</sup>, number of alleles; <sup>i</sup>, observed heterozygosity; <sup>j</sup>, expected heterozygosity; <sup>k</sup>, Garza-Williamson index (Garza & Williamson 2001);
### Table 3: Estimates of the model parameters for the selected 10-population model with a bottleneck on Borneo and a recent decline on Sumatra (IM10-BNBO-DECSU).

<table>
<thead>
<tr>
<th>Parameter(^a)</th>
<th>Prior(^b)</th>
<th>Mode</th>
<th>Mean</th>
<th>90%-HPD(^c)</th>
</tr>
</thead>
<tbody>
<tr>
<td>N(_{NOW}) BO [ind] (6)</td>
<td>logunif[100;10,000]</td>
<td>974</td>
<td>1,028</td>
<td>348–3,011</td>
</tr>
<tr>
<td>N(_{NOW}) NT [ind] (3)</td>
<td>logunif[100;10,000]</td>
<td>963</td>
<td>933</td>
<td>239–3,613</td>
</tr>
<tr>
<td>N(_{NOW}) ST [ind] (1)</td>
<td>logunif[100;10,000]</td>
<td>1,034</td>
<td>952</td>
<td>189–4,514</td>
</tr>
<tr>
<td>N(_{BN}) BO [ind] (1)</td>
<td>logunif[100;10,000]</td>
<td>2,598</td>
<td>1,486</td>
<td>286–9,988</td>
</tr>
<tr>
<td>N(_{ANC}) BO [ind] (1)</td>
<td>logunif[1,000;100,000]</td>
<td>17,046</td>
<td>12,344</td>
<td>2,171–89,115</td>
</tr>
<tr>
<td>N(_{ANC}) NT [ind] (3)</td>
<td>logunif[100;10,000]</td>
<td>10,508</td>
<td>11,278</td>
<td>1,886–78,264</td>
</tr>
<tr>
<td>T(_{BNEND}) BO [yrs]</td>
<td>unif[8,750;400,000]</td>
<td>81,946</td>
<td>149,580</td>
<td>8,848–283,785</td>
</tr>
<tr>
<td>T(_{STRUC}) NT [yrs]</td>
<td>unif[75;1,500]</td>
<td>861</td>
<td>820</td>
<td>267–1,398</td>
</tr>
<tr>
<td>T(_{DEC}) SU [yrs]</td>
<td>unif[1.0;3.5]</td>
<td>23,651</td>
<td>36,200</td>
<td>4,119–67,272</td>
</tr>
<tr>
<td>T(_{SPLIT}) NT [yrs]</td>
<td>unif[1,500;4,000]</td>
<td>3,392</td>
<td>2,995</td>
<td>2,101–3,999</td>
</tr>
<tr>
<td>T(_{MIGSTOP}) [yrs]</td>
<td>unif[2.5;4.2]</td>
<td>87,034</td>
<td>161,862</td>
<td>8,849–310,833</td>
</tr>
<tr>
<td>Log(m(_{BO-NT})) [migrants/ind/gen]</td>
<td>unif[-5.0;-3.0]</td>
<td>-3.55</td>
<td>-3.96</td>
<td>-4.79–3.09</td>
</tr>
<tr>
<td>Log(m(_{NT-BO})) [migrants/ind/gen]</td>
<td>unif[-5.0;-3.0]</td>
<td>-3.42</td>
<td>-3.84</td>
<td>-4.61–3.10</td>
</tr>
<tr>
<td>Log(m(_{ST-NT})) [migrants/ind/gen]</td>
<td>unif[-5.0;-3.0]</td>
<td>-3.65</td>
<td>-3.92</td>
<td>-4.71–3.06</td>
</tr>
<tr>
<td>Log(m(_{NT-BO})) [migrants/ind/gen]</td>
<td>unif[-4.0;-2.0]</td>
<td>-2.52</td>
<td>-2.90</td>
<td>-3.66–2.02</td>
</tr>
<tr>
<td>Log(m(_{ST-NT})) [migrants/ind/gen]</td>
<td>unif[-4.0;-2.0]</td>
<td>-2.51</td>
<td>-2.89</td>
<td>-3.65–2.03</td>
</tr>
</tbody>
</table>

\(^a\) BO = Borneo, NT = Sumatra north of Lake Toba, ST = Sumatra south of Lake Toba, N\(_{NOW}\) = current effective population size, N\(_{BN}\) = effective population size during population bottleneck, N\(_{ANC}\) = ancestral effective population size, N\(_{STRUC}\) = effective population size before recent decline, T\(_{BNEND}\) = time since population bottleneck ended, T\(_{BNSTART}\) = time when population bottleneck started, T\(_{SPLIT}\) = population split time, T\(_{DEC}\) = time since population decline, T\(_{STRUC}\) = time since establishment of population structure, T\(_{MIGSTOP}\) = time since migration between Borneo and Sumatra stopped, m = migration rate per individual per generation (an illustration of the meaning of the different model parameters can be found in Figure 3), the number in brackets next to the population size parameters refer to the number of simulated populations of this size each; \(^b\) The prior distributions for the parameter values were either uniform or loguniform within the boundaries provided in squared brackets; \(^c\) 90%-highest posterior density interval.
Figure 1: Map of sampling regions in Sundaland used for the demographic modelling. Shaded areas represent the current distribution of the Sumatran orang-utans and the three subspecies of Bornean orang-utans. The grey line indicates the extent of the exposed Sunda shelf during the LGM (19–26 ka, -120 meters below current sea level).
Demographic History of Orang-utans (*Pongo* spp.)
Figure 2: Schematic representation of the hierarchical model testing procedure. The 12 tested demographic models can be divided into four 2-population models and eight 10-populations models (IM10-DECBO not shown). The box above the left-right arrow shows the model posterior probabilities for each model comparison pair. The overall best-fitting model (IM10-BNBO-DECSU) is shown in a black frame. The box below the left-right arrow shows the power to distinguish between the two compared models as evaluated in a cross-validation procedure with 100 validations for each model, with the upper left and lower right boxes showing the correct model assignments for model 1 and model 2, respectively (SU = Sumatra, BO = Borneo, NT = Sumatra north of Lake Toba, ST = Sumatran south of Lake Toba). A) Comparison of four 2-population models, testing gene flow after the population split, exponential population growth or decline after the population split, and sudden population size change followed by exponential growth or decline. B) Comparison between the best fitting 2-population model and two 10-population models incorporating population structure. C) Tests of recent population declines on Sumatra, and Sumatra as well as Borneo. D) Test of population bottleneck on Borneo. E) Testing of a population bottleneck on Sumatra associated with the Toba supereruption 65–75 ka. The leftmost model implements a bottleneck in all four populations on Sumatra, followed by exponential population recovery. The rightmost model is similar, but restricts the bottleneck to a size of less than 100 surviving individuals per population, thus representing a scenario where regions devastated by the Toba eruption were recolonized from other areas after restoration of the rain forest habitat.
Figure 3: Schematic representation of the selected 10-population model with a bottleneck on Borneo and recent population declines in all Sumatran populations (IM10-BNBO-DECSU).

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Figure 4: Posterior distributions of important model parameter under the selected 10-population model (IM10-BNBO-DECsu). The abbreviations of the model parameters correspond to the labels in Figure 3. For better comparability, the effective populations sizes of the structured meta-populations on Borneo and north of Lake Toba are given as the total effective sizes according to the formula $N_e = D \times N \times (1 + (1/(4 \times N \times m)))$, with $D$ corresponding to the number of subpopulations, $N$ to the mean subpopulation size and $m$ to the total migration rate per individual per generation within the meta-population (Nichols et al. 2001).