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1 **Ultrastructure of the anterior adhesive apparatus of the gill parasite**
2 ***Macrogyrodactylus clarii* and skin parasite *M. congolensis* (Monogenea;**
3 **Gyrodactylidae) from the catfish *Clarias gariepinus***

4
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12

13 **Abstract**

14 Transmission electron microscopy (TEM) was used to study the anterior adhesive
15 apparatus of the gill parasite *Macrogyrodactylus clarii* Gussev, 1961 and skin parasite *M.*
16 *congolensis* (Prudhoe, 1957) Yamaguti, 1963. Despite the different microhabitats occupied by
17 these parasites, they have a similar anterior adhesive system. In both parasites, this consists of
18 three types of gland cells: G1 cells that produce rod-shaped bodies (S1), G2 cells
19 manufacture irregularly shaped bodies (S2) and G3 cells form mucoid-like secretions (S3). In
20 the cytoplasm of G1 cells, a single layer of microtubules encloses each developing rod-
21 shaped body. Some fully developed S1 bodies are attached to each other, forming large
22 condensed globules. S1 bodies are extruded through multiple apertures whereas S2 and S3
23 bodies are released through ducts each with a single opening. The adhesive sacs are lined
24 with two types of tegument (st1 and st2). A third tegument type (st3) connects the st2
25 tegument with the general body tegument. Only st1 has microvilli. Each adhesive sac is

26 provided with a spike-like sensillum and single unciliated sense organ. The possible
27 functions of microvilli in increasing the surface area and assistance in spreading and mixing of
28 the adhesive secretion and the role of sense organs associated with the adhesive sacs are
29 discussed.

30

31 **Key words:** Platyhelminthes, Monogenea, fish ectoparasite, temporary adhesion, adhesive
32 apparatus, ultrastructure

33 **1. Introduction**

34 Monogenean ectoparasites attach to their hosts primarily with their posterior attachment
35 organ (haptor), which is equipped with hamuli and marginal hooklets [1], but in order to
36 move from one position to another they rely on their anterior adhesive apparatus [2].
37 Typically, monogeneans move on the host or artificial substrates by stretching out their
38 bodies and attaching with head lobes to the host tissue, releasing and moving the haptor to
39 attach close to the adhesive areas of the head lobes, and then they detach the head lobes to
40 move anteriorly where they attach again to a new site. Some can move in a similar leech-like
41 manner upside down, using the water surface tension [3].

42 The anterior adhesive apparatus has been studied with transmission electron microscopy
43 (TEM) and/or scanning electron microscopy (SEM) in many monogenean parasites,
44 including the gyrodactylids [4,5,6], dactylogyrids [7,8], entobdellids [2,9], acanthocotylids
45 [10], monocotylids [11,12] and ancycrocephalids [13]. They have various kinds of gland cells
46 that open either into the outer syncytial tegumental layer [7,14], or onto the specialized
47 haptoral [13,15,16,17] or ventrally-located head regions (see for example, El-Naggar and
48 Khidr [8], Wong et al. [13]). The monogenean anterior adhesive apparatus produces one to
49 three types of secretion bodies. Species with rod-shaped bodies (S1), spherical bodies (S2)

50 and irregularly-shaped, electron-lucent vesicles (S3) include *Gyrodactylus eucaliae* (see
51 Kritsky, 1978) [4], *G. sprostonae* (see Yuan and Long [18]), *Dactylogyrus amphibothrium*
52 and *D. hemiamphibothrium* (see El-Naggar and Kearns [7]), *D. aristichthys* (see Yuan and
53 Long [19]), *Cichlidogyrus halli* (see El-Naggar and Khidr [8]) and *Merizocotyle icopae* (see
54 Cribb et al. [20]). Two types of secretion, rod-shaped bodies (S1) and spherical bodies (S2),
55 were reported in *Entobdella soleae* (see Kearns and Evans-Gowing [9]), *Acanthocotyle*
56 *lobianchi* (see Rees and Kearns [10]) and *Caballeria liewi* (see Wong et al. [13]). Only one
57 kind of secretion, rods, is produced in the anterior adhesive apparatus of *Monocotyle*
58 *spiremae* (see Cribb et al. [11]) and spherical bodies in *Enterogyrus cichlidarum* (see Khidr
59 et al. [21]).

60 Two gyrodactylid monogeneans of the Nile catfish, *Clarias gariepinus*,
61 *Macrogyrodactylus clarii* [22] and *M. congolensis* [23,24] infect the gills [25], and the skin
62 and fins [26], respectively. Although the haptors of *M. clarii* and *M. congolensis* show the
63 same basic structure, there are some differences [25,26] possibly reflecting the different
64 habitats of the parasites. The haptor of *M. clarii* possesses two lateral rows of tegumental
65 papillae, whilst that of *M. congolensis* has three rows (two lateral and one anterior). The
66 dorsal bar consists of two articulating sclerites in *M. clarii* and just one in *M.*
67 *congolensis*. Moreover, the ventral bar of *M. clarii* is posteriorly associated with three long
68 accessory sclerites, while that of *M. congolensis* has two long horns and possesses two
69 posterior accessory sclerites [25,26].

70 Light microscopy of the anterior adhesive apparatus of *M. clarii* (see El-Naggar and
71 Serag [25] and *M. congolensis* (see El-Naggar et al. [26]) revealed two kinds of gland cells,
72 one producing two types of secretion (rod-shaped bodies and spherical bodies) and the other
73 manufacturing irregularly-shaped bodies. With the exception of Kritsky [4], no ultrastructural
74 studies have been conducted on the anterior adhesive apparatus of gyrodactylid parasites.

75 However, SEM has been used to study the head lobes of *Gyrodactylus groschafti* (see El-
76 Naggar [5]), *M. clarii* (see El-Naggar [6]) and *M. congolensis* (see Arafa et al. [27]). In these
77 three gyrodactylids, each head lobe bears a single, ventrally-located adhesive sac provided
78 with emergent papillae which are densely covered with microvilli and perforated by gland
79 duct openings [5,6,27].

80 The present study assesses whether there are any adaptive differences in the anterior
81 adhesive apparatus of *M. clarii* and *M. congolensis* based on their microhabitat. *M. clarii*
82 lives on the gill filaments of *Clarias gariepinus* and are exposed to strong gill ventilating
83 water currents, while *M. congolensis* parasitizes the skin and fins of the same host.

84

85 2. Materials and Methods

86 Specimens of the Nile catfish *Clarias gariepinus* (Burchell, 1822) were caught from the
87 Demietta branch of the River Nile near Mansoura City, Daqahlia Province, Egypt and
88 transported alive to the Faculty of Sciences, Mansoura University. Here, fish were maintained
89 for a few days in an aquarium containing aerated river water at room temperature (25 ± 5 °C)
90 with natural daylight. The catfish (n = 50) were killed by pithing and severing the spinal cord.
91 The gills, fins and scrapings of the skin were removed and placed in Petri dishes containing
92 filtered river water. Gills were searched for *Macrogyrodactylus clarii*, while fins and scrapings
93 of the skin were searched for *M. congolensis* using a dissecting microscope. Some living
94 specimens of both species were flattened between a glass slide and a coverslip and stained with
95 light green and eosin according to El-Naggar et al. [23]. Living and stained flattened specimens
96 (N=10) were examined using light and phase-contrast microscopy with oil immersion, and the
97 different kinds of gland cells were counted.

98 For TEM, specimens of *M. clarii* and *M. congolensis* were washed in distilled water and
99 then fixed in 2.5% glutaraldehyde buffered to pH 7.3 with 0.1 M sodium cacodylate-HCl

100 buffer at 4 °C for 2 h. They were then washed for at least 1 h in several changes of cold
101 buffer (0.1 M sodium cacodylate-HCl containing 3% sucrose and 0.1 M CaCl₂), post-fixed in
102 1% osmium tetroxide in sodium cacodylate buffer at 4 °C for 1 h, washed overnight in the
103 same buffer, then dehydrated using an ascending series of ethanol solutions before transfer to
104 a 1 : 1 mixture of propylene oxide and Spurr resin. Specimens were transferred into gelatin
105 capsules containing pure resin and placed in an oven overnight at 60 °C. Ultrathin sections
106 were cut at 70–90 nm using an LKB NOVA ultramicrotome and glass knives. The sections
107 were mounted on single-hole and 75 mesh coated grids and stained in a solution of 1-2%
108 aqueous or alcoholic uranyl acetate for about 30 min followed by 2-3% lead citrate for 5 min.
109 The sections were examined using a JEOL 100SX transmission electron microscope
110 operating at 80 kV. Measurements of different secretory bodies are based on >10 organelles
111 from electron micrographs.

112 3. Results

113 The head region of both *Macrogyrodactylus clarii* and *M. congolensis* consists of two
114 head lobes. Each bears a single adhesive sac located ventrally at its distal extremity and
115 terminates in a single spike-like sensillum (Fig. 1). The lateral regions of the head contain
116 numerous unicellular glands with their ducts converging on and opening into the two
117 adhesive sacs (Fig. 1). Three kinds of gland cells (G1, G2 and G3) are present in both *M.*
118 *clarii* and *M. congolensis*. Generally, the anterior adhesive apparatus of *M. congolensis*
119 resembles that of *M. clarii* with just minor differences in the number of G2 gland cells. In *M.*
120 *congolensis*, the G2 glands comprise at least 10 cells while in *M. clarii* they constitute only
121 seven cells. The G1 gland cells produce rod-shaped bodies (S1a) and relatively large spherical
122 globules (S1b) (Fig. 1). On each lateral side of the head of both *M. clarii* and *M. congolensis*,
123 there are sixteen G1 cells that are arranged in three groups, one lies lateral to the cerebral
124 region and comprises five cells and the second consists of six cells and lies lateral to the

125 anterior region of the pharynx, while the third comprises five cells and lies lateral to the
126 anterior unbranched region of the intestine. In both *M. clarii* and *M. congolensis*, the G2 cells
127 are found in a single group lying lateral to the posterior region of the pharynx and the anterior
128 unbranched region of the intestine. The G2 cells are larger than the G1 cells and produce
129 irregularly-shaped secretory bodies (S2). In both *M. clarii* and *M. congolensis*, the G3 cells
130 are three in number located lateral to the cerebral region and produce translucent mucoid
131 secretory bodies (S3) (Fig. 1).

132 **3.1 Gland cells**

133 TEM of both *M. clarii* and *M. congolensis* revealed that each G1 gland cell has a nearly
134 spherical nucleus with granular nucleoplasm, conspicuous nucleolus and condensed chromatin
135 (Fig. 2). The cytoplasm is moderately electron-dense and contains abundant granular
136 endoplasmic reticulum (GER), numerous ribosomes (Figs. 2-5), a few Golgi bodies, small
137 electron-lucent vesicles and mitochondria. The dilated cisternae of the GER enclose an
138 amorphous, finely granular material with an electron density slightly higher than that of the
139 basal cytoplasm (Fig. 2). Generally, each fully-developed, rod-shaped S1 body is of high
140 electron density and measures 0.3-0.5 (average 0.4) μm in diameter. The maximum length
141 measured in sections is 4- 6.5 (average 6) μm . These bodies are membrane bounded and contain
142 a finely granular dense matrix in which small particles are embedded within higher electron-
143 dense material (Figs. 2-5). In sections, immature S1 bodies have a greater diameter than that of
144 the fully developed ones (Figs. 4, 6). They measure 0.4-0.7 (average 0.6) μm in *M. congolensis*
145 and *M. clarii* and contain granular material, with electron-density lower than that of the smaller
146 S1 bodies (Figs. 4, 6). Each of the large, immature, less electron-dense bodies and some of the
147 small highly electron-dense bodies are enclosed by a single layer of microtubules, which appear
148 to be parallel with each other and with the long axis of the rod (Figs. 4-6). Some of the less
149 electron-dense S1 bodies contain a peripheral layer of small electron-dense granules and have no

150 bounding membrane (Fig. 4). In cross sections, some fully developed S1 bodies attach to each
151 other, forming large condensed globules with various sizes and shapes (Figs. 2, 4, 5). Their
152 number varies from 3-7 S1 bodies in each globule. In a few sections of *M. congolensis*, some
153 fully formed S1 secretory bodies with peripheral translucent vesicles were detected (Fig. 7).

154 Each G2 cell is enclosed by a layer of fibrous interstitial material. They have an irregularly
155 shaped nucleus with a relatively large, conspicuous nucleolus, granular nucleoplasm, small
156 chromatin patches and nuclear membrane with characteristic nuclear pores (Fig. 8). The
157 cytoplasm is moderately electron-dense, but it is slightly darker than that of the G1 cells. It
158 contains abundant GER, free ribosomes, and numerous Golgi complexes, which in many
159 sections are aggregated (in groups of 2-4) in close proximity to the nuclear membrane (Fig. 9).
160 Each Golgi complex consists of 3-5 narrow parallel cisternae terminating with small and large
161 vesicles (Fig. 9). Both cisternae and vesicles are filled with homogeneous, highly electron-dense
162 material. In sections, the irregularly shaped bodies (S2) have different sizes ranging from 0.7-1.5
163 (average 1.2) μm in diameter. They are abundant and contain granular, highly electron-dense
164 material (Figs. 8, 9). However, in *M. clarii* with higher magnification, each S2 body contains
165 tubular structures with lower electron-density, which are embedded in highly electron-dense
166 ground substance (Fig. 10). In most regions of the G2 cells, fully developed S2 bodies are
167 surrounded by cytoplasm characterized by translucent ground substance (Figs. 8, 10).

168 Each G3 gland cell has a nearly oval nucleus with granular nucleoplasm, conspicuous
169 nucleolus and condensed chromatin patches (Fig. 11). Some GER have dilated cisternae. The
170 mucoid secretory bodies (S3) are abundant, irregularly shaped (1-1.9, average 1.4, μm) and
171 contain granular moderately electron-dense material (Fig. 11).

172 **3.2 Gland ducts and adhesive areas**

173 Ducts of the G1, G2 and G3 gland cells carrying the secretory bodies S1, S2 and S3,
174 respectively, extend anteriorly as cytoplasmic processes where they converge on adhesive
175 papillae through which they open into the adhesive sacs (Figs. 1, 12-21). As the gland ducts
176 approach the adhesive sac, they dilate and become closely packed (Figs. 12,19). At this point,
177 some of the gland ducts are associated with muscle fibers that are present beneath the tegument
178 lining the adhesive sac (Fig. 13). Most ducts of the G1 cells are filled with completely formed
179 rod-shaped bodies (Figs. 12,13,14), but in some sections, a few condensed globules of attached
180 rods are found beside S1 bodies (Fig. 15). There are no microtubules in any of the gland ducts.
181 Each G1 gland duct opens to the exterior via multiple apertures (Figs. 14-16). At the openings of
182 the G1 ducts, five layers, three electron-dense and two electron-lucent (Fig. 16), bound each
183 aperture. The outer layer membrane connects with the surrounding tegument by means of
184 septate desmosomes (Fig. 14). Each one of the multiple apertures allows passage of a single rod
185 (Fig. 14). Although large globules were detected in the terminal portion of the G1 ducts just
186 beneath the multiple apertures, none of them were seen passing through the openings or outside
187 the body (Fig.15). Each of the G2 and G3 gland ducts opens to the exterior by a single aperture
188 (Figs.17-20).

189 The adhesive sac is lined with three types of tegumental layer (st1, st2 and st3) (Figs. 14,
190 15, 17, 22, 24). The first (st1) represents the outer tegumental layer covering the ventral
191 surface of the adhesive papillae surrounding the gland duct openings (Figs. 12, 14, 15), while
192 st2 represents the outer tegumental layer covering the lateral surfaces of the adhesive papillae
193 (Figs. 12, 14, 22). The third type (st3) is the outer tegumental layer of the inner rim of the
194 adhesive sac and connects st2 and the tegumental layer of the general body surface (i.e. the
195 outer surface of the head lobe) (Figs. 12, 22). Comparing the three tegumental layers, the st1
196 layer is relatively thin, electron-dense and has numerous microvilli but lacks secretory bodies
197 (Figs. 14, 15). The st2 layer is highly electron-dense and contains abundant electron-dense

198 bodies (Figs. 14, 22). No cytoplasmic organelles like mitochondria, Golgi bodies, GER or free
199 ribosomes were found in st1 or st2 tegument. The st3 tegumental layer connects with the st2
200 tegument by means of junctional complexes (Fig. 22) and contains a few translucent vesicles
201 containing moderately electron-dense particles (Fig. 24). These vesicles are restricted to the
202 outer region of the tegument. Some electron-dense granular bodies, abundant rod-shaped,
203 electron-dense bodies and a few mitochondria were also seen (Fig. 22 inset). The general body
204 tegument contains abundant translucent vesicles and some electron-dense granular secretory
205 bodies, but no rod-shaped bodies (Fig. 22).

206 No experimental work was performed to study the mechanism of attachment and
207 detachment of the head lobes of *Macrogyrodactylus* species. However, in most sections the
208 terminal portions of G1 ducts, homogeneous particulate material was detected around the S1
209 bodies (Figs. 14, 15) while sections of the terminal portions of G2 and G3 ducts revealed
210 considerable change in appearance of the secretory bodies particularly S2 and S3. The S3 bodies
211 lose their membranes and their secretory components form homogeneous particulate material
212 (Figs. 18,19), while S2 bodies become slightly smaller in size and their particulate components
213 diffuse into the lumen of the duct in-between bodies that are still membrane-bounded (Figs.
214 19,21). Moreover, in the same region, these sections show a network of homogeneous material
215 covering the surface of the adhesive papilla (Figs. 19, 21).

216 TEM revealed the presence of a single sensillum on each adhesive papilla (Fig. 20), in the
217 intervening region between the adhesive sac and general body tegument (Fig. 23) and on the
218 anterior region, which is covered by general body tegument (Fig. 24). Each sensillum has an
219 elongated nerve bulb, which terminates in a single opening through which a single cilium
220 protrudes (Figs. 20, 23, 24). Close to the opening, there is an electron-dense thickening and the
221 lining of the opening is connected with the intervening tegument st3 via desmosomes (Figs. 23,

222 24). The nerve bulb contains neurotubules, electron-dense bodies and mitochondria (Figs. 23,
223 24).

224 4. Discussion

225 This is the first ultrastructural study of the anterior adhesive apparatus of the monogeneans
226 *Macrogyrodactylus clarii* [22] from the gills of *Clarias gariepinus* and *M. congolensis*
227 [23,24] from the skin and fins of the same host. TEM revealed that the anterior adhesive
228 apparatus of both parasites consists of three types of gland cells (G1, G2 and G3). The G1 cells
229 produce rod-shaped bodies (S1) and roughly spherical large globules, G2 cells secrete
230 irregularly shaped, highly electron-dense bodies with tubular contents (S2) and G3 cells
231 manufacture irregularly shaped, mucoid-like secretion (S3). These glands resemble those of the
232 anterior adhesive apparatus of other monogenean parasites [28,29,30]. Previous studies
233 illustrated that congeners in the same microhabitat tend to have similar types of anterior
234 adhesive secretions [7,29,31]. In the present study, the anterior adhesive apparatus of *M. clarii*
235 and *M. congolensis* have the similar morphological features, despite the differences in their
236 microhabitat, with the exception of the number of G2 cells: 10 pairs in *M. congolensis* and 7
237 pairs in *M. clarii*. Morphological similarities, however, do not exclude the possibility of
238 chemical and/or functional differences [30].

239 In *D. amphibothrium*, El-Naggar and Kern [7] found that S1 bodies in the G1 ducts
240 connect with each other by membrane-like structures and a similar feature of interlinking band-
241 like structures was observed between S1 bodies and S2 bodies in *Bravohollisia gussevi* and
242 *Caballeria liewi* (see Wong et al. [13,16], respectively). Also, the bounding membranes of S1
243 bodies in *Entobdella australis* and *Entobdella* spp. (see Whittington and Cribb [29]) showed
244 periodic dense bandings. None of these structures, however, were observed in either *M.*
245 *congolensis* or *M. clarii*. A unique feature of these parasites though is the presence of large
246 globular bodies in the cytoplasm and ducts of the G1 cells, in addition to fully formed S1 bodies.

247 With TEM, it became evident that these globules are aggregations of S1 bodies. There was no
248 evidence that S2 or S3 bodies in *M. clarii* and *M. congolensis* aggregate and coalesce in the
249 cytoplasm of their cells but they become closely packed as they reach the terminal portions of
250 the ducts. Another important feature of *M. clarii* and *M. congolensis* is that S2 bodies contain
251 tubular structures, a feature not reported in any other monogeneans studied by TEM. In addition,
252 the present study indicates that the fully formed rods in *M. clarii* and *M. congolensis* are
253 considerably larger than the S1 bodies in *Entobella* spp. (see Whittington and Cribb [29]).

254 During the early stage of assembly, the large, less electron-dense rods, and some of the
255 smaller highly electron-dense rods of *M. clarii* and *M. congolensis* are enclosed by
256 microtubules. The microtubules disappear when the rods are fully formed and become bounded
257 by membrane. Microtubules have been reported in most other monogeneans studied (see for
258 example Wong et al. [13]) except for monocotylids [11,12] and *Benedenia* spp. [32]. Moreover,
259 the rods of *Monocotyle spiremae* have no bounding membrane and possess an outer electron-
260 dense cortex and a more electron-lucent core [11]. El-Naggar and Kearn [7] suggested that
261 encircling microtubules may play a role in transporting products from different parts of the cell
262 prior to assembly of the secretory bodies. In addition, the microtubules may orientate the rods
263 during their passage from within the gland cells to the lumen of their gland ducts, and help to
264 maintain the parallel arrangement of rods into bundles [7].

265 The monogeneans *M. clarii* and *M. congolensis* resemble other gyrodactylids in that the
266 secretions of the anterior adhesive apparatus open into a single pair of adhesive sacs, one
267 situated antero-ventrally on each of the two head lobes [4]. Other monogeneans, with the
268 exception of gyrodactylids and some monocotylids, have three distinct zones on each side of the
269 head for the release of secretions (see, for example, El-Naggar and Kearn [7]). Such
270 organization into six separate points of contact presumably allows the parasite to be more
271 resistant to detachment caused by water currents [10].

272 In the present study of *Macrogyrodactylus* spp., it has been established that the rod-shaped
273 bodies and roughly spherical large globules produced by G1 gland cells are transported through
274 ducts terminating with multiple apertures. Each aperture apparently permits the passage of only
275 one rod but there is no evidence that the large globules pass through multiple apertures. Multiple
276 apertures were reported in *G. eucaliae*, *Entobdella soleae* and *M. spiremae* (see Kritsky [4]).
277 [2,11], respectively. However, in *D. amphibothrium* and *D. hemiamphibothrium* the rod-shaped
278 bodies are released from ducts with single apertures (see El-Naggar and Kearns [7]). The unique
279 feature of *M. clarii* and *M. congolensis* is that the ducts that carry the rod-shaped bodies also
280 carry larger globules of the same secretion, but the globules were not seen passing through duct
281 apertures. It is possible that the multilayered boundary of one of the small multiple openings
282 dilate to permit passage of the larger globules. Alternatively, the large spherical globules may
283 liquefy or fragment before passing through the multiple apertures. Presence of material similar
284 to the contents of S1 bodies and large globules in the terminal portion of G1 ducts (Figs. 10, 11)
285 supports the latter suggestion. There is also some evidence that the large globules in the G1 cells
286 are composite structures, perhaps made by accumulation of rod-shaped bodies or components of
287 them. If correct, then the globules might escape from the duct openings after disintegration into
288 their small rod-like components. Moreover, in *M. clarii* and *M. congolensis*, the S2 and S3
289 bodies showed considerable change in their appearance inside the terminal portions of the ducts
290 indicating that they are released from duct openings in a liquid form. A similar feature was
291 reported by Kearns and Evans-Gowing [9] who found that the spheroidal secretory bodies
292 associated with the anterior adhesive apparatus of *E. soleae* transform within the duct
293 terminations immediately prior to attachment of the head region.

294 A characteristic feature of *M. clarii* and *M. congolensis* is that each adhesive sac is lined
295 with three types of tegument (st1, st2, and st3) that are different from the general body surface.
296 The first kind (st1) is thickly covered with microvilli, a feature that has been reported in the

297 adhesive areas of many monogeneans [2,4,5,7,9,11,33,34]. These specialized microvilli may be
298 important during attachment of the head lobes by increasing the surface area available for
299 binding the adhesive secretions to the head region. Lyons [34] suggested that these microvilli in
300 *Gyrodactylus* spp. may assist in spreading the adhesive secretion of the head glands over the
301 skin of the host into a thin "tacky" film. The microvilli may help to mix the products of different
302 gland cells, which might have to interact with each other or with water before the sticky
303 properties are developed.

304 Rod-shaped bodies are the most abundant component of the anterior adhesive apparatus of
305 *M. clarii* and *M. congolensis*. These bodies also represent the main component of the anterior
306 adhesive secretions of many monogeneans, which produce two or three types of secretory bodies
307 such as *D. amphibothrium* and *D. hemiamphibothrium* (see El-Naggar and Kearns [7]) and *E.*
308 *soleae* (see Kearns and Evans-Gowing [9]). Furthermore, rod-shaped bodies are the only
309 secretory body recorded in the anterior adhesive apparatus of the monocotylid, *Monocotyle*
310 *spiremae* (see Cribb et al. 11]).

311 The mechanism of attachment of *M. clarii* and *M. congolensis* may involve adhesion of
312 the adhesive sac rim to the host tissues, protrusion of the adhesive papillae by means of
313 associated muscles and release of secretory bodies through gland duct openings. The spike
314 sensillum and other ciliary structures may serve as chemoreceptors that control attachment of the
315 adhesive sacs. In monogeneans, it was suggested that stickiness could be a property of one type
316 of secretory body or could develop by mixing between two types of secretion [2,7,20,35].
317 Interaction between secretory bodies and water [2] or between secretory bodies and host mucus
318 [11] are possible alternative mechanisms.

319 Detachment of the head lobes of *M. clarii* and *M. congolensis* may occur mechanically by
320 contraction of muscle fibres attached to the anterior region [11]. In *E. soleae*, tegument of the

321 adhesive pads may play a part in detachment of the head region, by release of secretory bodies,
322 which are abundant in this layer, or by some other physical or chemical change mediated via the
323 tegumentary membrane [9]. In *M. spiremae*, where only one type of secretion (rods) was found,
324 detachment may involve additional glue, physical detachment by muscle contraction or
325 extrusion of material surrounding the rods [11]. Experimental studies are still needed in this field
326 to determine which secretion is responsible for attachment and how detachment takes place: a
327 potentially lucrative area for industry in relation to binding agents in water.

328 Regarding parasite-host specificity, it has been reported that the epidermal mucous cells of
329 specific fish hosts may influence parasite attachment (see review in Whittington et al. [31]). The
330 anterior attachment region of *Gyrodactylus derjavini* contains mannose-rich glycoproteins,
331 which are implicated in stimulating the alternative complement pathway in the host [36].
332 Specific differences in host fish epithelium and differences in monogenean anterior adhesive
333 chemistry or in the chemistry of the specialized tegument of the anterior adhesive area may all
334 contribute to host specificity amongst monogeneans [30].

335

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449 **Figures**

450 **Fig. 1.** Diagrammatic representation of the anterior adhesive apparatus and anterior region of the
 451 digestive system of *Macrogyrodactylus clarii* (ventral view). aph, Anterior region of the
 452 pharynx; as, adhesive sac; co, cerebral organ; G1, gland cell producing rod-shaped bodies
 453 (S1a) and roughly spherical bodies (S1b); G2, gland cell producing irregularly-shaped
 454 bodies (S2); G3, gland cell producing translucent secretory bodies (S3). gd, gland duct; hl,
 455 head lobe; it, intestine; mo, mouth opening; oes, oesophagus; pph, posterior region of the
 456 pharynx; sp, spike-like sensillum; ui, unbranched region of the intestine.

457 **Fig. 2.** G1 gland cell of *Macrogyrodactylus clarii* containing S1 rod-shaped bodies and large
 458 spherical globules (lg). dS1, Developing rod-shaped bodies; ch, chromatin; GER, granular
 459 endoplasmic reticulum; N, nucleus; Nu, nucleolus; r, ribosomes; S1, rod-shaped secretory
 460 bodies.

461 **Fig. 3.** Cytoplasm of the G1 gland cell of *Macrogyrodactylus clarii* containing fully formed S1
 462 rod-shaped bodies and granular endoplasmic reticulum (GER). r, Ribosomes.

463 **Fig. 4.** G1 gland cell of *Macrogyrodactylus congolensis* containing fully formed rod-shaped
 464 bodies (S1), large globule (lg) and developing S1 (dS1) secretory bodies. Note that the
 465 developing rod-shaped bodies (ds1) have different sizes and are surrounded by
 466 microtubules (mt). Note also that some of the developing S1 bodies contain a peripheral
 467 layer of small electron-dense granules (arrows) and have no bounding membrane. GER,
 468 granular endoplasmic reticulum.

469 **Fig. 5.** G1 gland cell of *Macrogyrodactylus congolensis* containing fully formed rod-shaped
 470 bodies (S1) and large globules (lg) each containing many S1 bodies (arrow). Note the
 471 presence of small translucent vesicles (v) and developing rod-shaped bodies (dS1).

472 **Fig. 6.** G1 gland cell of *Macrogyrodactylus congolensis* showing longitudinal sections of the
473 large developing rod-shaped bodies (dS1) and fully formed rod-shaped bodies (S1). Note
474 the microtubules (mt) associated with dS1.

475 **Fig. 7.** Magnified S1 secretory bodies of *Macrogyrodactylus congolensis* with translucent
476 vesicles (arrow heads).

477 **Fig. 8.** G2 gland cell of *Macrogyrodactylus clarii* surrounded by fibrous interstitial material (fm)
478 and containing large nucleus (N) with conspicuous nucleolus (Nu) and irregularly shaped
479 secretory bodies (S2) surrounded by translucent area (*).

480 **Fig. 9.** Magnified part of G2 gland cell of *Macrogyrodactylus clarii* with Golgi bodies (Go),
481 granular endoplasmic reticulum (GER), ribosomes (r) and irregularly shaped secretory
482 bodies (S2).

483 **Fig. 10.** Magnified S2 of *Macrogyrodactylus clarii* containing tubular structures with lower
484 electron density and surrounded by a translucent area (*).

485 **Fig. 11.** G3 gland cell of *Macrogyrodactylus clarii* containing nucleus (N) with chromatin (ch),
486 dilated cisternae of granular endoplasmic reticulum (dGER) and translucent mucoid
487 secretory bodies (S3).

488 **Fig. 12.** Section through adhesive sac (as) of *Macrogyrodactylus clarii* showing the ventral
489 surface of adhesive papillae (ap) covered with st1 tegument. S1, rod-shaped bodies.

490 **Fig. 13.** Section through adhesive sac (as) of *Macrogyrodactylus clarii* showing muscle fibres
491 (mf) in between S1 ducts. S1, rod-shaped bodies.

492 **Fig. 14.** Adhesive papilla of *Macrogyrodactylus clarii* showing S1 body protruding from its
493 aperture. Note the st1 tegument covering the ventral surface of the adhesive papillae and

494 st2 tegument covering the lateral surface of adhesive papillae. as, Adhesive sac; mi,
495 microvilli; *, homogeneous material around S1 bodies.

496 **Fig. 15.** Duct of G1 gland cell of *Macrogyrodactylus congolensis* containing rod-shaped bodies
497 (S1) and large globules (lg) close to the multiple apertures. Note that the membrane
498 bounding the outer layer of the multiple apertures is connected to the adjacent tegument
499 (st1) by means of septate desmosomes (d) and presence of homogeneous material (*)
500 around S1 bodies. mi, Microvilli; st1, tegument covering the ventral surface of adhesive
501 papillae.

502 **Fig. 16.** Cross section of the multiple apertures of a G1 gland duct of *Macrogyrodactylus*
503 *congolensis* showing that each aperture is bounded by five layers (l), three electron-dense
504 and two electron-lucent. S1, rod-shaped bodies.

505 **Fig. 17.** Terminal portion of G2 gland duct of *Macrogyrodactylus congolensis* carrying S2
506 secretory bodies. f, fibrous layer; mi, microvilli; st1, microvillous tegument.

507 **Fig. 18.** Section through adhesive sac of *Macrogyrodactylus clarii* showing terminal portions of
508 gland ducts carrying S1, S2 and S3 bodies. Note that secretory bodies of S3 bodies form
509 particulate material.

510 **Fig. 19.** Terminal portions of G1, G2 and G3 gland cells of *Macrogyrodactylus clarii* containing
511 S1, S2 and S3 secretory bodies, respectively. Note that the component of S2 bodies (*)
512 diffuse into the lumen of the duct. mf, muscle fibres.

513 **Fig. 20.** Duct of G2 gland cell carrying S2 secretory bodies of *Macrogyrodactylus clarii*. c,
514 Cilium; nb, nerve bulb; nt, neurotubule; St1, tegument covering the ventral surface of
515 adhesive papillae.

516 **Fig. 21.** The terminal portions of G2 gland ducts of *Macrogyrodactylus clarii* showing that the
517 components of S2 bodies (*) diffuse into the duct lumen.

518 **Fig. 22.** Intervening tegument (st3) connecting the sac tegument of type st2 with the general
519 body tegument (gt) of *Macrogyrodactylus congolensis*. Note that st3 tegument and st2 are
520 connected by a junctional complex (j). gb, Electron-dense granular secretory bodies; rb,
521 rod-shaped electron-dense bodies; v, translucent vesicle with electron-dense granule.
522 **Inset:** magnified st3 with abundant electron-dense, rod-shaped bodies (rb), few electron-
523 lucent vesicles with dark granules (v) and electron-dense granular bodies (gb), similar to
524 those in the general body tegument.

525 **Fig. 23.** Section through the terminal part of ciliary sensillum of *Macrogyrodactylus clarii*. c,
526 Cilium; edb, electron-dense bodies; et, electron-dense thickening; m, mitochondria; nb,
527 nerve bulb; nt, neurotubules.

528 **Fig. 24.** Section through ciliary sensillum of *Macrogyrodactylus clarii*. c, cilium; d,
529 desmosomes; et, electron-dense thickening; gt, general body tegument; m, mitochondria; r,
530 root of the cilium; st3, intervening tegument.

531