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Synchrotron science in the UK: NINA, the SRS and Diamond

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Introduction

One contribution of 14 to a theme issue

'Fifty years of synchrotron science: achievements and opportunities'.

Subject Areas:
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The development of synchrotron science over the last 50 years is reviewed from the perspective of the authors' own scientific programmes.

This article is part of the theme issue 'Fifty years of synchrotron science: achievements and opportunities'.

1. Introduction

The application of techniques employing synchrotron radiation now permeates almost all areas of science. Here we give a personal account of how our work in the fields of structural molecular biology, materials and catalytic science developed and evolved using synchrotron techniques. S.S.H. first describes his early work using the NINA Synchrotron Radiation Facility (SRF) at Daresbury and the subsequent development of the Synchrotron Radiation Source (SRS); the growth of the molecular biology programme at the SRS and the increasing involvement of computation and theory are also discussed, as is the expansion of the international usage of the SRS and the transition to the Diamond Light Source. In the later sections, C.R.A.C. shows how early work with the SRS contributed to key areas of materials chemistry and describes the development of the SRS diffraction facilities; the major impact of both the SRS and the Diamond facility on catalytic science is also highlighted.

2. Arriving at the NINA Synchrotron Radiation Facility (SRF)

Having been awarded a J R Ashworth Research Scholarship by the University of Manchester for a project



Photo 1. Manchester team in 1976 at the experimental control station. Standing from left to right are Scott Hamilton, Manolis Pantos and Samar Hasnain. Sitting with Ian Munro are Itzhak Steinberger (a visiting scientist from Tel Aviv) and Paul Brint, a PDRA. (Online version in colour.)

to work in the extended group of Prof Frank Read, FRS (who was a reader at the time), I arrived at Manchester airport on an autumn Sunday evening on 29 September 1974. I was met by Ian Munro and taken to his family home where I lived for a whole week. This moving from home to home enabled me to adjust to the change (that was oceans apart) seamlessly. During the first fortnight, Prof Read asked me to meet various sub-groups of the Atomic and Molecular Physics group covering his own interest ‘electron collisions with atoms and molecules’, atomic/molecular physics undertaken at Jodrell Bank and photophysics/photochemistry. Three weeks later, having visited Jodrell Bank observatory and NINA at Daresbury, I was able to tell Prof Read that I was most excited by what I saw at Daresbury and what I had gleaned from Ian Munro’s excitement. Frank Read generously agreed. Ian became my official supervisor with Scott Hamilton as my additional supervisor. What a lucky combination—the two pioneers who started it all (the journey of synchrotron radiation science in 1967) at Daresbury were my supervisors. Given the isolated nature of Daresbury and difficulties of travelling from Manchester, I moved to Daresbury and stayed at the lovely Hinstock Mount for the rest of the time of my PhD. I was fortunate at the time that the North Beamline at the NINA Synchrotron Radiation Facility (SRF) had some instruments already installed and commissioned, and initial results were coming out.

In addition to Ian and Scott, Manolis Pantos and Malcolm Howells were the PDRAs in the Manchester team with a visiting scientist Professor Itzhak Steinberger from Tel Aviv on his sabbatical leave (see photo 1). Manolis’s enthusiasm was infectious and Itzhak was like a ‘child in a toy store’. Our instrument intercepted the first portion of the beam on the Northern beamline with the other parts shared by the Reading group and others. Malcolm (who went on to contribute significantly to the Brookhaven light source NSLS and Berkeley’s Advanced Light Source, ALS) had wonderfully put together our instrument. We were engaged with trapping organic molecules in rare gas matrices and studying their photophysical/photochemical properties. Anything we touched provided new results. The pioneering spirit was all around both on the North and South beamlines. This small team of enthusiasts (photo 2) with the community were able to put together a case for what became the first dedicated multi-GeV Synchrotron Radiation Source (SRS). Mike Hart played an important part in incorporating hard X-rays option via a superconducting wavelength shifter. When the NINA SRF closed with the switching off of NINA at midnight of 31 March 1977, a number of us went away to other SR facilities—Ian Munro to SSRL (Stanford), Malcolm Howells and Gwyn Williams to Brookhaven (New York), Joan Bordas to EMBL outstation and I to DESY at Hamburg, leaving my 3-year PDRA fellowship with the Manchester team that had commenced in October 1976.



Photo 2. A photograph of the NINA SRF team taken a few hours before the final switch off of NINA on 31 March 1977. At the time, there were 10 user groups coming from the universities of Manchester, Reading, Oxford, Coleraine, Durham, Bristol, Warwick, Leicester, Edinburgh and MRC Cambridge who successfully put the case with the wider potential users community to build the world's first dedicated SR source, the SRS. From left to right: (sitting) John West, Ian Munro, Jeff Worgan and Ken Lea. (standing) Pat Ridley, Iggy McGovern, Bill Smith, Tony Bourdillon, John Beaumont, John Morton, Paul Brint, Samar Hasnain, Robert Pettifer, Joan Bordas, and Tony Cox. (Online version in colour.)

3. From the NINA SRF to the Synchrotron Radiation Source (SRS)

I joined the late Prof Ernst Eckard Koch at DESY in Hamburg as a DESY Fellow where I was able to participate in a variety of experiments ranging from spectroscopy to diffraction of molecular crystals. Koch was again one of the pioneers who together with Ruprecht Haensel and Christoph Kunz had established the synchrotron radiation facility on the DESY synchrotron. During my stay at DESY, the synchrotron radiation team established HASYLAB. On my return to the UK in October 1978, I took a conscious decision to use my physics and synchrotron radiation background at the interface of chemistry and biomedical science, thus joining an interdisciplinary team at Manchester derived from the Chemistry (Dave Garner, FRS) and Medical Biophysics (David Hukins) Departments who had just started the UK's first biological XAFS project working on metalloenzymes and biological calcification. Again, I decided to locate myself at Daresbury where I had the good fortune of being given a temporary office (which became my office for the next 15 years) that was only two doors away from Sir John Pendry, FRS, who had put forward the most comprehensive modern theory of EXAFS (more of this later) [1]. A year later, in 1979, I joined the national effort of establishing the world's first dedicated synchrotron radiation source (SRS) as a full time scientific staff member of the Daresbury Laboratory where I remained until March 2008, having formed a Molecular Biophysics Group in 1989 after returning from a sabbatical in the protein crystallographic laboratories of Charlie Bugg (Birmingham) and Lyle Jensen (Seattle) during the high brightness lattice shutdown of the SRS. I became Max Perutz Professor of Molecular Biophysics at the University of Liverpool in April 2008.

4. Establishment of XAFS as an important structural biology technique at the SRS

The 2-year period 1978–1980 was a steep learning curve getting to grips with X-ray instrumentation (no gratings but radiation-resistant single perfect crystals such as germanium or silicon; mirrors of different size, smoothness and quality and detectors ranging from solid-state devices to ionization chambers), data analysis and interpretation of extracted EXAFS data. I was fortunate, as mentioned above, to have Sir John Pendry two doors away, whom I found most welcoming for a science discussion, prepared to translate difficult theoretical concepts into

simple language for experimentalists such as me. Even though an approximation known as ‘the plane wave approximation’ of the theory was readily usable and requiring little computer time, I immediately grasped the importance of the curvature of the electron wave for accurate structure determination and began to put effort into its full implementation in the form of EXCURVE [2,3] (Norman Binsted, Steve Gurman and Richard Strange played major roles). From John, I also learned an ‘open door policy’ to encourage younger members of the team to come and talk, which I still maintain at the University. Likewise, I was inducted into the new and emerging field of bioinorganic chemistry by David Garner, FRS, who was one of my supervisors for a year of post-doctoral research (1978–1979) and then a great collaborator until the early 1990s when my science interest and approach began to change. In these early years many leading biochemists placed their trust in us with their precious protein samples that they obtained through hard labour in a highly purified form in the hope that we would be able to provide some important structural information that would provide evidence to differentiate between alternate mechanisms. An initial success came from the late Bob Bray of the University of Sussex who had provided more than a gram of purified lyophilized molybdenum containing xanthine oxidase in two forms. We were successful in collecting data and extracting reliable structural information that resulted in the first significant biological XAFS publication from the UK [4].

Towards the end of 1980, Max Perutz approached me to see if I was prepared to help him resolve a serious challenge to his stereochemical mechanism of haem–haem interaction that had come about from some EXAFS work that was conducted in the United States by some leading and highly influential scientists [5]. I accepted the request despite the obvious difficulties (see below).

From the wonderful book “**Science is not a Quiet Life**” By Max Perutz
Extract from Chapter entitled “Haemoglobin Battle”

One day in the mid-seventies, James Lauterbrunner from the famous IGC Laboratories in USA stormed into my room like a Roman gladiator entering the arena and declared that he had disproved my mechanism, because he had found no significant difference between the positions of the iron relative to the porphyrin in deoxy- and oxyhaemoglobin. I offered to show him the difference on my electron density maps, but he was not interested. He had measured the Fe–N distances by EXAFS (Extended X-ray Absorption Fine Structure) and found them to be almost the same in oxy- and deoxyhaemoglobin. Soon afterwards, his results were published in *Nature*, and that vital part of my stereochemical mechanism was dead.

I decided that the only way to prove Lauterbrunner wrong was to repeat his work myself, but EXAFS was a new technique not yet available in Europe. Fortunately, my old friend Jacob Pinder in New York offered to let me prepare my samples in his laboratory and to carry out the EXAFS measurements with me at the Stanford Linear Accelerator, where Lauterbrunner had done his measurements some years earlier. I decided to compare the EXAFS of human deoxyhaemoglobin with that of the deoxygenated picket fence complex, an iron porphyrin synthesized by the Stanford chemist James Collman which mimicked haemoglobin in its ability to combine reversibly with molecular oxygen. Its structure was accurately known, the iron being displaced by $0.399 \pm 0.004 \text{ \AA}$ from the plane of the porphyrin nitrogens and $0.426 \pm 0.004 \text{ \AA}$ from the mean porphyrin plane. I was confident that its EXAFS would match that of deoxyhaemoglobin, which would vindicate my mechanism.

I returned to the guest-house weary from the sleepless night, but much relieved to have done the measurements. I had rejoiced too soon! In the afternoon, Pinder dropped in to bring me the raw EXAFS records; he told me that only Jane had the programs for converting them to interatomic distances, but she would have nothing to do with my data, because Lauterbrunner would have her sacked if he heard that she had collaborated with me. Now I understood why Pinder had tried to keep me away. I lacked the knowledge to devise my own program, and it took me a year until I found out that Samar Hasnain at the Hamburg synchrotron had a suitable program and was willing to evaluate my data.

Max Perutz's view on the James Lauterbrunner (in real life Peter Eisenberger) result was that his theory of a stereochemical mechanism was dead. Typical of him, not knowing the technique, he set about making arrangements for doing the XAFS measurements on a sample prepared by himself. He recruited his friends worldwide to get the measurements done at the Stanford Synchrotron in May 1980 on BL15 and BL23. But Max then faced the problem of data analysis. This luckily brought him to me in late 1980 when I had joined Daresbury as a staff scientist. I was aware of the controversy and had learnt of the difficulties of anyone looking at the data in the USA, for fear of harming their career. In fact, an Englishman who had done his PhD at Stanford was at the EMBL in Hamburg at the time; he could have analysed the data but decided not to, as he did not wish to rule out the possibility of working in the USA.

Over the next nine months, I rigorously analysed the data using the most accurate curved wave implementation of EXAFS theory where it took overnight computation on the best IBM computer available at Daresbury (Daresbury was one of the major national computer centres at the time) to complete a single iteration for a fraction of the XAFS data range. In May 1981, I had the result, which confirmed the original EXAFS structural parameters. This compelled me to think where the problem regarding the lack of movement of iron from the porphyrin plane in Eisenberger's study might originate. I set about looking at all of the chemical porphyrin compounds that had been used for comparison and as standards in the original study and our own. The answer was obvious—Eisenberger had used the triangulation method where the assumption was made that the distance of the centre of the porphyrin plane to nitrogens between the compounds and haemoglobin is transferable. Eisenberger had used a value of 2.045 Å for centre to nitrogen distance rather than the more commonly used value of 2.02 Å. I wrote a detailed letter to Max on 14 May 1981 describing the problem in detail. I received an instant response via a handwritten letter on 18th May expressing his excitement. With some additional data on related compounds collected and analysed, we quickly wrote the paper and submitted it to *Nature* on 22 September 1981. This was just a few months after the SRS had come into operation with its initial energy of 1.8 GeV and two bending magnet beamlines, line 7 for X-rays and line 6 for VUV and soft X-ray primarily for Surface Science experiments. The paper was accepted in December 1981 and published on 11 February 1982 [6]. It is remarkable that both of us were so focused on getting the data analysed and resolving the problem scientifically that we never met prior to the publication. This was remedied by many visits including him staying at our home. On one of my visits to LMB, he mounted a very large MbCO single crystal for the first angle resolved XANES study of a protein crystal using polarized X-rays from the SRS [7]. This early work led to two distinct major contributions: first the realization that multiple scattering events in XAFS needed to be handled accurately and second that the combination of XAFS and crystallography would be very powerful for structure–function studies of metalloproteins, hence giving birth to a combined methods approach that I have continued to develop with new approaches joining the toolbox of structural biology.

Through a BBSRC/MRC grant, we were eventually able to build a dedicated experimental beamline for combined crystallography and single crystal XAFS at the SRS that was opened by Cherie Blair on 28 January 2005 [8–10]. The use of this combined approach has led to a global effort to pursue damage-free crystallographic data collection by using spectroscopic methods to validate redox states. Using the most advanced synchrotrons and X-ray lasers, the serial crystallography approach is being developed for obtaining damage-free structures of functional states of redox enzymes [11–14].

5. Establishment of the SRS as the home for structural biologists

The efforts to make the SRS the home for structural molecular biology dates as far back as the establishment of the NINA SRF. To understand some complexity, it is worth mentioning that the UK's Science Research Council (SRC) established NINA and subsequently the SRS at Daresbury. There were a number of other research councils at the time including the Medical Research Council and the Agricultural and Food Research Council, each jealously guarding their territories

29th November 1972

MRC

Dear Mr Thatcher,

May I confirm the gist of the telephone discussion we had on Monday of this week about the proposals for the joint use by Professor Wilkins and Dr Kendrew of the Synchrotron facility at Daresbury?

When the question was raised in general terms by Mr Jolliffe (his letter of 2nd December 1971 to Dr Lush), we asked all the interested directors of MRC establishments to let us have their views, and the MRC's answer was given in Dr Neale's letter of 22nd June to Mr Jolliffe. The second paragraph of that letter sets out the position at that time in relation to the teams at King's and at the Laboratory of Molecular Biology.

We have now discussed the matter again with Professor Wilkins and with Dr Perutz and they do not feel that the new proposals in any way run contrary to their earlier views. Professor Wilkins regards his contribution to the joint programme as a departmental activity and the MRC will therefore be involved only if he decides to submit grant applications to us for support of this work; I understand that he is, in fact, intending instead to approach the SRC. Dr Perutz is still emphatic that Arndt's use of the facility is in a consultant capacity (in relation to the Daresbury Laboratory) and that no charge should be levied. It may well be that Professor Wilkins may also wish to take up this stance.

NOTE OF A MEETING ABOUT USE OF THE SRF FOR MOLECULAR BIOLOGY

The following were present at a meeting in State House on 22 January 1973:-

Mrs J O Paton
Professor A Ashmore
Dr M F Perutz
Professor D C Phillips
Dr T Vickers
Professor M H F Wilkins
Dr J A Fendley
Mr D Thatcher

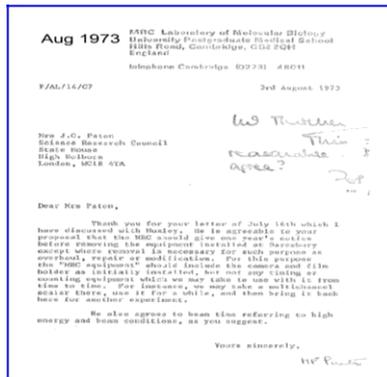
Dr Perutz indicated that the equipment presently under construction by Dr Arndt in the MRC Laboratory of Molecular Biology at Cambridge is essentially a copy of what was developed by Professor Holmes (ex-MRC) under ERNC auspices at DESY in Hamburg. When the MRC equipment is working its use will be shared with Professor Wilkins' group at Kings College (the Department of Biophysics and within it the MRC unit) and Professor Phillips' Laboratory of Molecular Biophysics (MRC grant supported) in the Oxford Department of Zoology. The equipment should be completed in late 1973 but would need to be tested in the SRF from Easter 1973 onwards.

Insert 1. Some correspondence between MRC and SRC.

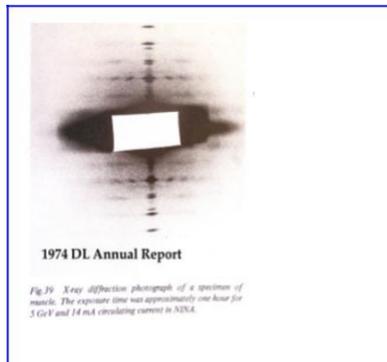
and budgets. Simply put, anyone outside the scope and remit of the SRC had to get their funding council to pay their way for the use of the NINA SRF but more so for the SRS owing to what was a significant investment by a single research council.

Max Perutz (MRC Cambridge), David Phillips (Oxford Molecular Biophysics) and Maurice Wilkins (King's College) represented the interests of MRC at meetings on 22 January and 3 August 1973—inserts 1 and inserts 2. Fibre diffraction was identified as the main beneficiary and as such Hugh Huxley was nominated to coordinate the activities at the NINA SRF. Huxley was able to obtain impressive static diffraction pictures from frog muscle (inserts 3) in early 1974 and was able to progress towards initial time resolved muscle diffraction using this synchrotron source before the closure of NINA on 31 March 1977. In 1978, when Joan Bordas moved to EMBL Hamburg, where fibre diffraction and XAFS instruments had been located on the storage ring DORIS, Hugh Huxley joined in the effort. He only returned to the SRS in the mid-1980s. Joan returned to Daresbury as the head of MRC's Structural Biology Laboratory in 1983. At this time MRC also decided to build a dedicated beamline 2.1 for biological solution scattering and fibre diffraction.

John Helliwell, who was a DPhil student at Oxford with Dr Margaret Adams, attended the 10th IUCr Congress in Amsterdam in August 1975 where he heard Keith Hodgson (Stanford) talk about some early crystallographic experiments performed at the 3.7 GeV SPEAR storage ring at Stanford [15]. When he asked his supervisory team to go to Stanford to gain experience, he was taken to David Phillips who told him about the existence of the Daresbury Synchrotron Radiation Facility. This turned out to be good fortune for the development of crystallographic activities. It also clearly showed how important the 1973 meeting was at which David Phillips was present. John obtained his beamtime on the NINA SRF in December 1976 but the tests were unfortunately



Insert 2. Max Perutz identifying Hugh Huxley as the key driver from MRC.



Insert 3. Muscle Diffraction obtained by Huxley *et al.* at NINA SRF.

inconclusive, primarily due to insufficient intensity and the operating mode of NINA—one would have to wait until the SRS storage ring came on line, providing steady intensity. In the meantime, elsewhere in Europe, Roger Fourme had put together a dedicated facility for protein crystallography on the positron storage ring DCI at LURE using an electronic detector [16], which began to attract users, including some from the UK, as the news of possible gains of up to 20 over a 24 kW Elliott GX6 rotating anode spread among the community. It attracted groups from Oxford led by Louise Johnson in which Dave Stuart, FRS and Keith Wilson were also involved [17], and MRC Cambridge led by Max Perutz [18].

With the closure of NINA SRF in spring 1977, the construction of the SRS began. On the X-ray beamline, XAFS (station 7.1: Greaves/Hasnain), fibre diffraction (station 7.2: Watson Fuller), topography (station 7.5/7.6, the longest beamline on a synchrotron for some time: Brian Tanner) and X-ray interferometry (station 7.4: Michael Hart) were planned and built. Neville Greaves led the first publication from the SRS using the XAFS station 7.1 in November 1981 [19]. Watson Fuller, who was a Professor of Biophysics at Keele University, negotiated a lectureship position jointly funded by Daresbury and Keele and advertised the position so that the individual could take the responsibility for station 7.2 as a station scientist. Fortunately, John Helliwell decided to apply and was appointed to this important job; he was then able to steer the design of this important station to include both fibre diffraction and protein crystallography. The versatility of the instrument provided evidence for many crystallographic groups to join the UK's effort of SR structural biology while providing some exciting science related to fibre diffraction [20].

While line 7 was beginning to produce its first results, I started working on a plan to develop beamline 8 where the source properties were much superior to the initial beamlines as the source was at an upstream point of an even-numbered magnet. The beamline was to provide XAFS facilities for dilute systems, particularly biological systems and solution scattering/fibre diffraction. Hugh Huxley, among others, was also involved in detailed specification of the SAXS component of the beamline. The beamline received a real boost with the arrival of the Dutch when NWO signed an agreement in 1982 with SERC to fund this beamline and its two experimental stations. Things began to move rapidly. In 1983 Joan Bordas arrived as part of the MRC signing a cooperation agreement with the SERC for building a biology support laboratory and another dedicated beamline for SAXS/muscle diffraction. SERC funded several experimental stations on the first superconducting wavelength shifter where crystallographic station 9.6 was to be installed; this eventually helped solve the foot and mouth disease virus structure [21] (capturing the national television high spot at 9 pm News in 1989) and F1-ATPase structure [22] that brought the first Nobel prize to the Synchrotron world in 1997 to Sir John Walker from MRC LMB [23]. My own efforts to combine all of the X-ray techniques (XAFS, SAXS and crystallography) came to fruition at the end of the first decade of the SRS [24–26] on iron transport protein, transferrins. This has been the integrated approach of my career since then—fostered by the interdisciplinary environment of the SRS where scientific and technical approaches had no boundaries—with the only important aspect of the enterprise being the scientific question.

6. Collaborative Computational Projects (CCPs) helped to expand the community and science

The SRC's Science Board approved the CCP programme proposed by the Atlas Laboratory (which became a division of Rutherford laboratory in 1975) in October 1973 with the following aims:

- to provide rapid interchange of information in the selected area of study
- to collect, maintain and develop relevant items of software
- to encourage basic research in a given area
- to disseminate information by organizing symposia and workshops

A CCP steering panel was established with Prof Phil Burke, FRS and Prof John Murrell, FRS (Sussex) as members. CCP1 (Quantum Chemistry) was initiated in February 1974 with John Murrell as the Chair. Membership included Prof Ian Hillier (Manchester) and Martyn Guest and Vic Saunders from ATLAS. Scientific results from the NINA SRF and the anticipated science from the SRS became a stimulus for a number of new CCP projects. The Science Research Council (SRC) agreed in 1976 that 10 posts supporting science board work should move from Rutherford to Daresbury. Phil Burke played a crucial role in the development of theory and CCPs. He had a joint appointment with Queen's University and the Daresbury Laboratory as Head of the Theory and Computational Science Division from 1977 until 1982 when he returned to Belfast full time. In September 1977, Vic Saunders, Martyn Guest, Mike Elder and Pella Machin moved to Daresbury; all played a major role in the success of CCPs. During the construction phase of the SRS, eight CCP projects were funded and established, four of which were directly linked to the SRS, namely CCP2 (atomic and molecular processes, 1978, Mike Seaton and Phil Burke), CCP3 (Surface Science, 1979, John Pendry, Tom Grimley and Martin Prutton), CCP4 (X-ray Diffraction and Crystallography, 1979, David Phillips, Tom Blundell, Mike Elder and Pella Machin) and CCP9 (electronic structure of solids, 1981, Balaza Gyorffy and Volker Heine). Each of these had and are continuing to have a major impact on science and access to methodology by a much larger community than would otherwise have been possible. In the context of structural molecular biology, the success of CCP4 is only matched by 'user friendly' facilities at SR centres. CCP4 continues to support collaboration between researchers working on methods and software development for protein crystallography in the UK. It has expanded to become a global example of collaboration and has been one of



Photo 3. Location of CCP4 usage (yellow) and regular workshops/schools (red). (Online version in colour.)

the key contributory factors in the success of biological crystallography. The world map showing CCP4 usage and location of training workshops that are held neatly illustrates this (photo 3).

7. Bringing the international community together

During 1978–1980 I was getting to grips with XAFS instrumentation, theory and analysis. David Garner was asked in the summer of 1980 by the Royal Society of Chemistry to organize a workshop for the chemistry community so that the technique would become more widely accessible to that community. I was acutely aware that EXAFS groups worldwide were working on similar problems (theory, instrumentation and data analysis packages) in this rapidly developing field. It was thus an opportunity to bring together experts from Europe and the United States. Daresbury was good at organizing focused meetings called ‘study weekends’, which were held over a weekend. After some discussions, Dave agreed to make it an international workshop/conference and that if we were to include ‘inorganic systems’ in the title, the RSC would be happy. This was needed as the request had come from the Inorganic Division of the RSC. We thus called the study weekend ‘EXAFS for Inorganic Systems’. With a modest contribution from the RSC and strong backing from the Daresbury directorate, we were able to include three speakers from the USA (Peter Eisenberger from Bell Labs, Ed Stern from Seattle and Steve Cramer from Exxon), three from continental Europe (Peter Rabe from DESY/Keele, Antonio Bianconi from Rome and Alain Fontaine from Orsay) and a number of speakers from the UK (John Pendry, the late Robert Pettifer, David Norman and myself). It was sufficiently successful and filled a much-needed gap, so it was spontaneously decided to make this a conference series that is still growing. Daresbury Laboratory published the proceedings of the meeting [27]; and the conference is held every 3 years now. The last conference in this series was held in Krakow, Poland on 22–27 July 2018 with some 500 delegates. In 1990, it returned to the UK when I organized the conference in the lovely campus of York.

The availability of synchrotron radiation provided a real boost to biophysical methods in the early 1980s. Following a study weekend organized by Greg Diakun in 1984, the first international conference on ‘Biophysics and Synchrotron Radiation (BSR)’ took place in Frascati in 1986. In 1988 I, together with Joan Bordas, organized the second conference in the series in the beautiful city of Chester only 15 miles away from Daresbury with generous support from the International Union of Crystallography (IUCr). The conference produced a well referenced book, ‘Synchrotron



Photo 4. Delegates of the 9th International BSR conference jointly organized by Louise Johnson and Samar Hasnain in Manchester 13–17 August 2007. From left to right S.S.H. is fourth with Tom Blundell. Louise Johnson and Hans Deissenhoffer in the front row. (Online version in colour.)

Radiation and Biophysics’, published by Ellis Horwood, to which many of the synchrotron pioneers (Ian Munro, Roger Fourme, Keith Hodgson, Louise Johnson, Janos Hajdu, Watson Fuller, John Helliwell, David Garner, Wayne Hendrickson, Heinrich Stuhrmann, Yoshiyuki Amemiya, Malcolm Howells, Ed Rubenstein, Ron Burge, etc.) contributed authoritative chapters. I was pleased to bring this conference back to the UK in 2007 when I had the pleasure of chairing the 9th conference in the series with one of my mentors, the late Dame Louise Johnson, FRS (photo 4). The conference attracted some 300 delegates with a number of Nobel laureates (Venki Ramakrishnan, Hans Deisenhoffer and Hartmut Michel) as plenary speakers. Other plenary speakers included Tom Blundell, David Stuart, Keith Hodgson, Janos Hajdu, Dmitri Svergun, Malcolm Irving and So Iwata. The 13th conference in the series will be held in Shanghai 21–24 September 2019.

8. 25 Years of synchrotron research at the SRS

To mark 25 years of the SRS, a celebratory science event was organized on 12 September 2005, combining it with the annual UK SR users meeting. The purpose of the occasion was not only to highlight the achievements but also to recognize the collaborative spirit which the SRS helped to engender in the scientific community. In addition to many of the UK pioneers, several major SR facilities (ESRF, SPring-8 and SSRL) were represented by their Directors and others. The growth of SR science around the world is a testimony to the collaborative spirit fostered by the SRS community.

I helped to assemble the scientific programme with a small advisory committee (Sir Tom Blundell FRS, Phil Burke FRS, David Garner FRS, Sir John Pendry, FRS, and Michael Woolfson, FRS). The programme was organized under 6 themes, namely CCP, Theory-SR science interaction, Materials Science, Advances in SR sources and Instrumentation, Structural Biology and a forward-looking session. The 2-day event was held in Manchester with a visit to the laboratory on the evening before. There was a reception for many of the pioneers of synchrotron radiation, who had travelled far and wide at their own expense, including the heads of the Anglo–Dutch collaboration, Dr Guy Luijkcx and Dr H Weijma. The photograph below (photo 5) shows many of the leading delegates who attended the special reception.



Photo 5. A group photo of many of the eminent scientists associated with the SRS and the SR world. In the front row Samar Hasnain, Herman Winick, Louise Johnson, John Pendry, Ian Munro, Mike Chesters (Director at the time), Hugh Huxley, Akira Kira, Michael Woolfson, Gerd Materlik and Alan Leadbetter are clearly visible. Pat Ridley, John West, Phil Burke, John Inglesfield, John Evans. Keith Hodgson, Richard Catlow and Bill Stirling are visible in the third and fourth rows. (Online version in colour.)



Photo 6. SRS gives birth to Diamond which has become an exemplar facility for Structural Biology. (Online version in colour.)

9. From the SRS to Diamond

The SRS closed in 2008, having pioneered many of the techniques and research areas in the X-ray region. These are continuing to thrive at the Diamond Light Source (photo 6), and pioneering new



Photo 7. Installation as the Max Perutz Chair of Molecular Biophysics at Liverpool University. Front row from left to right are Kyosho Nagai, Tom Blundell, Robin Perutz, S.S.H., Yasmeen Hasnain, Giorgina Ferry, Richard Henderson and Louise Johnson. In the second row Roger Fourme, John Collinge, Colin Nave, Salman Hasnain, Keith Hodgson, Simon Phillips and Michel van der Rest among others are present. In the final row first from right is Michael Woolfson. (Online version in colour.)

frontiers. My inauguration as the first holder of the Max Perutz chair of Biophysics at Liverpool University took place in September 2008 (photo 7) with opening of the Barkla X-ray Laboratory of Biophysics in July 2011.

Diamond has broken new ground in rapid data collection, on-the-fly data processing and remote access. It was the first synchrotron centre to extend its structural biology capabilities to include a national cryo-EM facility under the same roof utilising the same infrastructure for access, user support and scheduling. It clearly has become an exemplar structural biology centre encouraging several other leading synchrotron centres to include cryo-EM in their structural biology toolkit [28,29]. Below, we will also see how it has made major contributions to the development of materials and catalytic science.

10. Synchrotron radiation, materials chemistry and catalytic science

Techniques based on synchrotron radiation have had a major impact on the fields of materials chemistry and catalytic science. In the sections which follow, a personal account is presented of C.R.A.C.'s involvement in these fields, together with a discussion of the likely future developments. I will discuss first my early work with the SRS, which focused on the application of EXAFS to defective ionic materials; next the development at the SRS of the diffraction facilities.

From the 1990s onwards I became increasingly involved with the harnessing of Synchrotron techniques in catalytic science initially with colleagues in the Royal Institution, but more recently with the team in the UK Catalysis Hub. I hope that this account illustrates how synchrotron based techniques have grown over the decades from specialized niche applications into core and crucial experimental methods in mainstream areas within chemistry and materials science.

11. EXAFS and the SRS

I first became aware of the potential of synchrotron radiation for my science in the late 1970s when my research programme had a strong focus on disordered ionic materials—both halides and oxides—for applications in solid-state electrochemistry. Much of my work was in computer modelling which was then emerging as a powerful technique in materials chemistry for developing models of structural and dynamical properties of materials and in particular of defect and dopant structures and energies. However, I had a strong interaction with experiment— particularly the group of Alan Chadwick at Kent which was at the forefront of experimental studies of ionic mobility in solids. At Alan’s instigation, we attended a ‘Study Weekend’ at Daresbury where we learned of the EXAFS technique and it was clear that EXAFS had huge potential in our field, by providing unique information on local structural properties in disordered solids. We began a very fruitful interaction with Neville Greaves at Daresbury and shortly after the SRS was opened, we succeeded in winning a grant from the SRC, ‘Synchrotron Radiation Facilities Committee’ for EXAFS studies of ionically conducting solids. We recruited a talented post doc, Lee Moroney, and together with her and Neville, we faced the challenges of collecting and analysing data from the EXAFS beam line (station 7.1).

Despite the difficulties, it was immensely exciting working on this pioneering facility and our science made rapid progress. Developments in data analysis software were crucial and our success owed much to our interactions with Norman Binsted who was developing effective EXAFS data reduction procedures. I will pick out just two highlights of this highly productive period. The first concerned rare earth doped fluorite (CaF_2)—a widely studied system in the 1970s and 1980s owing to its ionic conducting properties and to applications in laser technology. There had been a long-standing debate about the local structure around the dopant ions and EXAFS data when combined with computational modelling demonstrated that the dominant structure was a beautiful octahedral dopant cluster surrounded by a cloud of fluoride ion interstitials. This work, published in *Nature* [30] and highlighted on the front cover of the journal (reproduced in figure 1) clearly demonstrated the power of the technique, especially when combined with computational modelling. The second was a study of yttrium stabilized zirconia [31]—among the most intensively studied ceramic materials, owing to its applications as a structural ceramic and as an oxygen ion conducting solid. Yttrium ions replace those of zirconium and are compensated by oxygen vacancies; and the location of the latter with respect to the dopant had been a matter of controversy. A careful and detailed analysis of the yttrium and zirconium EXAFS data showed conclusively that the vacancies occupied not the nearest, but the next nearest oxygen site with respect to the dopant. The conclusions of the EXAFS analysis were again supported by computer modelling.

Many others contributed to the developing field of X-ray spectroscopic studies of complex and disordered materials and the technique is of course now standard in materials chemistry. The early work on the Daresbury station 7.1 played a very important role in this development.

12. The development of diffraction facilities

Although the first successful applications of SR techniques in materials chemistry were in the exploitation of X-ray spectroscopy, it was clear that powder diffraction had great potential for the field. In the 1970s, high resolution powder diffraction (HRPD) using neutron sources had had a major impact, owing to the possibility of using Rietveld techniques enabled by the Gaussian line shapes of neutron PD reflections; structures as complex as those of zeolites had been solved.

⋮

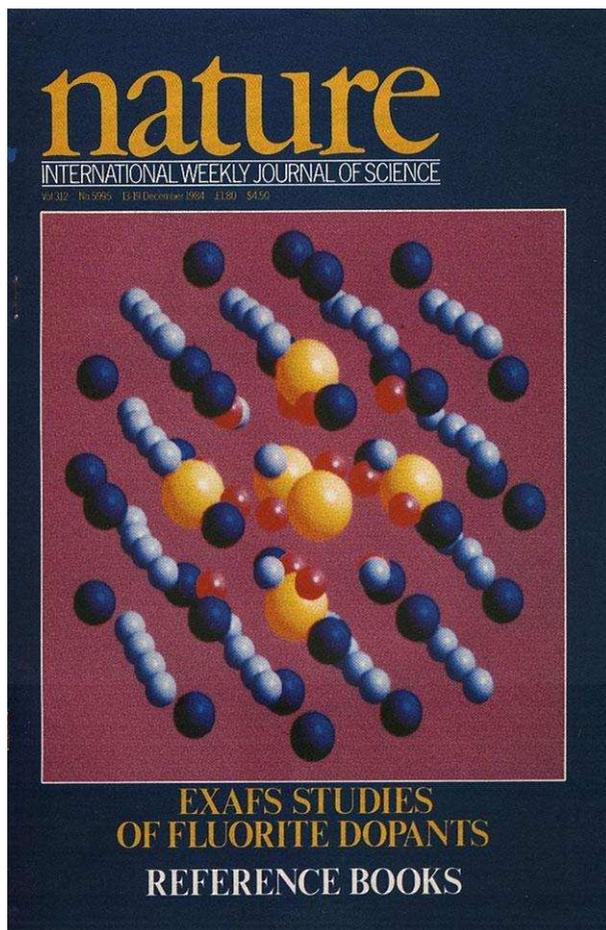


Figure 1. Front cover of Nature illustrating dopant interstitial cluster in rare-earth doped calcium fluoride. (Online version in colour.)

Synchrotron based techniques presented exciting opportunities as the peak shapes, although more complex than those of neutron data, can nevertheless still be parametrized; and they have the additional advantage of being narrow, due to the high collimation of the synchrotron beam, thereby minimizing peak overlap—the intrinsic difficulty with powder data.

The potential of SR based PD had been recognized early in the development of the instrumental programme on the SRS, with the construction of a high resolution instrument (station 9.1 on the wiggler beam line). In 1985, I moved to a joint professorial appointment between Keele University and The Daresbury Laboratory, where my role was to assist in the development of the PD instrumentation and community. I was given great support and encouragement from the then Vice-Chancellor of Keele, Brian Fender, and we were fortunate enough to recruit Andy Fitch (now leading the powder diffraction programme at ESRF) as a lecturer who brought valuable and extensive experience of neutron based PD. The team at Daresbury included Peter Hatton and Graham Bushnell-Wye, and was soon augmented by Bob Cernik, Phil Pattison and Simon Clark (photo 8). We were also helped by the expertise and vision of Michael Hart who was then playing a crucial role in developing the instrumental and scientific programme of the SRS.

It soon became clear that the instrument on beam line 9.1 was being required to function in too many modes, including operating both as an angular and energy dispersive (ED) diffractometer.



Photo 8. Shows the X-ray diffraction team at the SRS circa 1989: bottom row (from left) Phil Pattison, Bob Cernik, Simon Clark, Mike Miller and Killian O Reilly, top row (from left) Manolis Pantos, Dave Laundy, Richard Catlow, Graham Bushnell-Wye, Andy Fitch, Alf Neild, Eric Doohryee, one of our detector group summer students and Ian Sumner. (Online version in colour.)

The team therefore developed and implemented a plan for a suite of three instruments: station 9.1 was optimized for high resolution angle dispersive studies; a second station (9.4—again on the wiggler beam line) was constructed for energy dispersive work; and a third (station 8.3), for intermediate resolution angular dispersive XRPD. The latter had an intriguing design, proposed by Michael Hart with a set of long collimators between the sample and the detector (often referred to as the Hart-Parrish design), which improved line shape and resolution.

The plan proved to be a great success. Andy Fitch's expertise in structure solution using Rietveld techniques helped to develop station 8.3 into a workhorse diffractometer which solved a large number of crystal structures; there were also crucial contributions from Bob Cernik and an early success was the solution by Cernik *et al.* [32] of the structure of cimetidine ($C_{10}H_{16}N_6S$)—a powerful histamine antagonist—which clearly showed the ability of synchrotron based powder techniques to solve complex structures. (9.1) operated successfully as a high resolution instrument and was reserved for more specialist applications including anomalous dispersion experiments; while 9.4 proved highly successful as an ED diffractometer and made a key contribution to the emerging field of kinetic crystallography owing to the ability to collect data very quickly in the ED mode.

The three stations continued to operate throughout the lifetime of the SRS and a further account of their development and achievements is given by Cernik [33]. They were followed in the 1990s by a microcrystalline single crystal diffractometer in a project led by Bill Clegg, of which I was co-investigator and which is discussed in detail in Clegg's article in this issue. With these

instruments, synchrotron based diffraction at the SRS made a marked and important contribution to structural materials science.

13. Catalysis and the Royal Institution

In 1989, I moved to a professorial position at the Royal Institution where, together with the then Director, Sir John Meurig Thomas—one of the leading figures worldwide in catalytic science—we began to explore the potential of SR techniques in studying catalytic materials. EXAFS had been used to considerable effect by John Evans and others in homogeneous catalysts, but application to heterogeneous catalysis was less common. Our work centred around microporous catalysts, both zeolites and aluminophosphates, and was greatly assisted by Gopinathan Sankar, who had recently joined the team from CNR Rao's group in Bangalore. An early success was our study of nickel zeolite Y [34]—a widely studied catalytic material—where by combining results from separate XRD and EXAFS experiments we were able to characterize in detail the local environment of the nickel, located within the cages of the microporous crystal structure of the zeolite. The next step was to combine XRD and EXAFS in one experiment. The Daresbury team led by Greaves, Dent and Derbyshire successfully developed techniques in which diffraction and spectroscopy data could be measured simultaneously. Progress in detector design and technology reduced the data collection time and it now became possible to monitor the evolution of both local and long-range structures during a solid-state reaction. The RI/Daresbury team was at the forefront of exploiting these developments, with a pioneering study of the conversion of the mineral aurichalcite to a copper catalyst, which was able to follow the decomposition of the mineral and the growth of the metal particles during the reaction [35].

Other highlights from this very productive period relate first to the location of organic templates within microporous solids. Templates are used in the synthesis of these materials as they can direct the structures towards specific architectures. In the 1990s, Dewi Lewis and Dave Willock developed '*de novo*' design methods for the prediction of templates for specific architectures and successfully predicted a template for the synthesis of a microporous aluminophosphate—DAF-5—shown in figure 2. Following the synthesis of the solid, the structure was determined using the recently developed microcrystalline diffractometer on station 9.8, referred to earlier, and the location was revealed as shown in the figure. It was exactly as predicted by the computational modelling [36].

Another intensively studied system was the zeolite 'TS1' based on an all silica zeolite—silicalite—in which approximately 1% of the Si is replaced by Ti and which is an extensively used industrial oxidation catalyst. It also proved possible to develop catalysts with similar functionality by taking 'mesoporous' silicas—materials with pore dimensions in the 30–50 Å range (as opposed to microporous materials which are typically in the 4–12 Å range) and grafting tetrahedral Ti species on their internal pores. In a series of studies, EXAFS was able to confirm in detail the structure of the active site. Computer modelling also probed both the structure and mechanisms involved in using these catalysts in epoxidation reactions using H₂O₂ as oxidant. The predicted structures shown in figure 3 agree accurately with the results of the *in situ* EXAFS data analysis. The computer modelling was also able to elucidate the reaction mechanism, so that by this combination of modelling and EXAFS, the full catalytic cycle was understood at the molecular level [37], as illustrated in figure 4.

14. Synchrotron radiation and the UK Catalysis Hub

In 2013, Graham Hutchings, Matt Davidson, Chris Hardacre and I, after extensive discussions with EPSRC, successfully bid for funds to establish the UK Catalysis Hub involving a coordinated and comprehensive programme of catalytic science in the UK. Importantly, this national network comprising more than 40 university teams, has a physical base and hub in the Research Complex at Harwell (RCaH) on the UK Harwell Science and Innovation campus, which hosts the major facilities including the Diamond Light Source, the ISIS Neutron Source and the Central Laser

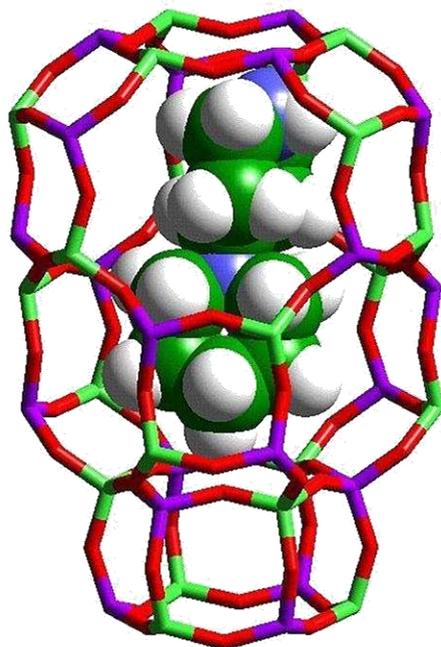


Figure 2. Predicted and experimentally determined structure of 4 piperidinopiperidine inside the pores of DAF-5. (Online version in colour.)

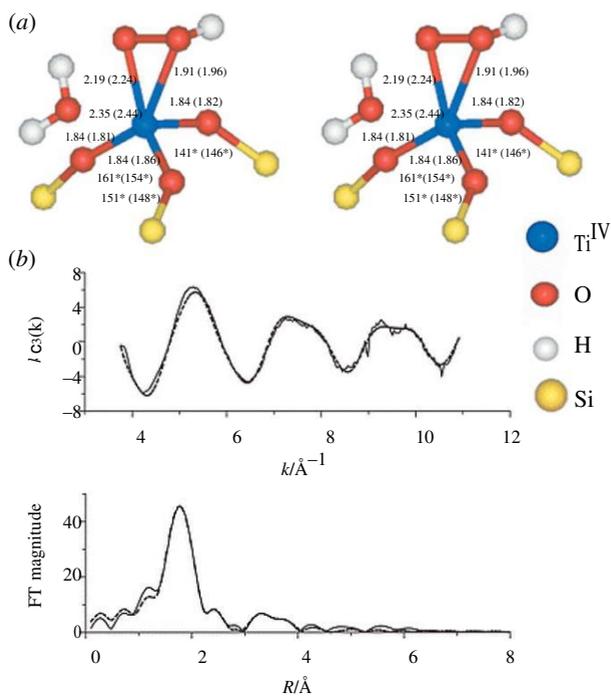


Figure 3. Intermediate in the TS1 oxidation catalyst created by reaction of the titanium centre with hydrogen peroxide. (Online version in colour.)

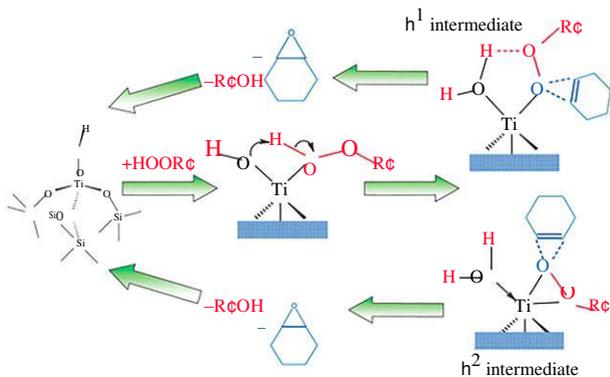


Figure 4. Proposed epoxidation mechanism for Ti micro/meso-porous catalysts for alkene epoxidation. (Online version in colour.)

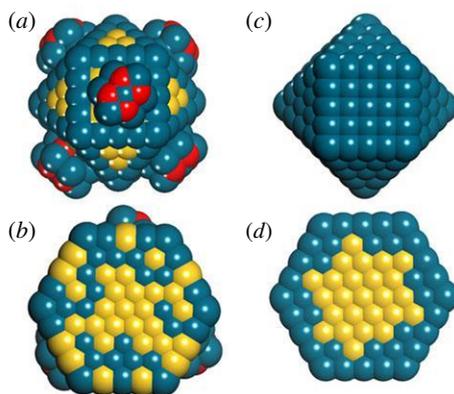


Figure 5. Schematic of restructuring of Au/Pd nanoparticles during CO oxidation as revealed by combined EXAFS/DRIFTS. Initial state of catalyst (a, showing external and b, giving cross-section) has PdO on the surface of NPs with an Au rich core and an AuPd alloy exterior. The final state (c,d) has a gold core, followed by an Au/Pd alloy and a top layer of Pd. (Online version in colour.)

Facility. The Hub has made and continues to make effective use of all these facilities. The use of X-ray spectroscopy on the Diamond beamlines, led by Emma Gibson and Peter Wells in collaboration with scientists from Diamond, has been one of the major features as illustrated by two recent examples.

The first relates to supported nano-metallic catalysts which are extensively used in heterogeneous catalysts. Bimetallic Au/Pd nanoparticulate systems have diverse catalytic functionality and have been particularly widely studied. In a series of experiments Gibson *et al.* [38] investigated how the structures of such nanoparticles evolve during CO oxidation catalysis. The work built on the development of combined *in situ* EXAFS with DRIFTS on Diamond Beam Line 18 and by piecing together the evidence from the two techniques, it was clear that the nanoparticle undergoes extensive restructuring during the catalytic cycle as illustrated in figure 5. Interestingly, during the cycle, the gold buries into the interior of the particle giving a gold-core, palladium-shell structure.

The second example relates again to nano-structured catalysts: in this case, gold on a carbon support, which pioneering work of Hutchings had shown to be effective for the catalytic conversion of acetylene to vinyl chloride—a key step in the production of PVC. The catalyst has now been commercialized by Johnson Matthey and is replacing mercury based catalysts, which

have been widely used in China and which have substantial associated environmental problems. It had been generally assumed that the active catalyst comprised Au nanoparticles. However, *in situ* EXAFS clearly showed during the catalytic operation, the gold is predominantly present as single gold cations. Computer modelling work then demonstrated a plausible mechanism for the catalytic cycle based on gold cations as the active site [39].

Many other examples could be given of the crucial role of *in situ* X-ray spectroscopy in the wide ranging science of the Catalysis Hub. Another very notable development has been the use of tomographic imaging of real industrial catalysts led by Beale using both facilities at Diamond and ESRF [40]. The role of these techniques in catalytic science has been amply demonstrated in extensive work of Weckhuysen and colleagues [41].

15. The future

Synchrotron based techniques are now integral components of biomolecular, materials and catalytic science. Future developments in sources will offer exciting new opportunities in time resolved structural science and in micro-focus experimentation. The power of *in situ* studies will continue to grow as well as the continuing spectacular developments in imaging of real systems complex materials and catalysts. There will also be continued rapid growth in the combination of synchrotron measurements with other spectroscopies and techniques. For structural biology, synchrotron X-ray crystallography will remain the single most important tool for proteins and multiple-protein complexes with molecular weight less than 200 kD. Nearly 90% of the structures in the protein data bank are for macromolecules less than 200 kD [29]. cryoEM is likely to become the most dominant structural biology approach for systems greater than 200 kD and for difficult-to-crystallize membrane proteins. For the latter, XFEL crystallography will also play a significant role with a pole position for providing damage-free and time-resolved structures as exemplified by recent examples of photosystem II [14] and retinol isomerization [42,43]. Synchrotron-based crystallography will remain unique at providing structures at a resolution that provides details at the chemical level necessary to define the mechanism of an enzyme or processes such as electron transfer, bond formation and breakage. Synchrotron-based serial crystallography is beginning to emerge and is likely to play an important role in kinetic crystallography at sub-seconds to milli-seconds time scale. XFEL sources also offer exciting opportunities for probing mechanisms of catalytic reactions. Light sources will continue to illuminate the science of biomolecules, complex materials and catalysts for the coming decades.

Data accessibility. This article has no additional data.

Competing Interests. We declare we have no competing interests.

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References

1. Lee PA, Pendry J. 1975 Theory of the extended X-ray absorption fine structure. *Phys. Rev. B* **11**, 2795–2811, 56–63. (doi:10.1103/PhysRevB.11.2795)
2. Binsted N, Strange RW, Hasnain SS. 1992 Constrained and restrained refinement in EXAFS data analysis with curved wave theory. *Biochemistry* **31**, 12 117–12 125. (doi:10.1021/bi00163a021)
3. Binsted N, Hasnain SS. 1996 State-of-the-art analysis of whole X-ray absorption spectra. *J. Synchrotron Radiat.* **3**, 185–196. (doi:10.1107/S0909049596005651)

4. Bordas J, Bray RC, Garner CD, Gutteridge S, Hasnain SS. 1980 X-ray absorption spectroscopy of xanthine oxidase. The molybdenum centres of the functional and the desulpho forms. *Biochem. J.* **191**, 499–508. (doi:10.1042/bj1910499)
5. Eisenberger P, Shulman RG, Kincaid BM, Brown GS, Ogawa S. 1978 Extended X-ray absorption fine structure determination of iron nitrogen distances in haemoglobin. *Nature* **274**, 30–34. (doi:10.1038/274030a0)
6. Perutz MF, Hasnain SS, Duke PJ, Sessler JL, Hahn JE. 1982 Stereochemistry of iron in deoxyhaemoglobin. *Nature* **295**, 535–538. (doi:10.1038/295535a0)
7. Bianconi A, Congiu-Castellano A, Durham PJ, Hasnain SS, Phillips S. 1985 The CO bond angle of carboxymyoglobin determined by angular-resolved XANES spectroscopy. *Nature* **318**, 685–687. (doi:10.1038/318685a0)
8. Cianci M *et al.* 2005 A high-throughput structural biology/proteomics beamline at the SRS on a new multipole wiggler. *J. Synchrotron Radiat.* **12**, 455–466. (doi:10.1107/S0909049505009131)
9. Arcovito A *et al.* 2007 X-ray structure analysis of a metalloprotein with enhanced active-site resolution using in situ X-ray absorption near edge structure spectroscopy. *Proc. Natl Acad. Sci. USA* **104**, 6211–6216. (doi:10.1073/pnas.0608411104)
10. Hough MA, Antonyuk SV, Strange RW, Eady RR, Hasnain SS. 2008 Crystallography with online optical and X-ray absorption spectroscopies demonstrates an ordered mechanism in copper nitrite reductase. *J. Mol. Biol.* **378**, 353–361. (doi:10.1016/j.jmb.2008.01.097)
11. Horrell S, Antonyuk SV, Eady RR, Hasnain SS, Hough MA, Strange RW. 2016 Serial crystallography captures enzyme catalysis in copper nitrite reductase at atomic resolution from one crystal. *IUCrJ* **3**, 271–281. (doi:10.1107/S205225251600823X)
12. Halsted TP *et al.* 2018 An unprecedented dioxygen species revealed by serial femtosecond rotation crystallography in copper nitrite reductase. *IUCrJ* **5**, 22–31. (doi:10.1107/S2052252517016128)
13. Suga M *et al.* 2015 Native structure of photosystem II at 1.95 Å resolution viewed by femtosecond X-ray pulses. *Nature* **517**, 99–103. (doi:10.1038/nature13991)
14. Suga M *et al.* 2017 Light-induced structural changes and the site of O=O bond formation in PSII caught by XFEL. *Nature* **543**, 131–135. (doi:10.1038/nature21400)
15. Phillips JC, Wlodawer A, Yevitz MM, Hodgson KO. 1976 Applications of synchrotron radiation to protein crystallography: preliminary results. *Proc. Natl Acad. Sci. USA* **73**, 128–132. (doi:10.1073/pnas.73.1.128)
16. Lemonnier M, Fourme R, Rousseaux F, Kahn R. 1978 X-ray curved-crystal monochromator system at the storage ring DCI. *Nucl. Instrum. Methods* **152**, 173–177. (doi:10.1016/0029-554X(78)90259-8)
17. Wilson KS *et al.* 1983 Macromolecular crystallography with synchrotron radiation. II. Results. *J. Appl. Crystallogr.* **16**, 28–41. (doi:10.1107/S0021889883009917)
18. Fermi G, Perutz MF, Shaanan B, Fourme R. 1984 The crystal structure of human deoxyhaemoglobin at 1.74 Å resolution. *J. Mol. Biol.* **175**, 159–174. (doi:10.1016/0022-2836(84)90472-8)
19. Greaves GN, Durham PJ, Diakun G, Quinn P. 1981 Near-edge X-ray absorption spectra for metallic Cu and Mn. *Nature* **294**, 139–142. (doi:10.1038/294139a0)
20. Mahendrasingam A, Forsyth VT, Hussain R, Greenall RJ, Pigram WJ, Fuller W. 1986 Time-resolved X-ray diffraction studies of the B in equilibrium D structural transition in the DNA double helix. *Science* **233**, 195–197. (doi:10.1126/science.3726529)
21. Acharya R, Fry E, Stuart D, Fox G, Rowlands D, Brown F. 1989 The three-dimensional structure of foot-and-mouth disease virus at 2.9 Å resolution. *Nature* **337**, 709–716. (doi:10.1038/337709a0)
22. Abrahams JP, Leslie AGW, Lutter R, Walker JE. 1994 Structure at 2.8 Å resolution of F1-ATPase from bovine heart mitochondria. *Nature* **370**, 621–628. (doi:10.1038/370621a0)
23. Hasnain SS, Helliwell JR, Kamitsubo H. 1999 Synchrotron radiation and structural biology. *J. Synchrotron Radiat.* **6**, 809–811. (doi:10.1107/S0909049599007232)
24. Garratt RC, Evans RW, Hasnain SS, Lindley PF. 1986 An extended-X-ray-absorption-fine-structure investigation of diferric transferrins and their iron-binding fragments. *Biochem. J.* **233**, 479–484. (doi:10.1042/bj2330479)
25. Bailey S *et al.* 1988 Molecular structure of serum transferrin at 3.3-Å resolution. *Biochemistry* **27**, 5804–5812. (doi:10.1021/bi00415a061)

26. Grossmann GJ, Neu M, Schwab FJ, Evans RW, Townes-Andrews E, Lindley PF, Appel H, Thies WG, Hasnain SS. 1992 X-ray solution scattering reveals conformational changes upon iron uptake in lactoferrin, serum and ovo-transferrins. *J. Mol. Biol.* **225**, 811–819. (doi:10.1016/0022-2836(92)90402-6)
27. Garner CD, Hasnain SS. 1981 DL/SCI/R17.
28. Saibil HR, Grunewald K, Stuart DI. 2015 A national facility for biological cryo-electron microscopy. *Acta Crystallogr. D Biol. Crystallogr.* **71**, 127–135. (doi:10.1107/S1399004714025280)
29. Muench SP, Antonyuk SV, Hasnain SS. 2019 The expanding toolkit for structural biology: synchrotrons, X-ray lasers and cryoEM. *IUCrJ* **6**, 167–177. (doi:10.1107/S2052252519002422)
30. Catlow CRA, Chadwick AV, Greaves GN, Moroney LM. 1984 Direct observations of the dopant environment in fluorites using EXAFS. *Nature* **312**, 601–604. (doi:10.1038/312601a0)
31. Catlow CRA, Chadwick AV, Greaves GN, Moroney LM. 1986 EXAFS study of yttria-stabilized zirconia. *J. Am. Ceram. Soc.* **69**, 272–277. (doi:10.1111/j.1151-2916.1986.tb07425.x)
32. Cernik RJ, Cheetham AK, Prout CK, Watkin DJ, Wilkinson AP, Willis BTM. 1991 The structure of cimetidine (C10H16N6S) solved from synchrotron-radiation X-ray powder diffraction data. *J. Appl. Crystallogr.* **24**, 222–226. (doi:10.1107/S0021889890013486)
33. Cernik RJ. 2016 The development of synchrotron X-ray diffraction at Daresbury Laboratory and its legacy for materials imaging. *J. Non. Cryst. Solids* **451**, 2–9. (doi:10.1016/j.jnoncrysol.2016.05.014)
34. Dooryhee E, Catlow CRA, Couves JW, Maddox PJ, Thomas JM, Greaves GN, Steel AT, Townsend RP. 1991 A study of cation environment and movement during dehydration and reduction of nickel-exchanged zeolite Y by X-ray absorption and diffraction. *J. Phys. Chem.* **95**, 4514–4521. (doi:10.1021/j100164a062)
35. Couves JW, Thomas JM, Waller D, Jones RH, Dent AJ, Derbyshire GE, Greaves GN. 1991 Tracing the conversion of aurichalcite to a copper catalyst by combined X-ray absorption and diffraction. *Nature* **354**, 465–468. (doi:10.1038/354465a0)
36. Lewis DW, Sankar G, Wyles JK, Thomas JM, Catlow CRA, Willock DJ. 1997 Synthesis of a small-pore microporous material using a computationally designed template. *Angew. Chem. Int. Ed. Engl.* **36**, 2675–2677. (doi:10.1002/anie.199726751)
37. Thomas JM, Catlow CRA, Sankar G. 2002 Determining the structure of active sites, transition states and intermediates in heterogeneously catalysed reactions. *Chem. Commun.* **24**, 2921–2925 (doi:10.1039/b210679p)
38. Gibson EK *et al.* 2015 Restructuring of AuPd nanoparticles studied by a combined XAFS/DRIFTS approach. *Chem. Mater.* **27**, 3714–3720. (doi:10.1021/acs.chemmater.5b00866)
39. Malta G *et al.* 2017 Identification of single-site gold catalysis in acetylene hydrochlorination. *Science* **355**, 1399–1402. (doi:10.1126/science.aal3439)
40. Senecal P *et al.* 2017 Real-time scattering-contrast imaging of a supported cobalt-based catalyst body during activation and Fischer–Tropsch synthesis revealing spatial dependence of particle size and phase on catalytic properties. *ACS Catal.* **7**, 2284–2293. (doi:10.1021/acscatal.6b03145)
41. Meirer F, Weckhuysen B. 2018 Spatial and temporal exploration of heterogeneous catalysts with synchrotron radiation. *Nat. Rev. Mater.* **3**, 324–340. (doi:10.1038/s41578-018-0044-5)
42. Nango E *et al.* 2016 A three-dimensional movie of structural changes in bacteriorhodopsin. *Science* **354**, 1552–1557. (doi:10.1126/science.aah3497)
43. Nogly P *et al.* 2018 Retinal isomerization in bacteriorhodopsin captured by a femtosecond X-ray laser. *Science* **361**, 145.