

Online Research @ Cardiff

This is an Open Access document downloaded from ORCA, Cardiff University's institutional repository: <http://orca.cf.ac.uk/97660/>

This is the author's version of a work that was submitted to / accepted for publication.

Citation for final published version:

Bassetto, Marcella, Leyssen, Pieter, Neyts, Johan, Yerukhimovich, Mark M., Frick, David N. and Brancale, Andrea 2017. Shape-based virtual screening, synthesis and evaluation of novel pyrrolone derivatives as antiviral agents against HCV. *Bioorganic & Medicinal Chemistry Letters* 27 (4) , pp. 936-940. 10.1016/j.bmcl.2016.12.087 file

Publishers page: <http://dx.doi.org/10.1016/j.bmcl.2016.12.087>
<<http://dx.doi.org/10.1016/j.bmcl.2016.12.087>>

Please note:

Changes made as a result of publishing processes such as copy-editing, formatting and page numbers may not be reflected in this version. For the definitive version of this publication, please refer to the published source. You are advised to consult the publisher's version if you wish to cite this paper.

This version is being made available in accordance with publisher policies. See <http://orca.cf.ac.uk/policies.html> for usage policies. Copyright and moral rights for publications made available in ORCA are retained by the copyright holders.



Shape-based virtual screening, synthesis and evaluation of novel pyrrolone derivatives as antiviral agents against HCV

Marcella Bassetto^a ¹, Pieter Leyssen^b, Johan Neyts^b, Mark M. Yerukhimovich^c, David N. Frick ^c, Andrea Brancale^a

^aCardiff School of Pharmacy and Pharmaceutical Sciences, Cardiff, King Edward VII Avenue, Cardiff CF103NB, UK

^bRega Institute for Medical Research, University of Leuven, Belgium

^cDepartment of Chemistry & Biochemistry, University of Wisconsin- Milwaukee, Milwaukee, Wisconsin 53211, United States

Hepatitis C virus (HCV) is a major cause of chronic liver disease and affected individuals are at high risk of developing hepatic steatosis, fibrosis, cirrhosis and hepatocellular carcinoma.¹ There is no vaccine available to prevent the infection and such vaccine is not soon expected. The standard of care used to be a combination of pegylated interferon (pegIFN) and ribavirin, a therapy not specific for HCV and efficient in 50% of treated patients, with many associated side effects.² However, new interferon-free combinations of direct acting antivirals (DAAs) have revolutionized HCV prognosis and treatment.

HCV ~9,000 nt single-stranded, positive-sense RNA genome encodes six non-structural proteins (NS2, NS3, NS4A, NS4B, NS5A and NS5B) essential for virus replication.³ Oral combination treatment regimens with NS5B polymerase, NS3 protease and/or NS5A inhibitors have become the standard of HCV care,⁴⁻⁸ but they are associated with the development of resistant mutants⁹⁻¹³ and high costs.¹⁴⁻¹⁵ Due to these limitations, the development of new therapeutics is still needed.

HCV NS3 helicase is one of the most underexploited targets in the search for anti-HCV compounds, with very few specific inhibitors reported so far, none of which reached the clinical stage.¹⁶ The protein main function, essential for viral replication, is the ATP-dependent unwinding of double-stranded RNA sequences.¹⁷⁻²⁰ Due to its fundamental role in HCV replication and the lack of inhibitors under development, this enzyme was chosen as target for the computer-aided identification and synthesis of viral replication inhibitors.

A successful computer-based approach for the identification of new biologically active compounds is represented by shape-complementarity search methods.²¹ Common docking-based techniques usually estimate the ligand binding energy by a scoring procedure that emphasises electrostatics rather than shape. In ligand-based methods instead, the shapes of known ligands are compared with unknown ones, while the chemical properties of known molecules can be included in the comparison as pharmacophoric or electrostatic models; for these approaches a higher accuracy in finding active compounds has been demonstrated.²² Ligand-based methods can evaluate two-dimensional similarities, such as fingerprints,²³ or three-dimensional similarities, focusing on the molecule occupational volume. While 2D comparison methods tend to find hits with the same functional groups of the query molecule,²⁴ 3D search programs such as ROCS focus on the occupational volume associated with each

¹ Corresponding author

E-mail address: bassettom@cardiff.ac.uk (Dr. M. Bassetto)

molecule,²⁵ thus allowing the identification of novel lead structures with a higher level of scaffold diversity.

While previous successful efforts in our research group focused on protein-based approaches for the identification of new antivirals,²⁶⁻²⁷ in order to explore the potential of a shape-comparison method in the search for novel anti-HCV agents, this technique was chosen to screen a library of commercially available compounds. The structures of the dye soluble blue HT **1** and a series of reported compounds derived from **1** were used as starting point,²⁸ being **1** the only known helicase inhibitor for which a crystal structure in complex with the enzyme is available (PDB ID 1ZJO, **Figure 1**).²⁹

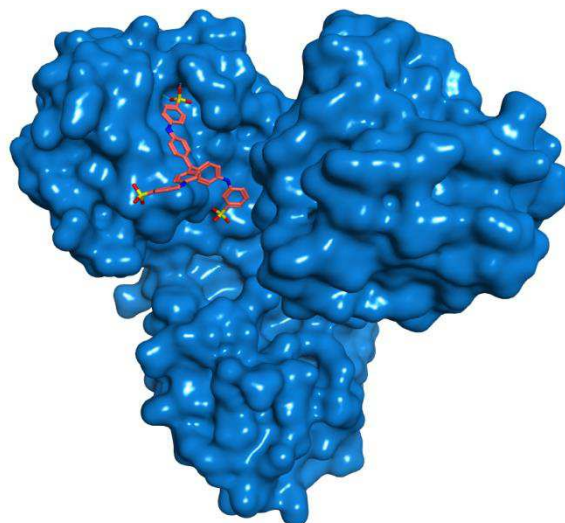


Figure 1: Crystal structure of **1** in complex with HCV NS3 helicase (PDB ID 1ZJO).

While **1** is toxic to cells and its effect against the viral replication cannot be evaluated in cell-based assays, several reported HCV NS3 helicase inhibitors have recently been tested in both an optimised enzymatic assay and a replicon cell-based assay:¹⁶ among them, one structural derivative of **1**, triphenylmethane analogue **2** (**Figure 2**),²⁸ has been confirmed to inhibit both the enzyme (IC_{50} 17 +/- 7 μ M) and the viral replication (~80% at 10 μ M) without relevant cytotoxicity (~75% cell viability at 10 μ M).¹⁶

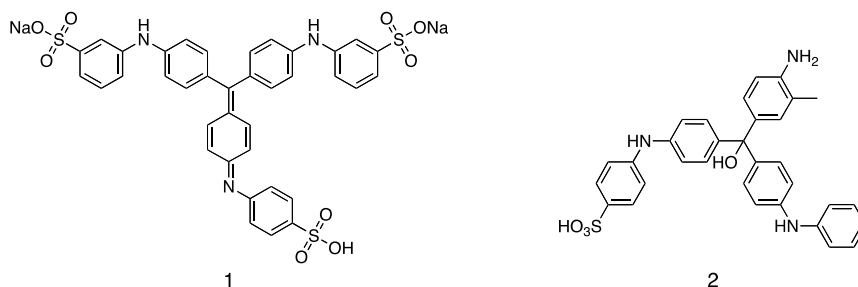


Figure 2: Chemical structures of **1** and **2**.

Due to its activity profile against the viral replication in cells, **2** was chosen as query for a shape-based screening of the SPECS library³⁰ using ROCS 3D.²⁵ In order to evaluate the shape complementarity between the target query and screened compounds, shape-density overlapping volumes of

superimposed molecules are considered by the program and scored according to a Tanimoto-like overlapping value. The conformational state of the query molecule plays an essential role for hit selection, since all analysed structures will be scored based on their overlapping with the target conformation. Along with the conformation selection for the query, the evaluation of the different three-dimensional states of the screened structures becomes crucial. First, a conformational search with MOE2015.10 was performed for the creation of a conformational database for the screening compounds.³¹ A conformational analysis was also performed on **2**, obtaining multiple low-energy three-dimensional states. The conformational results found for **2** were then compared with the conformation of **1** in the 1ZJO crystal structure using MOE2015.10 Flexible Alignment tool.³² The low-energy conformations found for **2** were rigidly aligned to **1**, and the one corresponding to the best alignment was chosen as query (**Figure 3**).

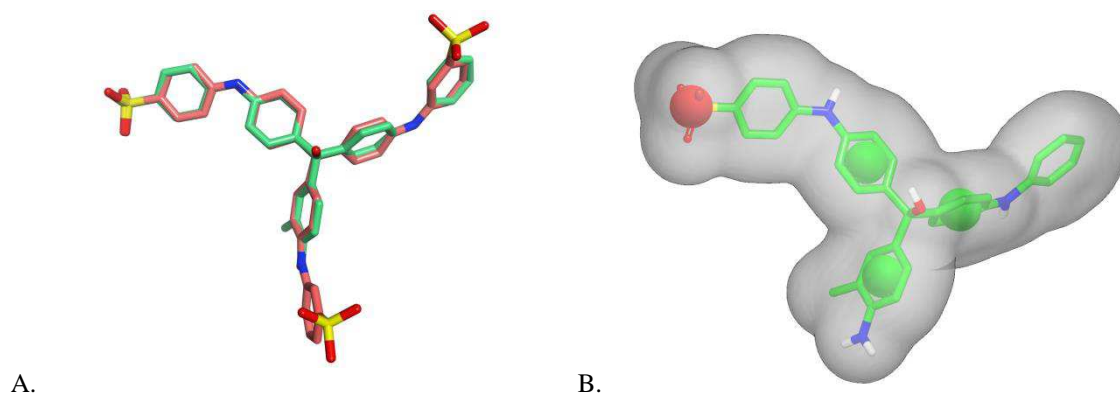


Figure 3: **A.** Best conformational superimposition between **1**, in pink, and **2**, in green; **B.** Query generated from **2** in vROCS3.1.2: the overall molecular shape is represented as grey surface, while the chemical features considered are an H-bond acceptor (red sphere) and three hydrophobic/aromatic centres (green spheres).

The conformational database obtained for the SPECS compounds was analysed against the selected conformation of **2** with vROCS3.1.2,²⁵ considering both shape and electrostatics complementarity. All screened conformations were ranked based on their similarity score with the given query. A selection was made among the screened molecules to prioritise those with the highest similarity indexes (Combo-shape Tanimoto score) and the biggest number of conformations matching the query. Six compounds were finally selected, purchased and evaluated against the viral replication in a cell-based assay (Supporting Information). Among them, **3a** (**Figure 4**) was found to inhibit the viral replication with an EC_{50} value of 38 μ M (**Table 1**). Its scaffold was chosen for the preparation of a small series of novel derivatives, with which to confirm the biological activity found.

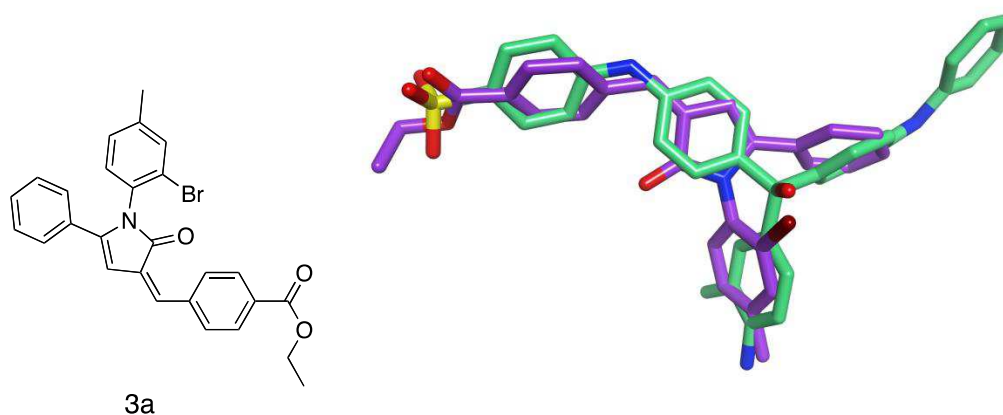
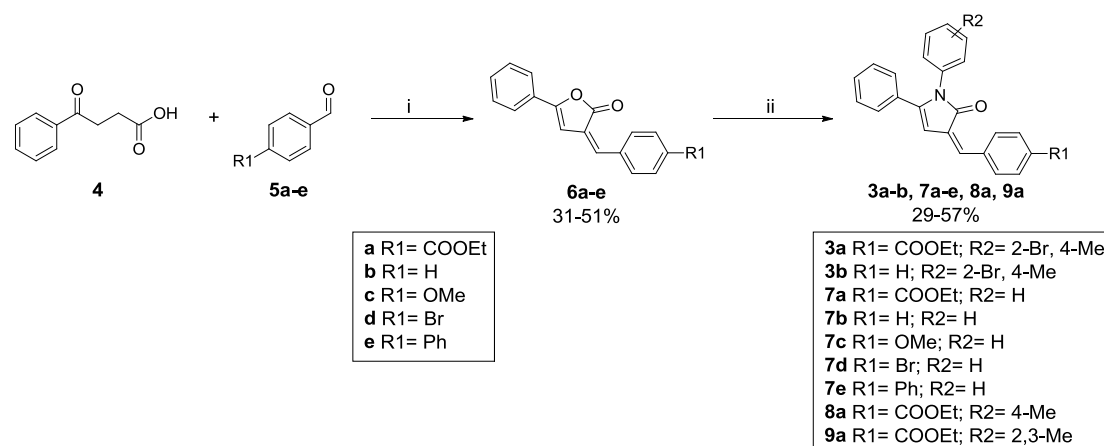


Figure 4: Chemical structure and best alignment found for **3a** (aligned in purple against **2**, in green).

3a is characterised by a central pyrrolone nucleus, substituted in position 1 and 5 with two aromatic rings, and linked in position 3 with a phenylidene system. A preliminary series of triphenyl-pyrrolone derivatives was designed by varying the aromatic substituents on the 1-phenyl and 3-phenylidene rings. This small series of compounds was envisaged to confirm the antiviral potential originally found for **3a** and to explore the effect of the aromatic substituents in the two substituted aromatic rings. In particular, removal of the 4-ethyl carboxylate group on the 1-phenyl ring of **3a** was envisaged to confirm its relevance for antiviral activity (**3b**, **7b**), along with its replacement with different electron withdrawing (4-Br in **7d**) and electron donating (4-OMe in **7c** and 4-Phenyl in **7e**) groups. Preliminary modifications designed for the substituted 3-phenylidene ring of **3a** included the removal of both substituents (**7a-e**, **8a**) or of the original 2-Br function only (**8a**), and their replacement with a 2,3-dimethyl substitution (**9a**). All desired compounds were prepared according to an optimised two-step synthetic pathway (**Scheme 1**).



Scheme 1: Reagents and conditions: i. AcONa, Ac₂O, 95 °C, 2h; ii. Substituted aniline, AcOH, rt, 4h, reflux, 22h.

The strategy carried out began with the condensation of 3-benzoyl propanoic acid **4** with different *para* substituted benzaldehydes **5a-e**, with the formation of butenolide intermediates **6a-e** through furanone ring closure, followed by phenyl-amine displacement of the cyclic ester with the formation of the

pyrrolone system in final products **3a-b**, **7a-e**, **8a** and **9**. According to literature procedures,³³⁻³⁴ furanone intermediates **6a-e** were obtained by heating the starting materials at 95 °C in acetic anhydride in the presence of sodium acetate. The formation of the central butenolide ring is believed to follow a Perkin condensation between β -benzoylpropionic acid and the substituted aryl aldehyde,³⁵ which are thought to form an aldol-condensation product that subsequently undergoes internal cyclisation with the formation of the furanone ring. In these conditions only one species, corresponding to the *cis* isomer, is formed.³³ The desired final products were subsequently obtained by treating intermediates **6a-e** with differently substituted anilines, stirring the reaction mixture in glacial acetic acid first at room temperature for 4 hours, and then heating to reflux for 22 hours. The furanone ring is believed to open as a result of the ammonolysis of the starting compound by the aromatic amine, followed by cyclisation and dehydration events to give the final pyrrolone system.³⁶ All newly synthesised pyrrolones were evaluated for their potential antiviral activity in the HCV replicon and cytostatic assay (Table 1).³⁷ The HCV protease inhibitor telaprevir (VX-950) was included as positive control.

Table 1: Antiviral effect of the test compounds on hepatitis C virus replication in the Huh5-2 replicon system and inhibition of NS3 helicase unwinding activity.

Compound	EC ₅₀ (μM) ^{a,d,37}	EC ₉₀ (μM) ^{b,d}	CC ₅₀ (μM) ^{c,d}	SI ^e	Unwinding IC ₅₀ (μM) ^{f,16}
3a (SPECS)	38	149	>204	>4	n.d.
3a (synthesised)	20.3±1.1	61.7±2.7	>205	>10	>1000
3b	4.1±2.5	12.4±6.3	>240	>58	>1000
7a	27.2±7.6	100	112	4	>1000
7b	4.7±0.5	14±4.2	30.3±5.9	6.4	>1000
7c	5.9±1.8	21.6±1.8	97.2±8.7	18	438
7d	4.3±1.5	11.6±0.8	25.7±6.4	6	>1000
7e	16.1±2.7	61.7	156±63.3	9.7	>1000
8a	22.5±0.9	81.1±13.1	>244	>11	>1000
9a	43.1±19.9	-	90.8±26.8	2.1	>1000
(VX-950)	0.8±0.2	-	47	58.8	n.d.
Primuline	-	-	-	-	10±2
Aurintricarboxylic Acid	-	-	-	-	0.3±0.1

^a EC₅₀ = 50% effective concentration (concentration at which 50% inhibition of virus replication is observed).

^b EC₉₀ = 90% effective concentration (concentration at which 90% inhibition of virus replication is observed).

^c CC₅₀ = 50% cytostatic/cytotoxic concentration (concentration at which 50% adverse effect is observed on the host cell).

^d The EC₅₀, EC₉₀ and CC₅₀ values are the mean of at least 2 independent experiments, with standard deviations of ±10% of the value quoted unless otherwise stated (mean value ± standard deviations).

^e SI = the ratio of CC₅₀ to EC₅₀.

^f Concentration needed to reduce rates of helicase-catalysed DNA unwinding by 50%

n.d. Not determined

Re-synthesised **3a** confirmed its activity profile, with EC₅₀, EC₉₀ and CC₅₀ values close to the ones previously found for the batch acquired from SPECS. The presence of an ethyl ester function in the 3-phenylidene ring does not appear to be essential for activity retention, since **3b**, where this group is removed, is the most active analogue found so far. A similar profile can be identified for derivatives **7b-e**, where the 1-phenyl ring is unsubstituted and the ethyl ester group is replaced respectively with 4-H, 4-Me, 4-Br and 4-phenyl groups. For analogs **7b-d** however an increased cytotoxic effect can also be observed. Activity retention found for **7a** and **8a** seems to indicate that also the presence of the 2-bromo, 4-methyl substituent in the 1-phenyl ring is not essential for antiviral activity, while the insertion of a methyl group in position 3 of this ring and the replacement of the original 2-bromo substituent with an extra methyl group are detrimental, as can be observed for **9a**.

All the final compounds synthesised were also tested for their potential interference with the HCV NS3 helicase activity (**Table 1**). Despite the fact that **7c** shows some interference with the helicase unwinding activity at high concentrations and **3b** was found to competitively inhibit RNA binding with an IC₅₀ of 620 µM (data not shown), a trend for this effect cannot be identified, and it does not seem to correlate with any specific structural feature. Moreover, the IC₅₀ values observed are dramatically higher than the range of activities found in the HCV replicon assay, and a correlation between the two sets of data cannot be found. This evidence suggests that the antiviral effect of the pyrrolone structures presented in this study might be due to a different target, viral or cellular, other than the HCV NS3 helicase. Additional studies aimed to further explore the structure-activity relationships associated to the pyrrolone scaffold and to determine the biological target of the newly synthesised compounds are ongoing and will be reported in due course. In particular, ongoing synthetic efforts are directed to increase the water solubility of the novel antiviral scaffold, to further explore the effect of different aromatic substitutions on the 1-phenyl, 3-phenylidene and 5-phenyl rings, to evaluate the importance of the rigidity of the molecule with the preparation of unsaturated analogues, and to replace the central pyrrolone nucleus with different heterocyclic systems.

In conclusion, starting from a ligand-based *in silico* approach, a substituted pyrrolone scaffold was identified as hit for the inhibition of HCV replication in the subgenomic replicon assay. Its structure was the starting point for the design and synthesis of a first series of new analogs, with which antiviral potential was confirmed and improved. Different novel derivatives with EC₅₀ values in the low micromolar range were found. Preliminary structure-activity relationships could be identified for the antiviral effect, while the first enzymatic evaluations performed seem to suggest that the antiviral potential associated with the newly prepared compounds is not directly correlated with interference with the NS3 helicase, but other viral or cellular targets might be involved. Further exploration of modified novel derivatives and understanding the biological target of these structures are the current focus of on-going investigations.

References

- 1 World Health Organization. Hepatitis C factsheet 164. July 2015. <http://www.who.int/mediacentre/factsheets/fs164/en/> (accessed November 22, 2016).

- 2 Marrero, J.A. Viral hepatitis and hepatocellular carcinoma, in *Chronic Viral Hepatitis: Diagnosis and Therapeutics* (Shetty, K.; Wu, G.Y., eds) (2nd edn) **2009**, pp.431-447, Springer.
- 3 Suzuki, T.; Ishii, K.; Aizaki, H.; Wakita, T. Hepatitis C viral life cycle. *Adv. Drug Deliv. Rev.* **2007**, *59*, 1200-1212.
- 4 U.S. Food and Drug Administration. www.fda.gov (accessed November 22, 2016).
- 5 FDA approves new treatment for hepatitis C virus. Food and Drug Administration. Nov 22, 2013.
- 6 FDA approves Sovaldi for chronic hepatitis C in FDA New Release, U.S. Food and Drug Administration, 2013-12-06.
- 7 Rai, R.; Deval, J. New opportunities in anti-hepatitis C virus drug discovery: Targeting NS4B. *Antiviral Res.* 2011, *90*, 93-101.
- 8 Anna Suk-Fong Lok. HCV NS5A Inhibitors in Development. *Clinics in Liver Disease*, 2013, *17*, 111-121.
- 9 Gao, M. Antiviral activity and resistance of HCV NS5A replication complex inhibitors. *Curr. Opin. Virol.*, 2013, *3*, 514-520.
- 10 Vermehren, J.; Sarrazin, C. The role of resistance in HCV treatment. *Best Pract. Res. Cl. Ga.* 2012, *26*, 487-503.
- 11 www.gilead.com/~media/Files/pdfs/medicines/liver-disease/harvoni/harvoni_pi (Accessed November 22, 2016).
- 12 Issur, M.; Gotte, M. Resistance patterns associated with HCV NS5A inhibitors provide limited insight into drug binding. *Viruses*, 2014, *6*, 4227-4241.
- 13 Poveda, E.; Wyles, D.; Mena, A.; Pedreira, A.; Castro-Iglesias, A.; Cachay, E. Update on hepatitis C virus resistance to direct-acting antiviral agents. *Antiviral Res.* 2014, *108*, 181-191.
- 14 Gritsenko, D.; Hughes, G. Ledipasvir/Sofosbuvir (Harvoni): Improving Options for Hepatitis C virus infection. *Pharm. Ther.: Drug Forecast.* 2015, *40*, 256-259, 276.
- 15 Hagan, L. M.; Sulkowski, M. S.; Schinazi, R. F. Cost analysis of sofosbuvir/ribavirin versus sofosbuvir/simeprevir for genotype 1 HCV infection ineligible/intolerant individuals. *Hepatology*, 2014, *60*, 37-45.
- 16 Li, K.; Frankowski, K. J.; Belon, C. A.; Neuenswander, B.; Ndjomou, J.; Hanson, A. M.; Shanahan, M. A.; Schoenen, F. J.; Blagg, B. S.; Aube, J.; Frick, D. N. Optimization of Potent Hepatitis C Virus NS3 Helicase Inhibitors Isolated from the Yellow Dyes Thioflavine S and Primuline *J. Med. Chem.* **2012**, *55*, 3319– 3330.
- 17 Lam, A.M.; Frick, D.N. Hepatitis C virus subgenomic replicon requires an active NS3 RNA helicase. *J. Virol.* **2006**, *80*, 404–411.
- 18 Kolykhalov, A.A.; Mihalik, K.; Feinstone, S.M.; Rice, C.M. Hepatitis C virus-encoded enzymatic activities and conserved RNA elements in the 3' nontranslated region are essential for virus replication in vivo. *J. Virol.* **2000**, *74*, 2046–2051.
- 19 Tai, C.L.; Chi, W.K.; Chen, D.S.; Hwang, L.H. The helicase activity associated with hepatitis C virus nonstructural protein 3 (NS3). *J. Virol.* **1996**, *70*, 8477–8484.

- 20** Morris, P.D.; Byrd, A.K.; Tackett, A.J.; Cameron, C.E.; Tanega, P.; Ott, R.; Fanning, E.; Raney, K.D. Hepatitis C virus NS3 and simian virus 40 T antigen helicases displace streptavidin from 5'-biotinylated oligonucleotides but not from 3'-biotinylated oligonucleotides: evidence for directional bias in translocation on single-stranded DNA. *Biochemistry* **2002**, 41, 2372–2378.
- 21** Willock, D.J.; Lewis, D.W.; Catlow, C.R.A.; Hutchings, G.J.; Thomas, J.M. Designing templates for the synthesis of microporous solids using de novo molecular design methods. *J. Mol. Catal. A-Chem.* **1997**, 119, 415-424.
- 22** Hawkins, P.C.D.; Skillman, A.G.; Nicholls, A. Comparison of shape-matching and docking as virtual screening tools. *J. Med. Chem.* **2007**, 50, 74-82.
- 23** Daylight, Version 4.9; Daylight Chemical Information System, Inc.: Aliso Viejo, CA, 2008.
- 24** McGaughey, G.B.; Sheridan, R.P.; Bayly, C.I.; Culberson, J.C.; Kretsoulas, C.; Lindsley, S.; Maiorov, V.; Truchon, J.; Cornell, W.D. Comparison of topological, shape, and docking methods in virtual screening. *J. Chem. Inf. Model.* **2007**, 47, 1504-1519.
- 25** OpenEye Scientific Software, Santa Fe, NM. www.eyesopen.com (accessed November 22, 2016).
- 26** Bassetto, M.; Leyssen, P.; Neyts, J.; Yerukhimovich, M.M.; Frick, D.N.; Brancale, A. Computer-aided identification, synthesis and evaluation of substituted thienopyrimidines as novel inhibitors of HCV replication. *Eur. J. Med. Chem.* **2016**, 123, 31-47.
- 27** Bassetto, M.; Leyssen, P.; Neyts, J.; Yerukhimovich, M.M.; Frick, D.N.; Cortney-Smith, M.; Brancale, A. In silico identification, design and synthesis of novel piperazine-based antiviral agents targeting the hepatitis C virus helicase. *Eur. J. Med. Chem.* **2016**, 125, 1115-1131.
- 28** Chen, C.S.; Chiou, C.T.; Chen, G.S.; Chen, S.C.; Hu, C.Y.; Chi, W.K.; Chu, Y.D.; Hwang, L.H.; Chen, P.J.; Chen, D.S.; Liaw, S.H.; Chern, J.W. Structure-based discovery of triphenylmethane derivatives as inhibitors of hepatitis C virus helicase. *J. Med. Chem.* **2009**, 52, 2716–2723.
- 29** Hu, C.Y.; Chen, S.J.; Liaw, S.H. Rational drug designs based on crystal structures of the Hepatitis C, virus NS3 helicase-inhibitor complexes. *KEK Prog. Rep.* **2003**, 2002-2002, 183.
- 30** Specs. www.specs.net (accessed November 22, 2016).
- 31** Chemical Computing Group, Montreal, Canada. www.chemcomp.com (accessed November 22, 2016).
- 32** Chan, S.L.; Labute, P. Training a scoring function for the alignment of small molecules. *J. Chem. Inf. Model.* **2010**, 50, 1724-1735.
- 33** Abbandonato, G.; Signore, G.; Nifosi, R.; Voliani, V.; Bizzarri, R.; Beltram, F. Cis–trans photoisomerization properties of GFP chromophore analogs. *Eur. Biophys. J.* 2011, 40, 1205–1214.
- 34** Bourotte, M.; Schmitt, M.; Follenius-Wund, A.; Pigault, C.; Haiech, J.; Bourguignon, J.J. Fluorophores related to the green fluorescent protein. *Tetrahedron Lett.* 2004, 45, 6343–6348.
- 35** Deo, S.; Inam, F.; Mahashabde, R.P.; Jadhav, A.N. Synthesis of 1-Phenyl naphthalene and pericarbonyl lignans. *Asian J. Chem.* 2010, 22, 3362-3368.
- 36** Wang, H. Et al. Conjugated polymers based on a new building block: dithienophthalimide. *Macromolecules*, **2011**, 44, 4213-4221.

37 Bartensclager, R. Hepatitis C virus replicons: potential role for drug development, *Nature Rev. Drug Disc.* **2002**, 1, 911-916.